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(54) Title: PROSTATE CANCER-RELEASED GENE 3 (PG-3) AND BIALLELIC MARKERS THEREOF

(57) Abstract: The invention concerns the genomic sequence and cDNA sequences of the PG-3 gene. The invention also concerns biallelic markers of the PG-3 gene. The invention also concerns polypeptides encoded by the PG-3 gene. The invention also deals with antibodies directed specifically against such polypeptides that are useful as diagnostic reagents.

PROSTATE CANCER-RELATED GENE 3 (PG3) AND BIALLELIC MARKERS THEREOF**FIELD OF THE INVENTION**

The present invention is directed to polynucleotides encoding a PG-3 polypeptide as well as
5 the regulatory regions located at the 5'- and 3'-ends of said coding region. The invention also relates
to polypeptides encoded by the PG-3 gene. The invention also relates to antibodies directed
specifically against such polypeptides that are useful as diagnostic reagents. The invention further
encompasses biallelic markers of the PG-3 gene useful in genetic analysis.

BACKGROUND OF THE INVENTION

10 Cancer is one of the leading causes of death in industrialized countries. This makes cancer
a serious burden in terms of public health, especially in view of the aging of the population. Indeed,
over the next 25 years there will be a dramatic increase in the number of people developing cancer.
Globally, 10 million new cancer patients are diagnosed each year and there will be 20 million new
cancer diagnoses by the year 2020.

15 In spite of a large number of available therapeutic techniques including but not limited to
surgery, chemotherapy, radiotherapy, bone marrow transplantation, and in spite of encouraging
results obtained with experimental protocols in immunotherapy or gene therapy, the overall survival
rate of cancer patients does not reach 50% after 5 years. Therefore, there is a strong need for both a
reliable diagnostic procedure which would enable early-stage cancer prognosis, and for preventive
20 and curative treatments of the disease.

A cancer is a clonal proliferation of cells produced as a consequence of cumulative genetic
damage that finally results in unrestrained cell growth, tissue invasion and metastasis (cell
transformation). Regardless of the type of cancer, transformed cells carry damaged DNA as gross
chromosomal translocations or, more subtly, as DNA amplification, rearrangement or even point
25 mutations.

Cancer is caused by the dysregulation of the expression of certain genes. The development
of a tumor requires an important succession of steps. Each of these comprises the dysregulation of
a gene either involved in cell cycle activity or in genomic stability and the emergence of an
abnormal mutated clone which overwhelms the other normal cell types because of a proliferative
30 advantage. Cancer indeed happens because of a combination of two mechanisms. Some mutations
enhance cell proliferation, increasing the target population of cells for the next mutation. Other
mutations affect the stability of the entire genome, increasing the overall mutation rate, as in the
case of mismatch repair proteins (reviewed in Arnheim N & Shibata D, 1997).

Recent studies have identified three groups of genes which are frequently mutated in
35 cancer. The first two groups are involved in cell cycle activity, which is a mechanism that drives
normal cell proliferation and ensures the normal development and homeostasis of the organism.
Conversely, many of the properties of cancer cells - uncontrolled proliferation, increased mutation

rate, abnormal translocations and gene amplifications - can be attributed directly to perturbations of the normal regulation or progression of the cycle.

The first group of genes, called oncogenes, are genes whose products activate cell proliferation. The normal non-mutant versions are called protooncogenes. The mutated forms are
5 excessively or inappropriately active in promoting cell proliferation and act in the cell in a dominant way such that a single mutant allele is enough to affect the cell phenotype. Activated oncogenes are rarely transmitted as germline mutations since they are probably be lethal when expressed in all the cells in the organism. Therefore oncogenes can only be investigated in tumor tissues. Oncogenes and protooncogenes can be classified into several different categories according to their function.
10 This classification includes genes that code for proteins involved in signal transduction such as: growth factors (*i.e.*, *sis*, *int-2*); receptor and non-receptor protein-tyrosine kinases (*i.e.*, *erbB*, *src*, *bcr-abl*, *met*, *trk*); membrane-associated G proteins (*i.e.*, *ras*); cytoplasmic protein kinases (*i.e.*, mitogen-activated protein kinase -*MAPK*- family, *raf*, *mos*, *pak*), or nuclear transcription factors (*i.e.*, *myc*, *myb*, *fos*, *jun*, *rel*) (for review see Hunter T, 1991 ; Fanger GR *et al.*, 1997 ; Weiss FU *et al.*, 1997).
15

The second group of genes which are frequently mutated in cancer, called tumor suppressor genes, are genes whose products inhibit cell growth. Mutant versions in cancer cells have lost their normal function, and act in the cell in a recessive way such that both copies of the gene must be inactivated in order to change the cell phenotype. Most importantly, the tumor phenotype can be
20 rescued by the wild type allele, as shown by cell fusion experiments first described by Harris and colleagues (Harris H *et al.*, 1969). Germline mutations of tumor suppressor genes are transmitted and thus studied in both constitutional and tumor DNA from familial or sporadic cases. The current family of tumor suppressors includes DNA-binding transcription factors (*i.e.*, *p53*, *WT1*), transcription regulators (*i.e.*, *RB*, *APC*, and *BRCA1*), and protein kinase inhibitors (*i.e.*, *p16*), among
25 others (for review, see Haber D & Harlow E, 1997).

The third group of genes which are frequently mutated in cancer, called mutator genes, are responsible for maintaining genome integrity and/or low mutation rates. Loss of function of both alleles increases cell mutation rates, and as a consequence, proto-oncogenes and tumor suppressor genes are mutated. Mutator genes can also be classified as tumor suppressor genes, except for the
30 fact that tumorigenesis caused by this class of genes cannot be suppressed simply by restoration of a wild-type allele, as described above. Genes whose inactivation may lead to a mutator phenotype include mismatch repair genes (*i.e.*, *MLH1*, *MSH2*), DNA helicases (*i.e.*, *BLM*, *WRN*) or other genes involved in DNA repair and genomic stability (*i.e.*, *p53*, possibly *BRCA1* and *BRCA2*) (For review see Haber D & Harlow E, 1997; Fishel & Wilson. 1997 ; Ellis, 1997).

35 The recent development of sophisticated techniques for genetic mapping has resulted in an ever expanding list of genes associated with particular types of human cancers. The human haploid genome contains an estimated 80,000 to 100,000 genes scattered on a 3×10^9 base-long double-

stranded DNA. Each human being is diploid, *i.e.*, possesses two haploid genomes, one from paternal origin, the other from maternal origin. The sequence of a given genetic locus may vary between individuals in a population or between the two copies of the locus on the chromosomes of a single individual. Genetic mapping techniques often exploit these differences, which are called

5 polymorphisms, to map the location of genes associated with human phenotypes.

One mapping technique, called the loss of heterozygosity (LOH) technique, is often employed to detect genes in which a loss of function results in a cancer, such as the tumor suppressor genes described above. Tumor suppressor genes often produce cancer via a two hit mechanism in which a first mutation, such as a point mutation (or a small deletion or insertion)

10 inactivates one allele of the tumor suppressor gene. Often, this first mutation is inherited from generation to generation. A second mutation, often a spontaneous somatic mutation such as a deletion which deletes all or part of the chromosome carrying the other copy of the tumor suppressor gene, results in a cell in which both copies of the tumor suppressor gene are inactive. As a consequence of the deletion in the tumor suppressor gene, one allele is lost for any genetic marker

15 located close to the tumor suppressor gene. Thus, if the patient is heterozygous for a marker, the tumor tissue loses heterozygosity, becoming homozygous or hemizygous. This loss of heterozygosity generally provides strong evidence for the existence of a tumor suppressor gene in the lost region.

LOH has allowed the identification of several chromosomal regions associated with cancer.

20 Indeed, substantial amounts of LOH data support the hypothesis that genes associated with distinct cancer types are located within 8p23 region of the human genome. Several regions of chromosome arm 8p were found to be frequently deleted in a variety of human malignancies including those of the prostate, head and neck, lung and colon. Emi *et al.* demonstrated the involvement of the 8p23.1-8p21.3 region in cases of hepatocellular carcinoma, colorectal cancer, and non-small cell lung

25 cancer (Emi *et al.*, 1992). Yaremko, *et al.*, (1994) showed the existence of two major regions of LOH for chromosome 8 markers in a sample of 87 colorectal carcinomas. The most prominent loss was found for 8p23.1-pter, where 45% of informative cases demonstrated loss of alleles. Scholnick *et al.* (Scholnick *et al.*, 1996 and Sunwoo *et al.*, 1996) demonstrated the existence of three distinct regions of LOH for the markers of chromosome 8 in cases of squamous cell carcinoma of the

30 supraglottic larynx. They showed that the allelic loss of 8p23 marker D8S264 serves as a statistically significant, independent predictor of poor prognosis for patients with supraglottic squamous cell carcinoma. The study of 51 squamous cell carcinomas of the head and neck and 29 oral squamous cell carcinoma cell lines showed a frequent allelic loss and homozygous deletion at 1 or more loci located in the 8p23 region (Ishwad CS *et al.*, 1999). In addition, a high resolution

35 deletion map of 150 squamous cell carcinomas of the larynx and oral cavity showed two distinct classes of deletion for the 8p23 region within the D8S264 to D8S1788 interval (Sunwoo *et al.*, 1999).

In other studies, Nagai *et al.* (1997) demonstrated the highest loss of heterozygosity in the specific region of 8p23 by genome wide scanning of LOH in 120 cases of hepatocellular carcinoma (HCC). Further studies using high-density polymorphic marker analysis identified three minimal deleted areas on chromosome 8p, one of them being a 5 cM area in 8p23, probably indicative of the presence of a tumor suppressor loci for HCC (Pineau P, *et al.*, 1999). Gronwald *et al.* (1997) also demonstrated 8p23-pter loss in renal clear cell carcinomas.

The same region is involved in specific cases of prostate cancer. Matsuyama *et al.* (1994) showed the specific deletion of the 8p23 band in prostate cancer cases, as monitored by FISH with D8S7 probe. They were able to document a substantial number of cases with deletions of 8p23 but retention of the 8p22 marker LPL. Moreover, Ichikawa *et al.* (1996) deduced the existence of a prostate cancer metastasis suppressor gene and localized it to 8p23-q12 by studies of metastasis suppression in highly metastatic rat prostate cells after transfer of human chromosomes. Recently Washburn *et al.* (1997) were able to find substantial numbers of tumors with the allelic loss specific to 8p23 by LOH studies of 31 cases of human prostate cancer. In these samples they were able to define the minimal overlapping region with deletions covering genetic interval D8S262-D8S277. In addition, using PCR analysis of polymorphic microsatellite repeat markers, 29% of 60 prostate tumors showed LOH, at the locus D8S262 of the 8p23 region (Perinchery *et al.*, 1999).

Recent studies have also implicated the 8p23 region in other types of cancers such as fibrous histiocytomas, ovarian adenocarcinomas and gastric cancers. Indeed, comparative genomic hybridization data showed the involvement of the 8p23.1 region in fibrous histiocytomas and detected a minimal amplified region between D8S1819 and D8S550 containing a gene MASL1, the overexpression of which might be oncogenic (Sakabe *et al.*, 1999). LOH was also observed for 27 ovarian adenocarcinomas on 8p. Detailed examination of nine tumours with partial deletions defined three regions of overlap including two in 8p23 (Wright *et al.*, 1998). Comparative genomic hybridization of 58 primary gastric cancers detected gain of the 8p22-23 region in 24% of the tumors and even high-level amplification of the same region in 5% of the tumors. This amplified region was narrowed down to 8p23.1 by reverse-painting FISH to prophase chromosomes (Sakakura *et al.*, 1999).

The present invention relates to the Prostate Cancer Related Gene 3 or PG-3 gene, a gene present in the 8p23 cancer candidate region, as well as diagnostic methods and reagents for detecting alleles of the PG-3 gene which may cause cancer, and therapies for treating cancer.

SUMMARY OF THE INVENTION

The present invention pertains to nucleic acid molecules comprising the genomic sequence and the cDNA sequence of a novel human gene which encodes a PG-3 protein. The PG-3 gene is localized in the 8p23 candidate region shown to be involved in several types of cancer by LOH studies and presents homology with the *BRCA1* gene involved in transcriptional control through modulation of chromatin structure (Bochar *et al.*, 2000), and in which mutations are thought to be

responsible for 45% of inherited breast cancer and more than 80% of inherited breast and ovarian cancer. In addition, *BRCA1* carriers have a 4-fold increased risk of colon cancer, whereas male carriers face a 3-fold increased risk of prostate cancer.

The PG-3 genomic sequence comprises regulatory sequences located upstream (5'-end) and downstream (3'-end) of the transcribed portion of said gene, these regulatory sequences being also part of the invention.

The invention also relates to the cDNA sequence encoding the PG-3 protein, as well as to the corresponding translation product.

Oligonucleotide probes or primers hybridizing specifically with a PG-3 genomic or cDNA sequence are also part of the present invention, as well as DNA amplification and detection methods using said primers and probes.

A further object of the invention relates to recombinant vectors comprising any of the nucleic acid sequences described herein, and in particular to recombinant vectors comprising a PG-3 regulatory sequence or a sequence encoding a PG-3 protein. The present invention also relates to host cells and transgenic non-human animals comprising said nucleic acid sequences or recombinant vectors.

The invention further encompasses biallelic markers of the PG-3 gene useful in genetic analysis.

Finally, the invention is directed to methods for the screening of substances or molecules that inhibit the expression of PG-3, as well as to methods for the screening of substances or molecules that interact with a PG-3 polypeptide or that modulate the activity of a PG-3 polypeptide.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 is a block diagram of an exemplary computer system.

Figure 2 is a flow diagram illustrating one embodiment of a process 200 for comparing a new nucleotide or protein sequence with a database of sequences in order to determine the homology levels between the new sequence and the sequences in the database.

Figure 3 is a flow diagram illustrating one embodiment of a process 250 in a computer for determining whether two sequences are homologous.

Figure 4 is a flow diagram illustrating one embodiment of an identifier process 300 for detecting the presence of a feature in a sequence.

BRIEF DESCRIPTION OF THE SEQUENCES PROVIDED IN THE SEQUENCE LISTING

SEQ ID No 1 is a genomic sequence of PG-3 comprising the 5' regulatory region (upstream untranscribed region), the exons and introns, and the 3' regulatory region (downstream untranscribed region).

SEQ ID No 2 is a cDNA sequence of PG-3.

SEQ ID No 3 is the amino acid sequence encoded by the cDNA of SEQ ID No 2.

SEQ ID No 4 is a primer containing the additional PU 5' sequence further described in Example 2.

SEQ ID No 5 is a primer containing the additional RP 5' sequence further described in Example 2.

- 5 In accordance with the regulations relating to Sequence Listings, the following codes have been used in the Sequence Listing to indicate the locations of biallelic markers within the sequences and to identify each of the alleles present at the polymorphic base. The code "r" in the sequences indicates that one allele of the polymorphic base is a guanine, while the other allele is an adenine. The code "y" in the sequences indicates that one allele of the polymorphic base is a thymine, while
- 10 the other allele is a cytosine. The code "m" in the sequences indicates that one allele of the polymorphic base is an adenine, while the other allele is a cytosine. The code "k" in the sequences indicates that one allele of the polymorphic base is a guanine, while the other allele is a thymine. The code "s" in the sequences indicates that one allele of the polymorphic base is a guanine, while the other allele is a cytosine. The code "w" in the sequences indicates that one allele of the
- 15 polymorphic base is an adenine, while the other allele is a thymine. The nucleotide code of the original allele for each biallelic marker is the following:

	<u>Biallelic marker</u>	<u>Original allele</u>
	5-390-177	C
	5-391-43	G
20	5-392-222	T
	5-392-280	T
	4-59-27	G
	4-58-289	C
	4-54-199	A
25	4-54-180	C
	4-51-312	G
	99-86-266	A
	4-88-107	G
	5-397-141	G
30	5-398-203	C
	99-12738-248	A
	99-109-358	C
	99-12749-175	T
	4-21-154	C
35	4-21-317	G
	4-23-326	G
	99-12753-34	A
	5-364-252	G
	99-12755-280	G
40	99-12755-329	C

	4-87-212	A
	99-12757-318	C
	99-12758-102	G
	99-12758-136	C
5	4-105-98	A
	4-105-86	G
	4-45-49	T
	4-44-277	T
	4-86-60	C
10	4-84-334	G
	99-78-321	T
	99-12767-36	G
	99-12767-143	T
	99-12767-189	T
15	99-12767-380	G
	4-80-328	C
	4-36-384	C
	4-36-264	G
	4-36-261	C
20	4-35-333	A
	4-35-240	G
	4-35-173	T
	4-35-133	C
	99-12771-59	T
25	99-12774-334	A
	99-12776-358	G
	99-12781-113	A
	4-104-298	C
	4-104-254	G
30	4-104-250	C
	4-104-214	A
	99-12818-289	T
	99-24807-271	C
	99-24807-84	G
35	99-12831-157	G
	99-12831-241	C
	99-12832-387	T
	99-12836-30	G
	99-12844-262	C
40	4-24-74	C
	4-24-246	C
	4-24-314	G

8

	4-27-190	A
	5-400-145	G
	5-400-149	G
	5-400-175	T
5	5-400-231	T
	5-400-367	A
	99-12852-110	T
	99-12852-325	A
	4-37-326	A
10	4-37-107	G
	5-270-92	G
	99-12860-47	G
	99-12860-57	T
	5-402-144	C

- 15 In some instances, the polymorphic bases of the biallelic markers alter the identity of an amino acid in the encoded polypeptide. This is indicated in the accompanying Sequence Listing by use of the feature VARIANT, placement of an Xaa at the position of the polymorphic amino acid, and definition of Xaa as the two alternative amino acids. For example if one allele of a biallelic marker is the codon CAC, which encodes histidine, while the other allele of the biallelic marker is
- 20 CAA, which encodes glutamine, the Sequence Listing for the encoded polypeptide will contain an Xaa at the location of the polymorphic amino acid. In this instance, Xaa would be defined as being histidine or glutamine.

DETAILED DESCRIPTION

- The present invention concerns polynucleotides and polypeptides related to the PG-3 gene.
- 25 Oligonucleotide probes and primers hybridizing specifically with a genomic or a cDNA sequence of PG-3 are also part of the invention. A further object of the invention relates to recombinant vectors comprising any of the nucleic acid sequences described in the present invention, and in particular recombinant vectors comprising a regulatory region of PG-3 or a sequence encoding the PG-3 protein, as well as host cells comprising said nucleic acid sequences or recombinant vectors. The
- 30 invention also encompasses methods of screening for molecules which inhibit the expression of the PG-3 gene or which modulate the activity of the PG-3 protein. The invention also relates to antibodies directed specifically against such polypeptides that are useful as diagnostic reagents.

- The invention also concerns PG-3-related biallelic markers which can be used in any method of genetic analysis including linkage studies in families, linkage disequilibrium studies in
- 35 populations and association studies of case-control populations. An important aspect of the present invention is that biallelic markers allow association studies to be performed to identify genes involved in complex traits. These biallelic markers may lead to allelic variants of the PG-3 protein.

Definitions

typically comprises about 50%, preferably 60 to 90% weight/weight of a polypeptide or polynucleotide sample, respectively, more usually about 95%, and preferably is over about 99% pure. Polypeptide and polynucleotide purity, or homogeneity, is indicated by a number of means well known in the art, such as agarose or polyacrylamide gel electrophoresis of a sample, followed
5 by visualizing a single band upon staining the gel. For certain purposes higher resolution can be provided by using HPLC or other means well known in the art. As an alternative embodiment, purification of the polypeptides and polynucleotides of the present invention may be expressed as "at least" a percent purity relative to heterologous polypeptides and polynucleotides (DNA, RNA or both). As a preferred embodiment, the polypeptides and polynucleotides of the present invention are at least;
10 10%, 20%, 30%, 40%, 50%, 60%, 70%, 80%, 90%, 95%, 96%, 96%, 98%, 99%, or 100% pure relative to heterologous polypeptides and polynucleotides, respectively. As a further preferred embodiment the polypeptides and polynucleotides have a purity ranging from any number, to the thousandth position, between 90% and 100% (e.g., a polypeptide or polynucleotide at least 99.995% pure) relative to either heterologous polypeptides or polynucleotides, respectively, or as a
15 weight/weight ratio relative to all compounds and molecules other than those existing in the carrier. Each number representing a percent purity, to the thousandth position, may be claimed as individual species of purity.

The term "polypeptide" refers to a polymer of amino acids without regard to the length of the polymer; thus, peptides, oligopeptides, and proteins are included within the definition of
20 polypeptide. This term also does not specify or exclude post-expression modifications of polypeptides, for example, polypeptides which include the covalent attachment of glycosyl groups, acetyl groups, phosphate groups, lipid groups and the like are expressly encompassed by the term polypeptide. Also included within the definition are polypeptides which contain one or more analogs of an amino acid (including, for example, non-naturally occurring amino acids, amino acids
25 which only occur naturally in an unrelated biological system, modified amino acids from mammalian systems etc.), polypeptides with substituted linkages, as well as other modifications known in the art, both naturally occurring and non-naturally occurring.

The term "recombinant polypeptide" is used herein to refer to polypeptides that have been artificially designed and which comprise at least two polypeptide sequences that are not found as
30 contiguous polypeptide sequences in their initial natural environment, or to refer to polypeptides which have been expressed from a recombinant polynucleotide.

As used herein, the term "non-human animal" refers to any non-human vertebrate, birds and more usually mammals, preferably primates, farm animals such as swine, goats, sheep, donkeys, and horses, rabbits or rodents, more preferably rats or mice. As used herein, the term "animal" is
35 used to refer to any vertebrate, preferable a mammal. Both the terms "animal" and "mammal" expressly embrace human subjects unless preceded with the term "non-human".

As used herein, the term “antibody” refers to a polypeptide or group of polypeptides which are comprised of at least one binding domain, where an antibody binding domain is formed from the folding of variable domains of an antibody molecule to form three-dimensional binding spaces with an internal surface shape and charge distribution complementary to the features of an antigenic determinant of an antigen, which allows an immunological reaction with the antigen. Antibodies include recombinant proteins comprising the binding domains, as well as fragments, including Fab, Fab', F(ab)₂, and F(ab')₂ fragments.

As used herein, an “antigenic determinant” is the portion of an antigen molecule, in this case a PG-3 polypeptide, that determines the specificity of the antigen-antibody reaction. An “epitope” refers to an antigenic determinant of a polypeptide. An epitope can comprise as few as 3 amino acids in a spatial conformation which is unique to the epitope. Generally an epitope consists of at least 6 such amino acids, and more usually at least 8-10 such amino acids. Methods for determining the amino acids which make up an epitope include x-ray crystallography, 2-dimensional nuclear magnetic resonance, and epitope mapping e.g. the Pepscan method described by Geysen *et al.* 1984; PCT Publication No. WO 84/03564; and PCT Publication No. WO 84/03506.

Throughout the present specification, the expression “nucleotide sequence” may be employed to designate indifferently a polynucleotide or a nucleic acid. More precisely, the expression “nucleotide sequence” encompasses the nucleic material itself and is thus not restricted to the sequence information (*i.e.* the succession of letters chosen among the four base letters) that biochemically characterizes a specific DNA or RNA molecule.

As used interchangeably herein, the terms “nucleic acids”, “oligonucleotides”, and “polynucleotides” include RNA, DNA, or RNA/DNA hybrid sequences of more than one nucleotide in either single chain or duplex form. The term “nucleotide” as used herein as an adjective to describe molecules comprising RNA, DNA, or RNA/DNA hybrid sequences of any length in single-stranded or duplex form. The term “nucleotide” is also used herein as a noun to refer to individual nucleotides or varieties of nucleotides, meaning a molecule, or individual unit in a larger nucleic acid molecule, comprising a purine or pyrimidine, a ribose or deoxyribose sugar moiety, and a phosphate group, or phosphodiester linkage in the case of nucleotides within an oligonucleotide or polynucleotide. The term “nucleotide” is also used herein to encompass “modified nucleotides” which comprise at least one of the following modifications (a) an alternative linking group, (b) an analogous form of purine, (c) an analogous form of pyrimidine, or (d) an analogous sugar, for examples of analogous linking groups, purine, pyrimidines, and sugars see for example PCT publication No. WO 95/04064. The polynucleotide sequences of the invention may be prepared by any known method, including synthetic, recombinant, *ex vivo* generation, or a combination thereof, as well as utilizing any purification methods known in the art.

A “promoter” refers to a DNA sequence recognized by the synthetic machinery of the cell required to initiate the specific transcription of a gene.

A sequence which is “operably linked” to a regulatory sequence such as a promoter means that said regulatory element is in the correct location and orientation in relation to the nucleic acid
5 to control RNA polymerase initiation and expression of the nucleic acid of interest. As used herein, the term “operably linked” refers to a linkage of polynucleotide elements in a functional relationship. For instance, a promoter or enhancer is operably linked to a coding sequence if it affects the transcription of the coding sequence. More precisely, two DNA molecules (such as a polynucleotide containing a promoter region and a polynucleotide encoding a desired polypeptide
10 or polynucleotide) are said to be “operably linked” if the nature of the linkage between the two polynucleotides does not (1) result in the introduction of a frame-shift mutation or (2) interfere with the ability of the polynucleotide containing the promoter to direct the transcription of the coding polynucleotide.

The term “primer” denotes a specific oligonucleotide sequence which is complementary to
15 a target nucleotide sequence and used to hybridize to the target nucleotide sequence. A primer serves as an initiation point for nucleotide polymerization catalyzed by either DNA polymerase, RNA polymerase or reverse transcriptase.

The term “probe” denotes a defined nucleic acid segment (or nucleotide analog segment, e.g., polynucleotide as defined herein) which can be used to identify a specific polynucleotide
20 sequence present in samples, said nucleic acid segment comprising a nucleotide sequence complementary of the specific polynucleotide sequence to be identified.

The terms “trait” and “phenotype” are used interchangeably herein and refer to any visible, detectable or otherwise measurable property of an organism such as symptoms of, or susceptibility to a disease for example. Typically the terms “trait” or “phenotype” are used herein to refer to
25 symptoms of, or susceptibility to a disease, a beneficial response to or side effects related to a treatment. Preferably, said trait can be, without being limited to, cancers, developmental diseases, and neurological diseases.

The term “allele” is used herein to refer to variants of a nucleotide sequence. A biallelic polymorphism has two forms. Typically the first identified allele is designated as the original allele
30 whereas other alleles are designated as alternative alleles. Diploid organisms may be homozygous or heterozygous for an allelic form.

The term “heterozygosity rate” is used herein to refer to the incidence of individuals in a population which are heterozygous at a particular allele. In a biallelic system, the heterozygosity rate is on average equal to $2P_a(1-P_a)$, where P_a is the frequency of the least common allele. In order
35 to be useful in genetic studies, a genetic marker should have an adequate level of heterozygosity to allow a reasonable probability that a randomly selected person will be heterozygous.

The term "genotype" as used herein refers the identity of the alleles present in an individual or a sample. In the context of the present invention, a genotype preferably refers to the description of the biallelic marker alleles present in an individual or a sample. The term "genotyping" a sample or an individual for a biallelic marker consists of determining the specific allele or the specific
5 nucleotide carried by an individual at a biallelic marker.

The term "mutation" as used herein refers to a difference in DNA sequence between or among different genomes or individuals which has a frequency below 1%.

The term "haplotype" refers to a combination of alleles present in an individual or a sample. In the context of the present invention, a haplotype preferably refers to a combination of biallelic
10 marker alleles found in a given individual and which may be associated with a phenotype.

The term "polymorphism" as used herein refers to the occurrence of two or more alternative genomic sequences or alleles between or among different genomes or individuals. "Polymorphic" refers to the condition in which two or more variants of a specific genomic sequence can be found in a population. A "polymorphic site" is the locus at which the variation occurs. A single
15 nucleotide polymorphism is the replacement of one nucleotide by another nucleotide at the polymorphic site. Deletion of a single nucleotide or insertion of a single nucleotide also gives rise to single nucleotide polymorphisms. In the context of the present invention, "single nucleotide polymorphism" preferably refers to a single nucleotide substitution. Typically, between different individuals, the polymorphic site may be occupied by two different nucleotides.

20 The term "biallelic polymorphism" and "biallelic marker" are used interchangeably herein to refer to a single nucleotide polymorphism having two alleles at a fairly high frequency in the population. A "biallelic marker allele" refers to the nucleotide variants present at a biallelic marker site. Typically, the frequency of the less common allele of the biallelic markers of the present invention has been validated to be greater than 1%, preferably the frequency is greater than 10%,
25 more preferably the frequency is at least 20% (*i.e.* heterozygosity rate of at least 0.32), even more preferably the frequency is at least 30% (*i.e.* heterozygosity rate of at least 0.42). A biallelic marker wherein the frequency of the less common allele is 30% or more is termed a "high quality biallelic marker".

The location of nucleotides in a polynucleotide with respect to the center of the
30 polynucleotide are described herein in the following manner. When a polynucleotide has an odd number of nucleotides, the nucleotide at an equal distance from the 3' and 5' ends of the polynucleotide is considered to be "at the center" of the polynucleotide, and any nucleotide immediately adjacent to the nucleotide at the center, or the nucleotide at the center itself is considered to be "within 1 nucleotide of the center." With an odd number of nucleotides in a
35 polynucleotide any of the five nucleotides positions in the middle of the polynucleotide would be considered to be within 2 nucleotides of the center, and so on. When a polynucleotide has an even number of nucleotides, there would be a bond and not a nucleotide at the center of the

polynucleotide. Thus, either of the two central nucleotides would be considered to be "within 1 nucleotide of the center" and any of the four nucleotides in the middle of the polynucleotide would be considered to be "within 2 nucleotides of the center", and so on. For polymorphisms which involve the substitution, insertion or deletion of 1 or more nucleotides, the polymorphism, allele or biallelic marker is "at the center" of a polynucleotide if the difference between the distance from the substituted, inserted, or deleted polynucleotides of the polymorphism and the 3' end of the polynucleotide, and the distance from the substituted, inserted, or deleted polynucleotides of the polymorphism and the 5' end of the polynucleotide is zero or one nucleotide. If this difference is 0 to 3, then the polymorphism is considered to be "within 1 nucleotide of the center." If the difference is 0 to 5, the polymorphism is considered to be "within 2 nucleotides of the center." If the difference is 0 to 7, the polymorphism is considered to be "within 3 nucleotides of the center," and so on.

The term "upstream" is used herein to refer to a location which is toward the 5' end of the polynucleotide from a specific reference point.

The terms "base paired" and "Watson & Crick base paired" are used interchangeably herein to refer to nucleotides which can be hydrogen bonded to one another by virtue of their sequence identities in a manner like that found in double-helical DNA with thymine or uracil residues linked to adenine residues by two hydrogen bonds and cytosine and guanine residues linked by three hydrogen bonds (See Stryer, L., 1995).

The terms "complementary" or "complement thereof" are used herein to refer to the sequences of polynucleotides which is capable of forming Watson & Crick base pairing with another specified polynucleotide throughout the entirety of the complementary region. For the purpose of the present invention, a first polynucleotide is deemed to be complementary to a second polynucleotide when each base in the first polynucleotide is paired with its complementary base. Complementary bases are, generally, A and T (or A and U), or C and G. "Complement" is used herein as a synonym of "complementary polynucleotide", "complementary nucleic acid" and "complementary nucleotide sequence". These terms are applied to pairs of polynucleotides based solely upon their sequences and not any particular set of conditions under which the two polynucleotides would actually bind.

30 **Variants and Fragments**

1- Polynucleotides

The invention also relates to variants and fragments of the polynucleotides described herein, particularly of a PG-3 gene containing one or more biallelic markers according to the invention.

Variants of polynucleotides, as the term is used herein, are polynucleotides that differ from a reference polynucleotide. A variant of a polynucleotide may be a naturally occurring variant such as a naturally occurring allelic variant, or it may be a variant that is not known to occur naturally. Such non-naturally occurring variants of the polynucleotide may be made by mutagenesis

techniques, including those applied to polynucleotides, cells or organisms. Generally, differences are limited so that the nucleotide sequences of the reference and the variant are closely similar overall and, in many regions, identical.

Variants of polynucleotides according to the invention include, without being limited to,
5 nucleotide sequences which are at least 95% identical to a polynucleotide selected from the group consisting of the nucleotide sequences of SEQ ID Nos 1 and 2 or to any polynucleotide fragment of at least 12 consecutive nucleotides of a polynucleotide selected from the group consisting of the nucleotide sequences of SEQ ID Nos 1 and 2, and preferably at least 99% identical, more particularly at least 99.5% identical, and most preferably at least 99.8% identical to a polynucleotide
10 selected from the group consisting of the nucleotide sequences of SEQ ID Nos 1 and 2, or to any polynucleotide fragment of at least 12 consecutive nucleotides of a polynucleotide selected from the group consisting of the nucleotide sequences of SEQ ID Nos 1 and 2.

Nucleotide changes present in a variant polynucleotide may be silent, which means that they do not alter the amino acids encoded by the polynucleotide. However, nucleotide changes may
15 also result in amino acid substitutions, additions, deletions, fusions and truncations in the polypeptide encoded by the reference sequence. The substitutions, deletions or additions may involve one or more nucleotides. The variants may be altered in coding or non-coding regions or both. Alterations in the coding regions may produce conservative or non-conservative amino acid substitutions, deletions or additions.

20 In the context of the present invention, particularly preferred embodiments are those in which the polynucleotides encode polypeptides which retain substantially the same biological function or activity as the mature PG-3 protein, or those in which the polynucleotides encode polypeptides which maintain or increase a particular biological activity, while reducing a second biological activity.

25 A polynucleotide fragment is a polynucleotide having a sequence that is entirely the same as part but not all of a given nucleotide sequence, preferably the nucleotide sequence of a PG-3 gene, and variants thereof. The fragment can be a portion of an intron or an exon of a PG-3 gene. It can also be a portion of the regulatory regions of PG-3. Preferably, such fragments comprise at least one of the biallelic markers A1 to A80 or the complements thereto or a biallelic marker in
30 linkage disequilibrium with one or more of the biallelic markers A1 to A80.

Such fragments may be "free-standing", *i.e.* not part of or fused to other polynucleotides, or they may be comprised within a single larger polynucleotide of which they form a part or region. Indeed, several of these fragments may be present within a single larger polynucleotide.

Optionally, such fragments may comprise, consist of, or consist essentially of a contiguous
35 span of at least 8, 10, 12, 15, 18, 20, 25, 35, 40, 50, 70, 80, 100, 250, 500 or 1000 nucleotides in length. A set of preferred fragments contain at least one of the biallelic markers A1 to A80 of the PG-3 gene which are described herein or the complements thereto.

2- Polypeptides

The invention also relates to variants, fragments, analogs and derivatives of the polypeptides described herein, including mutated PG-3 proteins.

The variant may be 1) one in which one or more of the amino acid residues are substituted
5 with a conserved or non-conserved amino acid residue and such substituted amino acid residue may
or may not be one encoded by the genetic code, or 2) one in which one or more of the amino acid
residues includes a substituent group, or 3) one in which the mutated PG-3 is fused with another
compound, such as a compound to increase the half-life of the polypeptide (for example,
polyethylene glycol), or 4) one in which the additional amino acids are fused to the mutated PG-3,
10 such as a leader or secretory sequence or a sequence which is employed for purification of the
mutated PG-3 or a preprotein sequence. Such variants are deemed to be within the scope of those
skilled in the art.

A polypeptide fragment is a polypeptide having a sequence that is entirely the same as part
but not all of a given polypeptide sequence, preferably a polypeptide encoded by a PG-3 gene and
15 variants thereof.

In the case of an amino acid substitution in the amino acid sequence of a polypeptide
according to the invention, one or several amino acids can be replaced by "equivalent" amino acids.
The expression "equivalent" amino acid is used herein to designate any amino acid that may be
substituted for one of the amino acids having similar properties, such that one skilled in the art of
20 peptide chemistry would expect the secondary structure and hydropathic nature of the polypeptide
to be substantially unchanged. Generally, the following groups of amino acids represent equivalent
changes: (1) Ala, Pro, Gly, Glu, Asp, Gln, Asn, Ser, Thr; (2) Cys, Ser, Tyr, Thr; (3) Val, Ile, Leu,
Met, Ala, Phe; (4) Lys, Arg, His; (5) Phe, Tyr, Trp, His.

A specific embodiment of a modified PG-3 peptide molecule of interest according to the
25 present invention, includes, but is not limited to, a peptide molecule which is resistant to
proteolysis, a peptide in which the -CONH- peptide bond is modified and replaced by a (CH₂NH)
reduced bond, a (NHCO) retro inverso bond, a (CH₂-O) methylene-oxy bond, a (CH₂-S)
thiomethylene bond, a (CH₂CH₂) carba bond, a (CO-CH₂) cetomethylene bond, a (CHOH-CH₂)
hydroxyethylene bond), a (N-N) bound, a E-alcene bond or also a -CH=CH- bond. The invention
30 also encompasses a human PG-3 polypeptide or a fragment or a variant thereof in which at least one
peptide bond has been modified as described above.

Such fragments may be "free-standing", *i.e.* not part of or fused to other polypeptides, or
they may be included within a single larger polypeptide of which they form a part or region.
However, several fragments may be included within a single larger polypeptide.

35 As representative examples of polypeptide fragments of the invention, there may be
mentioned those which are from about 5, 6, 7, 8, 9 or 10 to 15, 10 to 20, 15 to 40, or 30 to 55 amino

acids long. Preferred are those fragments containing at least one amino acid mutation in the PG-3 protein.

Identity Between Nucleic Acids Or Polypeptides

The terms "percentage of sequence identity" and "percentage homology" are used interchangeably herein to refer to comparisons among polynucleotides and polypeptides, and are determined by comparing two optimally aligned sequences over a comparison window, wherein the portion of the polynucleotide or polypeptide sequence in the comparison window may comprise additions or deletions (*i.e.*, gaps) as compared to the reference sequence (which does not comprise additions or deletions) for optimal alignment of the two sequences. The percentage is calculated by determining the number of positions at which the identical nucleic acid base or amino acid residue occurs in both sequences to yield the number of matched positions, dividing the number of matched positions by the total number of positions in the window of comparison and multiplying the result by 100 to yield the percentage of sequence identity. Homology is evaluated using any of the variety of sequence comparison algorithms and programs known in the art. Such algorithms and programs include, but are by no means limited to, TBLASTN, BLASTP, FASTA, TFASTA, and CLUSTALW (Pearson and Lipman, 1988; Altschul *et al.*, 1990; Thompson *et al.*, 1994; Higgins *et al.*, 1996; Altschul *et al.*, 1993). In a particularly preferred embodiment, protein and nucleic acid sequence homologies are evaluated using the Basic Local Alignment Search Tool ("BLAST") which is well known in the art (see, e.g., Karlin and Altschul, 1990; Altschul *et al.*, 1990, 1993, 1997). In particular, five specific BLAST programs are used to perform the following task:

- (1) BLASTP and BLAST3 compare an amino acid query sequence against a protein sequence database;
- (2) BLASTN compares a nucleotide query sequence against a nucleotide sequence database;
- (3) BLASTX compares the six-frame conceptual translation products of a query nucleotide sequence (both strands) against a protein sequence database;
- (4) TBLASTN compares a query protein sequence against a nucleotide sequence database translated in all six reading frames (both strands); and
- (5) TBLASTX compares the six-frame translations of a nucleotide query sequence against the six-frame translations of a nucleotide sequence database.

The BLAST programs identify homologous sequences by identifying similar segments, which are referred to herein as "high-scoring segment pairs," between a query amino or nucleic acid sequence and a test sequence which is preferably obtained from a protein or nucleic acid sequence database. High-scoring segment pairs are preferably identified (*i.e.*, aligned) by means of a scoring matrix, many of which are known in the art. Preferably, the scoring matrix used is the BLOSUM62 matrix (Gonnet *et al.*, 1992; Henikoff and Henikoff, 1993). Less preferably, the PAM or PAM250 matrices may also be used (see, e.g., Schwartz and Dayhoff, eds., 1978). The BLAST programs

evaluate the statistical significance of all high-scoring segment pairs identified, and preferably selects those segments which satisfy a user-specified threshold of significance, such as a user-specified percent homology. Preferably, the statistical significance of a high-scoring segment pair is evaluated using the statistical significance formula of Karlin (see, e.g., Karlin and Altschul, 5 1990). The BLAST programs may be used with the default parameters which are implemented in the absence of further instructions from the user. Alternatively, the BLAST programs may be used with parameters specified by the user.

Stringent Hybridization Conditions

By way of example and not limitation, procedures using conditions of high stringency are 10 as follows: Prehybridization of filters containing DNA is carried out for 8 h to overnight at 65°C in buffer composed of 6X SSC, 50 mM Tris-HCl (pH 7.5), 1 mM EDTA, 0.02% PVP, 0.02% Ficoll, 0.02% BSA, and 500 µg/ml denatured salmon sperm DNA. Filters are hybridized for 48 h at 65°C, the preferred hybridization temperature, in prehybridization mixture containing 100 µg/ml denatured salmon sperm DNA and 5-20 X 10⁶ cpm of ³²P-labeled probe. Alternatively, the 15 hybridization step can be performed at 65°C in the presence of SSC buffer, 1X SSC corresponding to 0.15M NaCl and 0.05 M Na citrate. Subsequently, filter washes can be done at 37°C for 1 h in a solution containing 2X SSC, 0.01% PVP, 0.01% Ficoll, and 0.01% BSA, followed by a wash in 0.1X SSC at 50°C for 45 min. Alternatively, filter washes can be performed in a solution containing 2X SSC and 0.1% SDS, or 0.5X SSC and 0.1% SDS, or 0.1X SSC and 0.1% SDS at 20 68°C for 15 minute intervals. Following the wash steps, the hybridized probes are detectable by autoradiography. Other conditions of high stringency which may be used are well known in the art and are cited in Sambrook *et al.*, 1989; and Ausubel *et al.*, 1989. These hybridization conditions are suitable for a nucleic acid molecule of about 20 nucleotides in length. There is no need to say that the hybridization conditions described above are to be adapted according to the length of the 25 desired nucleic acid, following techniques well known to the one skilled in the art. The suitable hybridization conditions may for example be adapted according to the teachings disclosed in Hames and Higgins (1985) or in Sambrook *et al.*(1989).

GENOMIC SEQUENCES OF THE PG-3 GENE

The present invention concerns the genomic sequence of PG-3. The present invention 30 encompasses the PG-3 gene, or PG-3 genomic sequences consisting of, consisting essentially of, or comprising the sequence of SEQ ID No 1, sequences complementary thereto, as well as fragments and variants thereof. These polynucleotides may be purified, isolated, or recombinant.

The invention also encompasses a purified, isolated, or recombinant polynucleotide comprising a nucleotide sequence having at least 70, 75, 80, 85, 90, or 95% nucleotide identity with 35 the nucleotide sequence of SEQ ID No 1 or a complementary sequence thereto or a fragment thereof. The nucleotide differences with regard to the nucleotide sequence of SEQ ID No 1 may be generally randomly distributed throughout the entire nucleic acid. Nevertheless, preferred nucleic

acids are those wherein the nucleotide differences as regards to the nucleotide sequence of SEQ ID No 1 are predominantly located outside the coding sequences contained in the exons. These nucleic acids, as well as their fragments and variants, may be used as oligonucleotide primers or probes in order to detect the presence of a copy of the PG-3 gene in a test sample, or alternatively in order to

5 amplify a target nucleotide sequence within the PG-3 sequences.

Another object of the invention relates to a purified, isolated, or recombinant nucleic acid that hybridizes with the nucleotide sequence of SEQ ID No 1 or a complementary sequence thereto or a variant thereof, under the stringent hybridization conditions as defined above.

Particularly preferred nucleic acids of the invention include isolated, purified, or

10 recombinant polynucleotides comprising a contiguous span of at least 12, 15, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 500, or 1000 nucleotides of SEQ ID No 1 or the complements thereof, wherein said contiguous span comprises at least 1, 2, 3, 5, or 10 of the following nucleotide positions of SEQ ID No 1: 1-97921, 98517-103471, 103603-108222, 108390-109221, 109324-114409, 114538-115723, 115957-122102, 122225-126876, 127033-157212, 157808-240825.

15 Additional preferred nucleic acids of the invention include isolated, purified, or recombinant polynucleotides comprising a contiguous span of at least 12, 15, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 500, or 1000 nucleotides of SEQ ID No 1 or the complements thereof, wherein said contiguous span comprises at least 1, 2, 3, 5, or 10 of the following nucleotide positions of SEQ ID No 1: 1-10000, 10001-20000, 20001-30000, 30001-40000, 40001-50000,

20 50001-60000, 60001-70000, 70001-80000, 80001-90000, 90001-97921, 98517-103471, 103603-108222, 108390-109221, 109324-114409, 114538-115723, 115957-122102, 122225-126876, 127033-157212, 157808-159000, 159001-160000, 160001-170000, 170001-180000, 180001-190000, 190001-200000, 200001-210000, 210001-220000, 220001-230000, 230001-240825. It should be noted that nucleic acid fragments of any size and sequence may also be comprised by the

25 polynucleotides described in this section.

The PG-3 genomic nucleic acid comprises 14 exons. The exon positions in SEQ ID No 1 are detailed below in Table A.

Table A

Exon	Position in SEQ ID No 1		Intron	Position in SEQ ID No 1	
	Beginning	End		Beginning	End
A	2001	2079	A-B	2080	4626
B	4627	4718	B-C	4719	10114
C	10115	10233	C-D	10234	26809
D	26810	26897	D-E	26898	31356
E	31357	31471	E-F	31472	34260
F	34261	34404	F-S	34405	37376
S	37377	37466	S-T	37467	39703
T	39704	40858	T-G	40859	50435
G	50436	50545	G-H	50546	72880
H	72881	72918	H-I	72919	75988
I	75989	76151	I-J	76152	95110

J	95111	95188	J-K	95189	216014
K	216015	216252	K-L	216253	237525
L	237526	238825			

Thus, the invention embodies purified, isolated, or recombinant polynucleotides comprising a nucleotide sequence selected from the group consisting of the 14 exons of the PG-3 gene, or a sequence complementary thereto. The invention also relates to purified, isolated, or recombinant nucleic acids comprising a combination of at least two exons of the PG-3 gene, wherein the polynucleotides are arranged within the nucleic acid, from the 5'-end to the 3'-end of said nucleic acid, in the same order as in SEQ ID No 1.

Intron A-B refers to the nucleotide sequence located between Exon A and Exon B, and so on. The position of the introns is detailed in Table A. The intron J-K is large. Indeed, it is 120 kb in length and comprises the whole angiopoietine gene.

Thus, the invention embodies purified, isolated, or recombinant polynucleotides comprising a nucleotide sequence selected from the group consisting of the 13 introns of the PG-3 gene, or a sequence complementary thereto.

While this section is entitled "Genomic Sequences of PG-3," it should be noted that nucleic acid fragments of any size and sequence may also be comprised by the polynucleotides described in this section, flanking the genomic sequences of PG-3 on either side or between two or more such genomic sequences.

PG-3 CDNA SEQUENCES

The expression of the PG-3 gene has been shown to lead to the production of at least one mRNA species which nucleic acid sequence is set forth in SEQ ID No 2. Three cDNAs have been independently cloned. They all have the same size but exhibit strong polymorphism between each other and between each cDNA and the genomic sequence. These polymorphisms are indicated in the appended sequence listing by the use of the feature "variation" in SEQ ID No 2.

Another object of the invention is a purified, isolated, or recombinant nucleic acid comprising the nucleotide sequence of SEQ ID No 2, complementary sequences thereto, as well as allelic variants, and fragments thereof. Moreover, preferred polynucleotides of the invention include purified, isolated, or recombinant PG-3 cDNAs consisting of, consisting essentially of, or comprising the sequence of SEQ ID No 2. Particularly preferred nucleic acids of the invention include isolated, purified, or recombinant polynucleotides comprising a contiguous span of at least 12, 15, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 500, or 1000 nucleotides of SEQ ID No 2 or the complements thereof. Additional preferred embodiments of the invention include isolated, purified, or recombinant polynucleotides comprising a contiguous span of at least 12, 15, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 500, or 1000 nucleotides of SEQ ID No 2 or the complements thereof, wherein said contiguous span comprises at least 1, 2, 3, 5, or 10 of the

following nucleotide positions of SEQ ID No 2: 1-500, 501-1000, 1001-1500, 1501-2000, 2001-2500, 2501-3000, 3001-3500, 3501-3809.

The invention also pertains to a purified or isolated nucleic acid comprising a polynucleotide having at least 80, 85, 90, or 95% nucleotide identity with a polynucleotide of SEQ ID No 2, advantageously 99 % nucleotide identity, preferably 99.5% nucleotide identity and most preferably 99.8% nucleotide identity with a polynucleotide of SEQ ID No 2, or a sequence complementary thereto or a biologically active fragment thereof.

Another object of the invention relates to purified, isolated or recombinant nucleic acids comprising a polynucleotide that hybridizes, under the stringent hybridization conditions defined herein, with a polynucleotide of SEQ ID No 2, or a sequence complementary thereto or a variant thereof or a biologically active fragment thereof.

The cDNA of SEQ ID No 2 includes a 5'-UTR region starting from the nucleotide at position 1 and ending at the nucleotide in position 57 of SEQ ID No 2. The cDNA of SEQ ID No 2 includes a 3'-UTR region starting from the nucleotide at position 2566 and ending at the nucleotide at position 3809 of SEQ ID No 2. The polyadenylation signal starts from the nucleotide at position 3795 and ends at the nucleotide in position 3800 of SEQ ID No 2.

Consequently, the invention concerns a purified, isolated, or recombinant nucleic acid comprising a nucleotide sequence of the 5'UTR of the PG-3 cDNA, a sequence complementary thereto, or an allelic variant thereof. The invention also concerns a purified, isolated, or recombinant nucleic acid comprising a nucleotide sequence of the 3'UTR of the PG-3 cDNA, a sequence complementary thereto, or an allelic variant thereof.

While this section is entitled "PG-3 cDNA Sequences," it should be noted that nucleic acid fragments of any size and sequence may also be comprised by the polynucleotides described in this section, flanking the PG-3 sequences on either side or between two or more such PG-3 sequences.

25

CODING REGIONS

The PG-3 open reading frame is contained in the corresponding mRNA of SEQ ID No 2. More precisely, the effective PG-3 coding sequence (CDS) includes the region between nucleotide position 58 (first nucleotide of the ATG codon) and nucleotide position 2565 (end nucleotide of the TGA codon) of SEQ ID No 2.

The present invention also embodies isolated, purified, and recombinant polynucleotides which encode a polypeptide comprising a contiguous span of at least 6 amino acids, preferably at least 8 or 10 amino acids, more preferably at least 12, 15, 20, 25, 30, 40, 50, or 100 amino acids of SEQ ID No 3. Preferably, the present invention also embodies isolated, purified, and recombinant polynucleotides which encode a polypeptide comprising a contiguous span of at least 6 amino acids, preferably at least 8 or 10 amino acids, more preferably at least 12, 15, 20, 25, 30, 40, 50, or 100 amino acids of SEQ ID No 3, wherein wherein said contiguous span comprises at least 1, 2, 3, 5, or

10 of the following amino acid positions of SEQ ID No 3: 1-100, 101-200, 201-300, 301-400, 401-500, 501-600, 601-700, 701-835.

The above disclosed polynucleotide that contains the coding sequence of the PG-3 gene may be expressed in a desired host cell or a desired host organism, when this polynucleotide is placed under the control of suitable expression signals. The expression signals may be either the expression signals contained in the regulatory regions in the PG-3 gene of the invention or in contrast the signals may be exogenous regulatory nucleic sequences. Such a polynucleotide, when placed under the suitable expression signals, may also be inserted in a vector for its expression and/or amplification.

10

REGULATORY SEQUENCES OF PG-3

As mentioned, the genomic sequence of the PG-3 gene contains regulatory sequences both in the non-transcribed 5'-flanking region and in the non-transcribed 3'-flanking region that border the PG-3 coding region containing the 14 exons of this gene.

The 5' regulatory region of the PG-3 gene is localized between the nucleotide in position 1 and the nucleotide in position 2000 of the nucleotide sequence of SEQ ID No 1. The 3' regulatory region of the PG-3 gene is localized between nucleotide position 238826 and nucleotide position 240825 of SEQ ID No 1.

Polynucleotides derived from the 5' and 3' regulatory regions are useful in order to detect the presence of at least a copy of a nucleotide sequence of SEQ ID No 1 or a fragment thereof in a test sample.

The promoter activity of the 5' regulatory regions contained in PG-3 can be assessed as described below.

In order to identify the relevant biologically active polynucleotide fragments or variants of SEQ ID No 1, one of skill in the art will refer to the book of Sambrook *et al.* (1989) which describes the use of a recombinant vector carrying a marker gene (*i.e.* beta galactosidase, chloramphenicol acetyl transferase, etc.) the expression of which will be detected when placed under the control of a biologically active polynucleotide fragments or variants of SEQ ID No 1. Genomic sequences located upstream of the first exon of the PG-3 gene are cloned into a suitable promoter reporter vector, such as the pSEAP-Basic, pSEAP-Enhancer, p β gal-Basic, p β gal-Enhancer, or pEGFP-1 Promoter Reporter vectors available from Clontech, or pGL2-basic or pGL3-basic promoterless luciferase reporter gene vector from Promega. Briefly, each of these promoter reporter vectors include multiple cloning sites positioned upstream of a reporter gene encoding a readily assayable protein such as secreted alkaline phosphatase, luciferase, β galactosidase, or green fluorescent protein. The sequences upstream the PG-3 coding region are inserted into the cloning sites upstream of the reporter gene in both orientations and introduced into an appropriate host cell. The level of reporter protein is assayed and compared to the level obtained from a vector which lacks an insert in the cloning site. The presence of an elevated expression level in the vector containing the

insert with respect to the control vector indicates the presence of a promoter in the insert. If necessary, the upstream sequences can be cloned into vectors which contain an enhancer for increasing transcription levels from weak promoter sequences. A significant level of expression above that observed with the vector lacking an insert indicates that a promoter sequence is present
5 in the inserted upstream sequence.

Promoter sequences within the upstream genomic DNA may be further defined by constructing nested 5' and/or 3' deletions in the upstream DNA using conventional techniques such as Exonuclease III or appropriate restriction endonuclease digestion. The resulting deletion fragments can be inserted into the promoter reporter vector to determine whether the deletion has
10 reduced or obliterated promoter activity, such as described, for example, by Coles *et al.* (1998). In this way, the boundaries of the promoters may be defined. If desired, potential individual regulatory sites within the promoter may be identified using site directed mutagenesis or linker scanning to obliterate potential transcription factor binding sites within the promoter individually or in combination. The effects of these mutations on transcription levels may be determined by
15 inserting the mutations into cloning sites in promoter reporter vectors. This type of assay is well-known to those skilled in the art and is described in WO 97/17359, US Patent No. 5,374,544; EP 582 796; US Patent No. 5,698,389; US 5,643,746; US Patent No. 5,502,176; and US Patent 5,266,488.

The strength and the specificity of the promoter of the PG-3 gene can be assessed through
20 the expression levels of a detectable polynucleotide operably linked to the PG-3 promoter in different types of cells and tissues. The detectable polynucleotide may be either a polynucleotide that specifically hybridizes with a predefined oligonucleotide probe, or a polynucleotide encoding a detectable protein, including a PG-3 polypeptide or a fragment or a variant thereof. This type of assay is well-known to those skilled in the art and is described in US Patent No. 5,502,176; and US
25 Patent No. 5,266,488. Some of the methods are discussed in more detail below.

Polynucleotides carrying the regulatory elements located at the 5' end and at the 3' end of the PG-3 coding region may be advantageously used to control the transcriptional and translational activity of an heterologous polynucleotide of interest.

Thus, the present invention also concerns a purified or isolated nucleic acid comprising a
30 polynucleotide which is selected from the group consisting of the 5' and 3' regulatory regions, or a sequence complementary thereto or a biologically active fragment or variant thereof.

The invention also pertains to a purified or isolated nucleic acid comprising a polynucleotide having at least 80, 85, 90, or 95% nucleotide identity with a polynucleotide selected from the group consisting of the 5' and 3' regulatory regions, advantageously 99 % nucleotide
35 identity, preferably 99.5% nucleotide identity and most preferably 99.8% nucleotide identity with a polynucleotide selected from the group consisting of the 5' and 3' regulatory regions, or a sequence complementary thereto or a variant thereof or a biologically active fragment thereof.

Another object of the invention relates to purified, isolated or recombinant nucleic acids comprising a polynucleotide that hybridizes, under the stringent hybridization conditions defined herein, with a polynucleotide selected from the group consisting of the nucleotide sequences of the 5'- and 3' regulatory regions, or a sequence complementary thereto or a variant thereof or a
5 biologically active fragment thereof.

Preferred fragments of the 5' regulatory region have a length of about 1500 or 1000 nucleotides, preferably of about 500 nucleotides, more preferably about 400 nucleotides, even more preferably 300 nucleotides and most preferably about 200 nucleotides.

Preferred fragments of the 3' regulatory region are at least 50, 100, 150, 200, 300 or 400
10 bases in length.

"Biologically active" polynucleotide derivatives of SEQ ID No 1 are polynucleotides comprising or alternatively consisting essentially of or consisting of a fragment of said polynucleotide which is functional as a regulatory region for expressing a recombinant polypeptide or a recombinant polynucleotide in a recombinant cell host. It could act either as an enhancer or as
15 a repressor.

For the purpose of the invention, a nucleic acid or polynucleotide is "functional" as a regulatory region for expressing a recombinant polypeptide or a recombinant polynucleotide if said regulatory polynucleotide contains nucleotide sequences which contain transcriptional and translational regulatory information, and such sequences are "operably linked" to nucleotide
20 sequences which encode the desired polypeptide or the desired polynucleotide.

The regulatory polynucleotides of the invention may be prepared from the nucleotide sequence of SEQ ID No 1 by cleavage using suitable restriction enzymes, as described for example in the book of Sambrook *et al.* (1989). The regulatory polynucleotides may also be prepared by digestion of SEQ ID No 1 by an exonuclease enzyme, such as Bal31 (Wabiko *et al.*, 1986). These
25 regulatory polynucleotides can also be prepared by nucleic acid chemical synthesis, as described elsewhere in the specification.

The regulatory polynucleotides according to the invention may be part of a recombinant expression vector that may be used to express a coding sequence in a desired host cell or host organism. The recombinant expression vectors according to the invention are described elsewhere
30 in the specification.

A preferred 5'-regulatory polynucleotide of the invention includes the 5'-untranslated region (5'-UTR) of the PG-3 cDNA, or a biologically active fragment or variant thereof.

A preferred 3'-regulatory polynucleotide of the invention includes the 3'-untranslated region (3'-UTR) of the PG-3 cDNA, or a biologically active fragment or variant thereof.

35 A further object of the invention relates to a purified or isolated nucleic acid comprising:

a) a nucleic acid comprising a regulatory nucleotide sequence selected from the group consisting of:

(i) a nucleotide sequence comprising a polynucleotide of the 5' regulatory region or a complementary sequence thereto; or

(ii) a nucleotide sequence comprising a polynucleotide having at least 80, 85, 90, or 95% of nucleotide identity with the nucleotide sequence of the 5' regulatory region or a complementary sequence thereto; or

(iii) a nucleotide sequence comprising a polynucleotide that hybridizes under stringent hybridization conditions with the nucleotide sequence of the 5' regulatory region or a complementary sequence thereto; or

(iv) a biologically active fragment or variant of the polynucleotides in (i), (ii) and (iii);

b) a polynucleotide encoding a desired polypeptide or a nucleic acid of interest, operably linked to the nucleic acid defined in (a) above;

c) Optionally, a nucleic acid comprising a 3'- regulatory polynucleotide, preferably a 3'- regulatory polynucleotide of the PG-3 gene.

In a specific embodiment of the nucleic acid defined above, said nucleic acid includes the 5'-untranslated region (5'-UTR) of the PG-3 cDNA, or a biologically active fragment or variant thereof.

In a second specific embodiment of the nucleic acid defined above, said nucleic acid includes the 3'-untranslated region (3'-UTR) of the PG-3 cDNA, or a biologically active fragment or variant thereof.

The regulatory polynucleotide of the 5' regulatory region, or its biologically active fragments or variants, is operably linked at the 5'-end of the polynucleotide encoding the desired polypeptide or polynucleotide.

The regulatory polynucleotide of the 3' regulatory region, or its biologically active fragments or variants, is advantageously operably linked at the 3'-end of the polynucleotide encoding the desired polypeptide or polynucleotide.

The desired polypeptide encoded by the above-described nucleic acid may be of various nature or origin, encompassing proteins of prokaryotic or eukaryotic origin. Among the polypeptides which may be expressed under the control of a PG-3 regulatory region are bacterial, fungal or viral antigens. Also encompassed are eukaryotic proteins such as intracellular proteins, like "house keeping" proteins, membrane-bound proteins, like receptors, and secreted proteins like endogenous mediators such as cytokines. The desired polypeptide may be the PG-3 protein, especially the protein of the amino acid sequence of SEQ ID No 3, or a fragment or a variant thereof.

The desired nucleic acids encoded by the above-described polynucleotide, usually an RNA molecule, may be complementary to a desired coding polynucleotide, for example to the PG-3 coding sequence, and thus useful as an antisense polynucleotide.

Such a polynucleotide may be included in a recombinant expression vector in order to express the desired polypeptide or the desired nucleic acid in host cell or in a host organism. Suitable recombinant vectors that contain a polynucleotide such as described herein are disclosed elsewhere in the specification.

5 **POLYNUCLEOTIDE CONSTRUCTS**

The terms "polynucleotide construct" and "recombinant polynucleotide" are used interchangeably herein to refer to linear or circular, purified or isolated polynucleotides that have been artificially designed and which comprise at least two nucleotide sequences that are not found as contiguous nucleotide sequences in their initial natural environment.

10 **DNA Construct That Enables Temporal And Spatial PG-3 Gene Expression In Recombinant Cell Hosts And In Transgenic Animals.**

In order to study the physiological and phenotypic consequences of a lack of synthesis of the PG-3 protein, both at the cell level and at the multi cellular organism level, the invention also encompasses DNA constructs and recombinant vectors enabling a conditional expression of a
 15 specific allele of the PG-3 genomic sequence or cDNA and also of a copy of this genomic sequence or cDNA harboring substitutions, deletions, or additions of one or more bases as regards to the PG-3 nucleotide sequence of SEQ ID Nos 1 and 2, or a fragment thereof, these base substitutions, deletions or additions being located either in an exon, an intron or a regulatory sequence, but preferably in the 5'-regulatory sequence or in an exon of the PG-3 genomic sequence or within the
 20 PG-3 cDNA of SEQ ID No 2. In a preferred embodiment, the PG-3 sequence comprises a biallelic marker of the present invention. In a preferred embodiment, the PG-3 sequence comprises at least one of the biallelic markers A1 to A80.

The present invention embodies recombinant vectors comprising any one of the polynucleotides described in the present invention. More particularly, the polynucleotide constructs
 25 according to the present invention can comprise any of the polynucleotides described in the "Genomic Sequences Of The PG3 Gene" section, the "PG-3 cDNA Sequences" section, the "Coding Regions" section, and the "Oligonucleotide Probes And Primers" section.

A first preferred DNA construct is based on the tetracycline resistance operon *tet* from *E. coli* transposon Tn10 for controlling the PG-3 gene expression, such as described by Gossen *et al.* (1992, 1995) and Furth *et al.* (1994). Such a DNA construct contains seven *tet* operator
 30 sequences from Tn10 (*tetop*) that are fused to either a minimal promoter or a 5'-regulatory sequence of the PG-3 gene, said minimal promoter or said PG-3 regulatory sequence being operably linked to a polynucleotide of interest that codes either for a sense or an antisense oligonucleotide or for a polypeptide, including a PG-3 polypeptide or a peptide fragment thereof. This DNA construct is
 35 functional as a conditional expression system for the nucleotide sequence of interest when the same cell also comprises a nucleotide sequence coding for either the wild type (tTA) or the mutant (rtTA) repressor fused to the activating domain of viral protein VP16 of herpes simplex virus, placed

under the control of a promoter, such as the HCMVIE1 enhancer/promoter or the MMTV-LTR. Indeed, a preferred DNA construct of the invention comprises both the polynucleotide containing the *tet* operator sequences and the polynucleotide containing a sequence coding for the tTA or the rTA repressor.

- 5 In a specific embodiment, the conditional expression DNA construct contains the sequence encoding the mutant tetracycline repressor rTA, the expression of the polynucleotide of interest is silent in the absence of tetracycline and induced in its presence.

DNA Constructs Allowing Homologous Recombination: Replacement Vectors

- A second preferred DNA construct comprises, from 5'-end to 3'-end: (a) a first nucleotide
10 sequence that is included within the PG-3 genomic sequence; (b) a nucleotide sequence comprising a positive selection marker, such as the marker for neomycine resistance (*neo*); and (c) a second nucleotide sequence that is included within the PG-3 genomic sequence, and is located on the genome downstream the first PG-3 nucleotide sequence (a).

- In a preferred embodiment, this DNA construct also comprises a negative selection marker
15 located upstream of the nucleotide sequence (a) or downstream from the nucleotide sequence (c). Preferably, the negative selection marker comprises of the thymidine kinase (*tk*) gene (Thomas *et al.*, 1986), the hygromycine beta gene (Te Riele *et al.*, 1990), the *hprt* gene (Van der Lugt *et al.*, 1991; Reid *et al.*, 1990) or the Diphtheria toxin A fragment (*Dt-A*) gene (Nada *et al.*, 1993; Yagi *et al.* 1990). Preferably, the positive selection marker is located within a PG-3 exon sequence so as to
20 interrupt the sequence encoding a PG-3 protein. These replacement vectors are described, for example, by Thomas *et al.* (1986; 1987), Mansour *et al.* (1988) and Koller *et al.* (1992).

- The first and second nucleotide sequences (a) and (c) may be indifferently located within a PG-3 regulatory sequence, an intronic sequence, an exon sequence or a sequence containing both regulatory and/or intronic and/or exon sequences. The size of the nucleotide sequences (a) and (c)
25 ranges from 1 to 50 kb, preferably from 1 to 10 kb, more preferably from 2 to 6 kb and most preferably from 2 to 4 kb.

DNA Constructs Allowing Homologous Recombination: Cre-LoxP System

- These new DNA constructs make use of the site specific recombination system of the P1 phage. The P1 phage possesses a recombinase called Cre which interacts specifically with a 34
30 base pairs *loxP* site. The *loxP* site is composed of two palindromic sequences of 13 bp separated by a 8 bp conserved sequence (Hoess *et al.*, 1986). The recombination by the Cre enzyme between two *loxP* sites having an identical orientation leads to the deletion of the DNA fragment.

- The Cre-*loxP* system used in combination with a homologous recombination technique has been first described by Gu *et al.* (1993, 1994). Briefly, a nucleotide sequence of interest to be
35 inserted in a targeted location of the genome harbors at least two *loxP* sites in the same orientation and located at the respective ends of a nucleotide sequence to be excised from the recombinant genome. The excision event requires the presence of the recombinase (Cre) enzyme within the

nucleus of the recombinant cell host. The recombinase enzyme may be provided at the desired time either by (a) incubating the recombinant cell hosts in a culture medium containing this enzyme, by injecting the Cre enzyme directly into the desired cell, such as described by Araki *et al.*(1995), or by lipofection of the enzyme into the cells, such as described by Baubonis *et al.*(1993); (b) 5 transfecting the cell host with a vector comprising the *Cre* coding sequence operably linked to a promoter functional in the recombinant host cell, said promoter being optionally inducible, said vector being introduced in the recombinant cell host, such as described by Gu *et al.*(1993) and Sauer *et al.*(1988); (c) introducing in the genome of the cell host a polynucleotide comprising the *Cre* coding sequence operably linked to a promoter functional in the recombinant cell host, which 10 promoter is optionally inducible, and said polynucleotide being inserted in the genome of the cell host either by a random insertion event or an homologous recombination event, such as described by Gu *et al.*(1994).

In a specific embodiment, the vector containing the sequence to be inserted in the PG-3 gene by homologous recombination is constructed in such a way that selectable markers are flanked 15 by *loxP* sites of the same orientation, it is possible, by treatment by the Cre enzyme, to eliminate the selectable markers while leaving the PG-3 sequences of interest that have been inserted by an homologous recombination event. Again, two selectable markers are needed: a positive selection marker to select for the recombination event and a negative selection marker to select for the homologous recombination event. Vectors and methods using the Cre-*loxP* system are described by 20 Zou *et al.*(1994).

Thus, a third preferred DNA construct of the invention comprises, from 5'-end to 3'-end: (a) a first nucleotide sequence that is included in the PG-3 genomic sequence; (b) a nucleotide sequence comprising a polynucleotide encoding a positive selection marker, said nucleotide sequence comprising additionally two sequences defining a site recognized by a recombinase, such as a *loxP* 25 site, the two sites being placed in the same orientation; and (c) a second nucleotide sequence that is included in the PG-3 genomic sequence, and is located on the genome downstream of the first PG-3 nucleotide sequence (a).

The sequences defining a site recognized by a recombinase, such as a *loxP* site, are preferably located within the nucleotide sequence (b) at suitable locations bordering the nucleotide 30 sequence for which the conditional excision is sought. In one specific embodiment, two *loxP* sites are located at each side of the positive selection marker sequence, in order to allow its excision at a desired time after the occurrence of the homologous recombination event.

In a preferred embodiment of a method using the third DNA construct described above, the excision of the polynucleotide fragment bordered by the two sites recognized by a recombinase, 35 preferably two *loxP* sites, is performed at a desired time, due to the presence within the genome of the recombinant host cell of a sequence encoding the Cre enzyme operably linked to a promoter sequence, preferably an inducible promoter, more preferably a tissue-specific promoter sequence

and most preferably a promoter sequence which is both inducible and tissue-specific, such as described by Gu *et al.*(1994).

The presence of the Cre enzyme within the genome of the recombinant cell host may result from the breeding of two transgenic animals, the first transgenic animal bearing the PG-3-derived
5 sequence of interest containing the *loxP* sites as described above and the second transgenic animal bearing the *Cre* coding sequence operably linked to a suitable promoter sequence, such as described by Gu *et al.*(1994).

Spatio-temporal control of the Cre enzyme expression may also be achieved with an adenovirus based vector that contains the Cre gene thus allowing infection of cells, or *in vivo*
10 infection of organs, for delivery of the Cre enzyme, such as described by Anton *et al.* (1995) and Kanegae *et al.*(1995).

The DNA constructs described above may be used to introduce a desired nucleotide sequence of the invention, preferably a PG-3 genomic sequence or a PG-3 cDNA sequence, and most preferably an altered copy of a PG-3 genomic or cDNA sequence, within a predetermined
15 location of the targeted genome, leading either to the generation of an altered copy of a targeted gene (knock-out homologous recombination) or to the replacement of a copy of the targeted gene by another copy sufficiently homologous to allow an homologous recombination event to occur (knock-in homologous recombination). In a specific embodiment, the DNA constructs described above may be used to introduce a PG-3 genomic sequence or a PG-3 cDNA sequence comprising at
20 least one biallelic marker of the present invention, preferably at least one biallelic marker selected from the group consisting of A1 to A80.

Nuclear Antisense DNA Constructs

Other compositions comprise a vector of the invention comprising an oligonucleotide fragment of the nucleic acid sequence of SEQ ID No 2, preferably a fragment including the start
25 codon of the PG-3 gene, as an antisense tool that inhibits the expression of the corresponding PG-3 gene. Preferred methods using antisense polynucleotide according to the present invention are the procedures described by Sczakiel *et al.*(1995) or those described in PCT Application No WO 95/24223.

Preferably, the antisense tools are chosen among the polynucleotides (15-200 bp long) that
30 are complementary to the 5'end of the PG-3 mRNA. In one embodiment, a combination of different antisense polynucleotides complementary to different parts of the desired targeted gene are used.

Preferred antisense polynucleotides according to the present invention are complementary to a sequence of the mRNAs of PG-3 that contains either the translation initiation codon ATG or a splicing site. Further preferred antisense polynucleotides according to the invention are
35 complementary of the splicing site of the PG-3 mRNA.

Preferably, the antisense polynucleotides of the invention have a 3' polyadenylation signal that has been replaced with a self-cleaving ribozyme sequence, such that RNA polymerase II

transcripts are produced without poly(A) at their 3' ends, these antisense polynucleotides being incapable of export from the nucleus, such as described by Liu *et al.*(1994). In a preferred embodiment, these PG-3 antisense polynucleotides also comprise, within the ribozyme cassette, a histone stem-loop structure to stabilize cleaved transcripts against 3'-5' exonucleolytic degradation, such as the structure described by Eckner *et al.*(1991).

Oligonucleotide Probes And Primers

Polynucleotides derived from the PG-3 gene are useful in order to detect the presence of at least a copy of a nucleotide sequence of SEQ ID No 1, or a fragment, complement, or variant thereof in a test sample.

Particularly preferred probes and primers of the invention include isolated, purified, or recombinant polynucleotides comprising a contiguous span of at least 12, 15, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 500, or 1000 nucleotides of SEQ ID No 1 or the complements thereof, wherein said contiguous span comprises at least 1, 2, 3, 5, or 10 of the following nucleotide positions of SEQ ID No 1: 1-97921, 98517-103471, 103603-108222, 108390-109221, 109324-114409, 114538-115723, 115957-122102, 122225-126876, 127033-157212, 157808-240825.

Additional preferred probes and primers of the invention include isolated, purified, or recombinant polynucleotides comprising a contiguous span of at least 12, 15, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 500, or 1000 nucleotides of SEQ ID No 1 or the complements thereof, wherein said contiguous span comprises at least 1, 2, 3, 5, or 10 of the following nucleotide positions of SEQ ID No 1: 1-10000, 10001-20000, 20001-30000, 30001-40000, 40001-50000, 50001-60000, 60001-70000, 70001-80000, 80001-90000, 90001-97921, 98517-103471, 103603-108222, 108390-109221, 109324-114409, 114538-115723, 115957-122102, 122225-126876, 127033-157212, 157808-159000, 159001-160000, 160001-170000, 170001-180000, 180001-190000, 190001-200000, 200001-210000, 210001-220000, 220001-230000, 230001-240825.

Another object of the invention is a purified, isolated, or recombinant nucleic acid comprising the nucleotide sequence of SEQ ID No 2, complementary sequences thereto, as well as allelic variants, and fragments thereof. Moreover, preferred probes and primers of the invention include purified, isolated, or recombinant PG-3 cDNAs consisting of, consisting essentially of, or comprising the sequence of SEQ ID No 2. Particularly preferred probes and primers of the invention include isolated, purified, or recombinant polynucleotides comprising a contiguous span of at least 12, 15, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 500, or 1000 nucleotides of SEQ ID No 2 or the complements thereof. Additional preferred embodiments of the invention include probes and primers comprising a contiguous span of at least 12, 15, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 500, or 1000 nucleotides of SEQ ID No 2 or the complements thereof, wherein said contiguous span comprises at least 1, 2, 3, 5, or 10 of the following nucleotide positions of SEQ ID No 2: 1-500, 501-1000, 1001-1500, 1501-2000, 2001-2500, 2501-3000, 3001-3500, 3501-3809.

Thus, the invention also relates to nucleic acid probes characterized in that they hybridize specifically, under the stringent hybridization conditions defined above, with a nucleic acid selected from the group consisting of the nucleotide sequences 1-97921, 98517-103471, 103603-108222, 108390-109221, 109324-114409, 114538-115723, 115957-122102, 122225-126876, 127033-
5 157212, 157808-240825 of SEQ ID No 1 or a variant thereof or a sequence complementary thereto. The invention relates to nucleic acid probes characterized in that they hybridize specifically, under the stringent hybridization conditions defined above, with a nucleic acid of SEQ ID No 2 or a variant or a fragment thereof or a sequence complementary thereto.

In one embodiment the invention encompasses isolated, purified, and recombinant
10 polynucleotides consisting of, or consisting essentially of a contiguous span of at least 8, 10, 12, 15, 18, 20, 25, 30, 35, 40, or 50 nucleotides in length of any one of SEQ ID Nos 1 and 2 and the complement thereof, wherein said span includes a PG-3-related biallelic marker in said sequence; optionally, said PG-3-related biallelic marker is selected from the group consisting of A1 to A80, and the complements thereof, or optionally the biallelic markers in linkage disequilibrium
15 therewith; optionally, wherein said PG-3-related biallelic marker is selected from the group consisting of A1 to A5 and A8 to A80, and the complements thereof, or optionally the biallelic markers in linkage disequilibrium therewith; optionally, wherein said PG-3-related biallelic marker is selected from the group consisting of A6 and A7, and the complements thereof, or optionally the biallelic markers in linkage disequilibrium therewith; optionally, said contiguous span is 18 to 35
20 nucleotides in length and said biallelic marker is within 4 nucleotides of the center of said polynucleotide; optionally, said polynucleotide comprises, consists essentially of, or consists of said contiguous span and said contiguous span is 25 nucleotides in length and said biallelic marker is at the center of said polynucleotide; optionally, the 3' end of said contiguous span is present at the 3' end of said polynucleotide; and optionally, the 3' end of said contiguous span is located at
25 the 3' end of said polynucleotide and said biallelic marker is present at the 3' end of said polynucleotide. In a preferred embodiment, said probes comprises, consists of, or consists essentially of a sequence selected from the following sequences: P1 to P4 and P6 to P80 and the complementary sequences thereto.

In another embodiment the invention encompasses isolated, purified or recombinant
30 polynucleotides comprising, consisting of, or consisting essentially of a contiguous span of at least 8, 10, 12, 15, 18, 20, 25, 30, 35, 40, or 50 nucleotides in length of SEQ ID Nos 1 and 2, or the complements thereof, wherein the 3' end of said contiguous span is located at the 3' end of said polynucleotide, and wherein the 3' end of said polynucleotide is located within 20 nucleotides upstream of a PG-3-related biallelic marker in said sequence; optionally, wherein said PG-3-related
35 biallelic marker is selected from the group consisting of A1 to A80, and the complements thereof, or optionally the biallelic markers in linkage disequilibrium therewith; optionally, wherein said PG-3-related biallelic marker is selected from the group consisting of A1 to A5 and A8 to A80, and the

complements thereof, or optionally the biallelic markers in linkage disequilibrium therewith; optionally, wherein said PG-3-related biallelic marker is selected from the group consisting of A6 and A7, and the complements thereof, or optionally the biallelic markers in linkage disequilibrium therewith; optionally, wherein the 3' end of said polynucleotide is located 1 nucleotide upstream of
5 said PG-3-related biallelic marker in said sequence; and optionally, wherein said polynucleotide consists essentially of a sequence selected from the following sequences: D1 to D4, D6 to D80, E1 to E4 and E6 to E80.

In a further embodiment, the invention encompasses isolated, purified, or recombinant polynucleotides comprising, consisting of, or consisting essentially of a sequence selected from the
10 following sequences: B1 to B52 and C1 to C52.

In an additional embodiment, the invention encompasses polynucleotides for use in hybridization assays, sequencing assays, and enzyme-based mismatch detection assays for determining the identity of the nucleotide at a PG-3-related biallelic marker in SEQ ID Nos 1 and 2, as well as polynucleotides for use in amplifying segments of nucleotides comprising a PG-3-related
15 biallelic marker in SEQ ID Nos 1 and 2; optionally, wherein said PG-3-related biallelic marker is selected from the group consisting of A1 to A80, and the complements thereof, or optionally the biallelic markers in linkage disequilibrium therewith; optionally, wherein said PG-3-related biallelic marker is selected from the group consisting of A1 to A5 and A8 to A80, and the complements thereof, or optionally the biallelic markers in linkage disequilibrium therewith;
20 optionally, wherein said PG-3-related biallelic marker is selected from the group consisting of A6 and A7, and the complements thereof, or optionally the biallelic markers in linkage disequilibrium therewith.

The invention concerns the use of the polynucleotides according to the invention for determining the identity of the nucleotide at a PG-3-related biallelic marker, preferably in
25 hybridization assay, sequencing assay, microsequencing assay, or an enzyme-based mismatch detection assay and in amplifying segments of nucleotides comprising a PG-3-related biallelic marker.

A probe or a primer according to the invention is between 8 and 1000 nucleotides in length, or is specified to be at least 12, 15, 18, 20, 25, 35, 40, 50, 60, 70, 80, 100, 250, 500 or 1000
30 nucleotides in length. More particularly, the length of these probes and primers can range from 8, 10, 15, 20, or 30 to 100 nucleotides, preferably from 10 to 50, more preferably from 15 to 30 nucleotides. Shorter probes and primers tend to lack specificity for a target nucleic acid sequence and generally require cooler temperatures to form sufficiently stable hybrid complexes with the template. Longer probes and primers are expensive to produce and can sometimes self-hybridize to
35 form hairpin structures. The appropriate length for primers and probes under a particular set of assay conditions may be empirically determined by one of skill in the art. A preferred probe or primer consists of a nucleic acid comprising a polynucleotide selected from the group of the

nucleotide sequences of P1 to P4 and P6 to P80 and the complementary sequence thereto, B1 to B52, C1 to C52, D1 to D4, D6 to D80, E1 to E4 and E6 to E80, for which the respective locations in the sequence listing are provided in Tables 1, 2, and 3.

The formation of stable hybrids depends on the melting temperature (T_m) of the DNA. The T_m depends on the length of the primer or probe, the ionic strength of the solution and the G+C content. The higher the G+C content of the primer or probe, the higher is the melting temperature because G:C pairs are held by three H bonds whereas A:T pairs have only two. The GC content in the probes of the invention usually ranges between 10 and 75 %, preferably between 35 and 60 %, and more preferably between 40 and 55 %.

The primers and probes can be prepared by any suitable method, including, for example, cloning and restriction of appropriate sequences and direct chemical synthesis by a method such as the phosphodiester method of Narang *et al.* (1979), the phosphodiester method of Brown *et al.* (1979), the diethylphosphoramidite method of Beaucage *et al.* (1981) and the solid support method described in EP 0 707 592.

Detection probes are generally nucleic acid sequences or uncharged nucleic acid analogs such as, for example peptide nucleic acids which are disclosed in International Patent Application WO 92/20702, morpholino analogs which are described in U.S. Patents Numbered 5,185,444; 5,034,506 and 5,142,047. The probe may have to be rendered "non-extendable" in that additional dNTPs cannot be added to the probe. In and of themselves analogs usually are non-extendable and nucleic acid probes can be rendered non-extendable by modifying the 3' end of the probe such that the hydroxyl group is no longer capable of participating in elongation. For example, the 3' end of the probe can be functionalized with the capture or detection label to thereby consume or otherwise block the hydroxyl group. Alternatively, the 3' hydroxyl group simply can be cleaved, replaced or modified, U.S. Patent Application Serial No. 07/049,061 filed April 19, 1993 describes modifications, which can be used to render a probe non-extendable.

Any of the polynucleotides of the present invention can be labeled, if desired, by incorporating any label known in the art to be detectable by spectroscopic, photochemical, biochemical, immunochemical, or chemical means. For example, useful labels include radioactive substances (including, ^{32}P , ^{35}S , ^3H , ^{125}I), fluorescent dyes (including, 5-bromodesoxyuridin, fluorescein, acetylaminofluorene, digoxigenin) or biotin. Preferably, polynucleotides are labeled at their 3' and 5' ends. Examples of non-radioactive labeling of nucleic acid fragments are described in the French patent No. FR-7810975, or by Urdea *et al.* (1988) or Sanchez-Pescador *et al.* (1988). In addition, the probes according to the present invention may have structural characteristics such that they allow the signal amplification, such structural characteristics being, for example, branched DNA probes as those described by Urdea *et al.* in 1991 or in the European patent No. EP 0 225 807 (Chiron).

A label can also be used to capture the primer, so as to facilitate the immobilization of either the primer or a primer extension product, such as amplified DNA, on a solid support. A capture label is attached to the primers or probes and can be a specific binding member which forms a binding pair with the solid's phase reagent's specific binding member (e.g. biotin and streptavidin). Therefore depending upon the type of label carried by a polynucleotide or a probe, it may be employed to capture or to detect the target DNA. Further, it will be understood that the polynucleotides, primers or probes provided herein, may, themselves, serve as the capture label. For example, in the case where a solid phase reagent's binding member is a nucleic acid sequence, it may be selected such that it binds a complementary portion of a primer or probe to thereby immobilize the primer or probe to the solid phase. In cases where a polynucleotide probe itself serves as the binding member, those skilled in the art will recognize that the probe will contain a sequence or "tail" that is not complementary to the target. In the case where a polynucleotide primer itself serves as the capture label, at least a portion of the primer will be free to hybridize with a nucleic acid on a solid phase. DNA Labeling techniques are well known to the skilled technician.

The probes of the present invention are useful for a number of purposes. They can be notably used in Southern hybridization to genomic DNA. The probes can also be used to detect PCR amplification products. They may also be used to detect mismatches in the PG-3 gene or mRNA using other techniques.

Any of the polynucleotides, primers and probes of the present invention can be conveniently immobilized on a solid support. Solid supports are known to those skilled in the art and include the walls of wells of a reaction tray, test tubes, polystyrene beads, magnetic beads, nitrocellulose strips, membranes, microparticles such as latex particles, sheep (or other animal) red blood cells, duracytes and others. The solid support is not critical and can be selected by one skilled in the art. Thus, latex particles, microparticles, magnetic or non-magnetic beads, membranes, plastic tubes, walls of microtiter wells, glass or silicon chips, sheep (or other suitable animal's) red blood cells and duracytes are all suitable examples. Suitable methods for immobilizing nucleic acids on solid phases include ionic, hydrophobic, covalent interactions and the like. A solid support, as used herein, refers to any material which is insoluble, or can be made insoluble by a subsequent reaction. The solid support can be chosen for its intrinsic ability to attract and immobilize the capture reagent. Alternatively, the solid phase can retain an additional receptor which has the ability to attract and immobilize the capture reagent. The additional receptor can include a charged substance that is oppositely charged with respect to the capture reagent itself or to a charged substance conjugated to the capture reagent. As yet another alternative, the receptor molecule can be any specific binding member which is immobilized upon (attached to) the solid support and which has the ability to immobilize the capture reagent through a specific binding reaction. The receptor molecule enables the indirect binding of the capture reagent to a solid support material before the performance of the assay or during the performance of the assay. The

allowing the amplification of a nucleic acid containing the polymorphic base of one PG-3 biallelic marker are listed in Table 1 of Example 2.

Eight PG-3-related biallelic markers A3, A6, A7, A14, A70, A71, A72 and A80, are located in the exonic regions of the genomic sequence of PG-3 at the following positions: 10228, 39944, 39973, 76060, 216026, 216082, 216218 and 237555 of the SEQ ID No 1. They are located in exons C, T, I, K and L of the PG-3 gene. Their respective positions in the cDNA and protein sequences are given in Table 2.

The invention also relates to a purified and/or isolated nucleotide sequence comprising a polymorphic base of a PG-3-related biallelic marker, preferably of a biallelic marker selected from the group consisting of A1 to A80, and the complements thereof. The sequence is between 8 and 1000 nucleotides in length, and preferably comprises at least 8, 10, 12, 15, 18, 20, 25, 35, 40, 50, 60, 70, 80, 100, 250, 500 or 1000 contiguous nucleotides of a nucleotide sequence selected from the group consisting of SEQ ID Nos 1 and 2 or a variant thereof or a complementary sequence thereto. These nucleotide sequences comprise the polymorphic base of either allele 1 or allele 2 of the considered biallelic marker. Optionally, said biallelic marker may be within 6, 5, 4, 3, 2, or 1 nucleotides of the center of said polynucleotide or at the center of said polynucleotide. Optionally, the 3' end of said contiguous span may be present at the 3' end of said polynucleotide. Optionally, biallelic marker may be present at the 3' end of said polynucleotide. Optionally, said polynucleotide may further comprise a label. Optionally, said polynucleotide can be attached to solid support. In a further embodiment, the polynucleotides defined above can be used alone or in any combination.

The invention also relates to a purified and/or isolated nucleotide sequence comprising a sequence between 8 and 1000 nucleotides in length, and preferably at least 8, 10, 12, 15, 18, 20, 25, 35, 40, 50, 60, 70, 80, 100, 250, 500 or 1000 contiguous nucleotides of a nucleotide sequence selected from the group consisting of SEQ ID Nos 1 and 2 or a variant thereof or a complementary sequence thereto. Optionally, the 3' end of said polynucleotide may be located within or at least 2, 4, 6, 8, 10, 12, 15, 18, 20, 25, 50, 100, 250, 500, or 1000 nucleotides upstream of a PG-3-related biallelic marker in said sequence. Optionally, said PG-3-related biallelic marker is selected from the group consisting of A1 to A80; Optionally, the 3' end of said polynucleotide may be located 1 nucleotide upstream of a PG-3-related biallelic marker in said sequence. Optionally, said polynucleotide may further comprise a label. Optionally, said polynucleotide can be attached to solid support. In a further embodiment, the polynucleotides defined above can be used alone or in any combination.

In a preferred embodiment, the sequences comprising a polymorphic base of one of the biallelic markers listed in Table 2 are selected from the group consisting of the nucleotide sequences comprising, consisting essentially of, or consisting of the amplicons listed in Table 1 or a variant thereof or a complementary sequence thereto.

solid phase thus can be a plastic, derivatized plastic, magnetic or non-magnetic metal, glass or silicon surface of a test tube, microtiter well, sheet, bead, microparticle, chip, sheep (or other suitable animal's) red blood cells, duracytes® and other configurations known to those of ordinary skill in the art. The polynucleotides of the invention can be attached to or immobilized on a solid support individually or in groups of at least 2, 5, 8, 10, 12, 15, 20, or 25 distinct polynucleotides of the invention to a single solid support. In addition, polynucleotides other than those of the invention may be attached to the same solid support as one or more polynucleotides of the invention.

Consequently, the invention also relates to a method for detecting the presence of a nucleic acid comprising a nucleotide sequence selected from the group consisting of SEQ ID Nos 1 and 2, a fragment or a variant thereof and a complementary sequence thereto in a sample, said method comprising the following steps of:

- a) bringing into contact a nucleic acid probe or a plurality of nucleic acid probes which can hybridize with a nucleotide sequence included in a nucleic acid selected from the group consisting of the nucleotide sequences of SEQ ID Nos 1 and 2, a fragment or a variant thereof and a complementary sequence thereto and the sample to be assayed; and
- b) detecting the hybrid complex formed between the probe and a nucleic acid in the sample.

The invention further concerns a kit for detecting the presence of a nucleic acid comprising a nucleotide sequence selected from a group consisting of SEQ ID Nos 1 and 2, a fragment or a variant thereof and a complementary sequence thereto in a sample, said kit comprising:

- a) a nucleic acid probe or a plurality of nucleic acid probes which can hybridize with a nucleotide sequence included in a nucleic acid selected from the group consisting of the nucleotide sequences of SEQ ID Nos 1 and 2, a fragment or a variant thereof and a complementary sequence thereto; and
- b) optionally, the reagents necessary for performing the hybridization reaction.

In a first preferred embodiment of this detection method and kit, said nucleic acid probe or the plurality of nucleic acid probes are labeled with a detectable molecule. In a second preferred embodiment of said method and kit, said nucleic acid probe or the plurality of nucleic acid probes has been immobilized on a substrate. In a third preferred embodiment, the nucleic acid probe or the plurality of nucleic acid probes comprise either a sequence which is selected from the group consisting of the nucleotide sequences of P1 to P4 and P6 to P80 and the complementary sequence thereto, B1 to B52, C1 to C52, D1 to D4, D6 to D80, E1 to E4 and E6 to E80 or a biallelic marker selected from the group consisting of A1 to A80 and the complements thereto.

Olig nucleotide Arrays

A substrate comprising a plurality of oligonucleotide primers or probes of the invention may be used either for detecting or amplifying targeted sequences in the PG-3 gene and may also be used for detecting mutations in the coding or in the non-coding sequences of the PG-3 gene.

Any polynucleotide provided herein may be attached in overlapping areas or at random locations on the solid support. Alternatively, the polynucleotides of the invention may be attached in an ordered array wherein each polynucleotide is attached to a distinct region of the solid support which does not overlap with the attachment site of any other polynucleotide. Preferably, such an ordered array of polynucleotides is designed to be "addressable" where the distinct locations are recorded and can be accessed as part of an assay procedure. Addressable polynucleotide arrays typically comprise a plurality of different oligonucleotide probes that are coupled to a surface of a substrate in different known locations. The knowledge of the precise location of each polynucleotide makes these "addressable" arrays particularly useful in hybridization assays. Any addressable array technology known in the art can be employed with the polynucleotides of the invention. One particular embodiment of these polynucleotide arrays is known as the Genechips™, and has been generally described in US Patent 5,143,854; PCT publications WO 90/15070 and 92/10092. These arrays may generally be produced using mechanical synthesis methods or light directed synthesis methods which incorporate a combination of photolithographic methods and solid phase oligonucleotide synthesis (Fodor *et al.*, 1991). The immobilization of arrays of oligonucleotides on solid supports has been rendered possible by the development of a technology generally identified as "Very Large Scale Immobilized Polymer Synthesis" (VLSIPS™) in which, typically, probes are immobilized in a high density array on a solid surface of a chip. Examples of VLSIPS™ technologies are provided in US Patents 5,143,854; and 5,412,087 and in PCT Publications WO 90/15070, WO 92/10092 and WO 95/11995, which describe methods for forming oligonucleotide arrays through techniques such as light-directed synthesis techniques. In designing strategies aimed at providing arrays of nucleotides immobilized on solid supports, further presentation strategies were developed to order and display the oligonucleotide arrays on the chips in an attempt to maximize hybridization patterns and sequence information. Examples of such presentation strategies are disclosed in PCT Publications WO 94/12305, WO 94/11530, WO 97/29212 and WO 97/31256.

In another embodiment of the oligonucleotide arrays of the invention, an oligonucleotide probe matrix may advantageously be used to detect mutations occurring in the PG-3 gene and preferably in its regulatory region. For this particular purpose, probes are specifically designed to have a nucleotide sequence allowing their hybridization to the genes that carry known mutations (either by deletion, insertion or substitution of one or several nucleotides). By known mutations, it is meant, mutations on the PG-3 gene that have been identified according, for example to the technique used by Huang *et al.*(1996) or Samson *et al.*(1996).

Another technique that may be used to detect mutations in the PG-3 gene is the use of a high-density DNA array. Each oligonucleotide probe constituting a unit element of the high density DNA array is designed to match a specific subsequence of the PG-3 genomic DNA or cDNA.

Thus, an array consisting of oligonucleotides complementary to subsequences of the target gene
5 sequence is used to determine the identity of the target sequence within a sample, measure its amount, and detect differences between the target sequence and the sequence of the PG-3 gene in the sample. In one such design, termed 4L tiled array, a set of four probes (A, C, G, T), preferably 15-nucleotide oligomers, is used. In each set of four probes, the perfect complement will hybridize more strongly than mismatched probes. Consequently, a nucleic acid target of length L is scanned
10 for mutations with a tiled array containing 4L probes, the whole probe set containing all the possible mutations in the known sequence. The hybridization signals of the 15-mer probe set tiled array are perturbed by a single base change in the target sequence. As a consequence, there is a characteristic loss of signal or a "footprint" for the probes flanking a mutation position. This technique was described by Chee *et al.* in 1996.

15 Consequently, the invention concerns an array of nucleic acid molecules comprising at least one polynucleotide described above as probes and primers. Preferably, the invention concerns an array of nucleic acid comprising at least two polynucleotides described above as probes and primers.

A further object of the invention consists of an array of nucleic acid sequences comprising
20 either at least one of the sequences selected from the group consisting of P1 to P4 and P6 to P80, B1 to B52, C1 to C52, D1 to D4, D6 to D80, E1 to E4 and E6 to E80, the sequences complementary thereto, a fragment thereof of at least 8, 10, 12, 15, 18, or 20 consecutive nucleotides thereof, or at least one sequence comprising a biallelic marker selected from the group consisting of A1 to A80 and the complements thereto.

25 The invention also pertains to an array of nucleic acid sequences comprising either at least two of the sequences selected from the group consisting of P1 to P4, P6 to P80, B1 to B52, C1 to C52, D1 to D4, D6 to D80, E1 to E4 and E6 to E80, the sequences complementary thereto, a fragment thereof of at least 8 consecutive nucleotides thereof, or at least two sequences comprising a biallelic marker selected from the group consisting of A1 to A80 and the complements thereof.

30 PG-3 PROTEINS AND POLYPEPTIDE FRAGMENTS

The term "PG-3 polypeptides" is used herein to embrace all of the proteins and polypeptides of the present invention. Also forming part of the invention are polypeptides encoded by the polynucleotides of the invention, as well as fusion polypeptides comprising such polypeptides. The invention embodies PG-3 proteins from humans, including isolated or purified
35 PG-3 proteins consisting, consisting essentially, or comprising the sequence of SEQ ID No 3. More particularly, the present invention concerns allelic variants of the PG-3 protein comprising at least one amino acid selected from the group consisting of an arginine or an isoleucine residue at the

amino acid position 304 of the SEQ ID No 3, a histidine or an aspartic acid residue at the amino acid position 314 of the SEQ ID No 3, a threonine or an asparagine residue at the amino acid position 682 of the SEQ ID No 3, an alanine or a valine residue at the amino acid position 761 of the SEQ ID No 3, and a proline or a serine residue at the amino acid position 828 of the SEQ ID No 3.

5 3. In addition, the invention also encompasses polypeptide variants of PG-3 comprising at least one amino acid selected from the group consisting of a methionine or an isoleucine residue at the position 91 of SEQ ID No 3, a valine or an alanine residue at the position 306 of SEQ ID No 3, a proline or a serine residue at the position 413 of SEQ ID No 3, a glycine or an aspartate residue at the position 528 of SEQ ID No 3, a valine or an alanine residue at the position 614 of SEQ ID No 3, 10 a threonine or an asparagine residue at the position 677 of SEQ ID No 3, a valine or an alanine residue at the position 756 of SEQ ID No 3, a valine or an alanine residue at the position 758 of SEQ ID No 3, a lysine or a glutamate residue at the position 809 of SEQ ID No 3, and a cysteine or an arginine residue at the position 821 of SEQ ID No 3.

The present invention includes isolated, purified, or recombinant polypeptides comprising a 15 contiguous span of at least 6 amino acids, preferably at least 8 to 10 amino acids, more preferably at least 12, 15, 20, 25, 30, 40, 50, or 100 amino acids of SEQ ID No 3. The present invention also embodies isolated, purified, and recombinant polypeptides comprising a contiguous span of at least 6 amino acids, preferably at least 8 to 10 amino acids, more preferably at least 12, 15, 20, 25, 30, 40, 50, or 100 amino acids of SEQ ID No 3, wherein said contiguous span includes at least 1, 2, 3, 5 20 or 10 of the following amino acid positions of SEQ ID No 3: 1-100, 101-200, 201-300, 301-400, 401-500, 501-600, 601-700, 701-835. In other preferred embodiments the contiguous stretch of amino acids comprises the site of a mutation or functional mutation, including a deletion, addition, swap or truncation of the amino acids in the PG-3 protein sequence.

The invention also encompasses purified, isolated, or recombinant polypeptides comprising 25 a sequence having at least 70, 75, 80, 85, 90, 95, 98 or 99% nucleotide identity with the sequence of SEQ ID No 3 or a fragment thereof.

PG-3 proteins are preferably isolated from human or mammalian tissue samples or expressed from human or mammalian genes. The PG-3 polypeptides of the invention can be made using routine expression methods known in the art. The polynucleotide encoding the desired 30 polypeptide, is ligated into an expression vector suitable for any convenient host. Both eukaryotic and prokaryotic host systems is used in forming recombinant polypeptides, and a summary of some of the more common systems. The polypeptide is then isolated from lysed cells or from the culture medium and purified to the extent needed for its intended use. Purification is by any technique known in the art, for example, differential extraction, salt fractionation, chromatography, 35 centrifugation, and the like. See, for example, Methods in Enzymology for a variety of methods for purifying proteins.

In addition, shorter protein fragments is produced by chemical synthesis. Alternatively the proteins of the invention is extracted from cells or tissues of humans or non-human animals. Methods for purifying proteins are known in the art, and include the use of detergents or chaotropic agents to disrupt particles followed by differential extraction and separation of the polypeptides by ion exchange chromatography, affinity chromatography, sedimentation according to density, and gel electrophoresis.

Any PG-3 cDNA, including SEQ ID No 2, may be used to express PG-3 proteins and polypeptides. The nucleic acid encoding the PG-3 protein or polypeptide to be expressed is operably linked to a promoter in an expression vector using conventional cloning technology. The PG-3 insert in the expression vector may comprise the full coding sequence for the PG-3 protein or a portion thereof. For example, the PG-3 derived insert may encode a polypeptide comprising at least 10 consecutive amino acids of the PG-3 protein of SEQ ID No 3, preferably least 10 consecutive amino acids including at least 1, 2, 3, 5 or 10 of the following amino acid positions of SEQ ID No 3: 1-100, 101-200, 201-300, 301-400, 401-500, 501-600, 601-700, 701-835.

The expression vector may be any of the mammalian, yeast, insect or bacterial expression systems known in the art. Commercially available vectors and expression systems are available from a variety of suppliers including Genetics Institute (Cambridge, MA), Stratagene (La Jolla, California), Promega (Madison, Wisconsin), and Invitrogen (San Diego, California). If desired, to enhance expression and facilitate proper protein folding, the codon context and codon pairing of the sequence may be optimized for the particular expression organism in which the expression vector is introduced, as explained by Hatfield, *et al.*, and U.S. Patent No. 5,082,767.

In one embodiment, the entire coding sequence of the PG-3 cDNA through the poly A signal of the cDNA is operably linked to a promoter in the expression vector. Alternatively, if the nucleic acid encoding a portion of the PG-3 protein lacks a methionine to serve as the initiation site, an initiating methionine can be introduced next to the first codon of the nucleic acid using conventional techniques. Similarly, if the insert from the PG-3 cDNA lacks a poly A signal, this sequence can be added to the construct by, for example, splicing out the Poly A signal from pSG5 (Stratagene) using BglII and SalI restriction endonuclease enzymes and incorporating it into the mammalian expression vector pXT1 (Stratagene). pXT1 contains the LTRs and a portion of the gag gene from Moloney Murine Leukemia Virus. The position of the LTRs in the construct allow efficient stable transfection. The vector includes the Herpes Simplex Thymidine Kinase promoter and the selectable neomycin gene. The nucleic acid encoding the PG-3 protein or a portion thereof is obtained by PCR from a bacterial vector containing the PG-3 cDNA of SEQ ID No 3 using oligonucleotide primers complementary to the PG-3 cDNA or portion thereof and containing restriction endonuclease sequences for Pst I incorporated into the 5' primer and BglII at the 5' end of the corresponding cDNA 3' primer, taking care to ensure that the sequence encoding the PG-3 protein or a portion thereof is positioned properly with respect to the poly A signal. The purified fragment obtained from the

resulting PCR reaction is digested with PstI, blunt ended with an exonuclease, digested with Bgl II, purified and ligated to pXT1, now containing a poly A signal and digested with BglII.

The ligated product is transfected into mouse NIH 3T3 cells using Lipofectin (Life Technologies, Inc., Grand Island, New York) under conditions outlined in the product specification.

- 5 Positive transfectants are selected after growing the transfected cells in 600ug/ml G418 (Sigma, St. Louis, Missouri).

The above procedures may also be used to express a mutant PG-3 protein responsible for a detectable phenotype or a portion thereof.

- The expressed protein is purified using conventional purification techniques such as
10 ammonium sulfate precipitation or chromatographic separation based on size or charge. The protein encoded by the nucleic acid insert may also be purified using standard immunochromatography techniques. In such procedures, a solution containing the expressed PG-3 protein or portion thereof, such as a cell extract, is applied to a column having antibodies against the PG-3 protein or portion thereof attached to the chromatography matrix. The expressed protein is allowed to bind the
15 immunochromatography column. Thereafter, the column is washed to remove non-specifically bound proteins. The specifically bound expressed protein is then released from the column and recovered using standard techniques.

- To confirm expression of the PG-3 protein or a portion thereof, the proteins expressed from host cells containing an expression vector containing an insert encoding the PG-3 protein or a portion
20 thereof can be compared to the proteins expressed in host cells containing the expression vector without an insert. The presence of a band in samples from cells containing the expression vector with an insert which is absent in samples from cells containing the expression vector without an insert indicates that the PG-3 protein or a portion thereof is being expressed. Generally, the band will have the mobility expected for the PG-3 protein or portion thereof. However, the band may have a mobility different
25 than that expected as a result of modifications such as glycosylation, ubiquitination, or enzymatic cleavage.

Antibodies capable of specifically recognizing the expressed PG-3 protein or a portion thereof are described below.

- If antibody production is not possible, the nucleic acids encoding the PG-3 protein or a portion
30 thereof is incorporated into expression vectors designed for use in purification schemes employing chimeric polypeptides. In such strategies the nucleic acid encoding the PG-3 protein or a portion thereof is inserted in frame with the gene encoding the other half of the chimera. The other half of the chimera is β -globin or a nickel binding polypeptide encoding sequence. A chromatography matrix having antibody to β -globin or nickel attached thereto is then used to purify the chimeric protein.
35 Protease cleavage sites are engineered between the β -globin gene or the nickel binding polypeptide and the PG-3 protein or portion thereof. Thus, the two polypeptides of the chimera is separated from one another by protease digestion.

One useful expression vector for generating β -globin chimeric proteins is pSG5 (Stratagene), which encodes rabbit β -globin. Intron II of the rabbit β -globin gene facilitates splicing of the expressed transcript, and the polyadenylation signal incorporated into the construct increases the level of expression. These techniques are well known to those skilled in the art of molecular biology. Standard methods are published in methods texts such as Davis *et al.*, (1986) and many of the methods are available from Stratagene, Life Technologies, Inc., or Promega. Polypeptide may additionally be produced from the construct using in vitro translation systems such as the In vitro ExpressTM Translation Kit (Stratagene).

ANTIBODIES THAT BIND PG-3 POLYPEPTIDES OF THE INVENTION

Any PG-3 polypeptide or whole protein may be used to generate antibodies capable of specifically binding to an expressed PG-3 protein or fragments thereof as described.

One antibody composition of the invention is capable of specifically binding to the PG-3 protein of SEQ ID No 3. For an antibody composition to specifically bind to the PG-3 protein, it must demonstrate at least a 5%, 10%, 15%, 20%, 25%, 50%, or 100% greater binding affinity for PG-3 protein than for another protein in an ELISA, RIA, or other antibody-based binding assay.

The invention also concerns antibody compositions which are specific for variants of the PG-3 protein, more particularly variants comprising at least one amino acid selected from the group consisting of a methionine or an isoleucine residue at the position 91 of SEQ ID No 3, a valine or an alanine residue at the position 306 of SEQ ID No 3, a proline or a serine residue at the position 413 of SEQ ID No 3, a glycine or an aspartate residue at the position 528 of SEQ ID No 3, a valine or an alanine residue at the position 614 of SEQ ID No 3, a threonine or an asparagine residue at the position 677 of SEQ ID No 3, a valine or an alanine residue at the position 756 of SEQ ID No 3, a valine or an alanine residue at the position 758 of SEQ ID No 3, a lysine or a glutamate residue at the position 809 of SEQ ID No 3, and a cysteine or an arginine residue at the position 821 of SEQ ID No 3. More preferably, the invention encompasses antibody compositions which are specific for an allelic variant of the PG-3 protein, more particularly a variant comprising at least one amino acid selected from the group consisting of an arginine or an isoleucine residue at the amino acid position 304 of SEQ ID No 3, a histidine or an aspartic acid residue at the amino acid position 314 of SEQ ID No 3, a threonine or an asparagine residue at the amino acid position 682 of SEQ ID No 3, an alanine or a valine residue at the amino acid position 761 of SEQ ID No 3, and a proline or a serine residue at the amino acid position 828 of SEQ ID No 3.

In a preferred embodiment, the invention concerns antibody compositions, either polyclonal or monoclonal, capable of selectively binding, or selectively bind to an epitope-containing a polypeptide comprising a contiguous span of at least 6 amino acids, preferably at least 8 to 10 amino acids, more preferably at least 12, 15, 20, 25, 30, 40, 50, or 100 amino acids of SEQ ID No 3; preferably, said epitope comprises at least 1, 2, 3, 5 or 10 of the following amino acid positions of SEQ ID No 3: 1-100, 101-200, 201-300, 301-400, 401-500, 501-600, 601-700, 701-835.

The invention also concerns a purified or isolated antibody capable of specifically binding to a mutated PG-3 protein or to a fragment or variant thereof comprising an epitope of the mutated PG-3 protein. In another preferred embodiment, the present invention concerns an antibody capable of binding to a polypeptide comprising at least 10 consecutive amino acids of a PG-3 protein and including at least one of the amino acids which can be encoded by the trait causing mutations.

In a preferred embodiment, the invention concerns the use in the manufacture of antibodies of a polypeptide comprising a contiguous span of at least 6 amino acids, preferably at least 8 to 10 amino acids, more preferably at least 12, 15, 20, 25, 30, 40, 50, or 100 amino acids of SEQ ID No 3; preferably, said contiguous span comprises at least 1, 2, 3, 5 or 10 of the following amino acid positions of SEQ ID No 3: 1-100, 101-200, 201-300, 301-400, 401-500, 501-600, 601-700, 701-835.

Non-human animals or mammals, whether wild-type or transgenic, which express a different species of PG-3 than the one to which antibody binding is desired, and animals which do not express PG-3 (*i.e.* a PG-3 knock out animal as described herein) are particularly useful for preparing antibodies. PG-3 knock out animals will recognize all or most of the exposed regions of a PG-3 protein as foreign antigens, and therefore produce antibodies with a wider array of PG-3 epitopes. Moreover, smaller polypeptides with only 10 to 30 amino acids may be useful in obtaining specific binding to any one of the PG-3 proteins. In addition, the humoral immune system of animals which produce a species of PG-3 that resembles the antigenic sequence will preferentially recognize the differences between the animal's native PG-3 species and the antigen sequence, and produce antibodies to these unique sites in the antigen sequence. Such a technique will be particularly useful in obtaining antibodies that specifically bind to any one of the PG-3 proteins.

Antibody preparations prepared according to either protocol are useful in quantitative immunoassays which determine concentrations of antigen-bearing substances in biological samples; they are also used semi-quantitatively or qualitatively to identify the presence of antigen in a biological sample. The antibodies may also be used in therapeutic compositions for killing cells expressing the protein or reducing the levels of the protein in the body.

The antibodies of the invention may be labeled using any one of the radioactive, fluorescent or enzymatic labels known in the art.

Consequently, the invention is also directed to a method for specifically detecting the presence of a PG-3 polypeptide according to the invention in a biological sample, said method comprising the following steps :

- a) bringing the biological sample into contact with a polyclonal or monoclonal antibody that specifically binds to a PG-3 polypeptide comprising an amino acid sequence of SEQ ID No 3, or to a peptide fragment or variant thereof; and
- b) detecting the antigen-antibody complex formed.

The invention also concerns a diagnostic kit for detecting the presence of a PG-3 polypeptide according to the present invention in a biological sample *in vitro*, wherein said kit comprises:

- a) a polyclonal or monoclonal antibody that specifically binds to a PG-3 polypeptide comprising the amino acid sequence of SEQ ID No 3, or to a peptide fragment or variant thereof; optionally the antibody may be labeled; and
- b) a reagent allowing the detection of the antigen-antibody complexes formed, said reagent optionally carrying a label, or being able to be recognized itself by a labeled reagent (particularly in the case when the above-mentioned monoclonal or polyclonal antibody itself is not labeled).

PG-3 -RELATED BIALLELIC MARKERS

Advantages Of The Biallelic Markers Of The Present Invention

The PG-3-related biallelic markers of the present invention offer a number of important advantages over other genetic markers such as RFLP (Restriction fragment length polymorphism) and VNTR (Variable Number of Tandem Repeats) markers.

The first generation of markers were RFLPs, which are variations that modify the length of a restriction fragment. But methods used to identify and to type RFLPs are relatively wasteful of materials, effort, and time. The second generation of genetic markers were VNTRs, which can be categorized as either minisatellites or microsatellites. Minisatellites are tandemly repeated DNA sequences present in units of 5-50 repeats which are distributed along regions of the human chromosomes ranging from 0.1 to 20 kilobases in length. Since they present many possible alleles, their informative content is very high. Minisatellites are scored by performing Southern blots to identify the number of tandem repeats present in a nucleic acid sample from the individual being tested. However, there are only 10^4 potential VNTRs that can be typed by Southern blotting. Moreover, both RFLP and VNTR markers are costly and time-consuming to develop and assay in large numbers.

Single nucleotide polymorphisms (SNPs) or biallelic markers can be used in the same manner as RFLPs and VNTRs but offer several advantages. SNPs are densely spaced in the human genome and represent the most frequent type of variation. An estimated number of more than 10^7 sites are scattered along the 3×10^9 base pairs of the human genome. Therefore, SNPs occur at a greater frequency and with greater uniformity than RFLP or VNTR markers which means that there is a greater probability that such a marker will be found in close proximity to a genetic locus of interest. SNPs are less variable than VNTR markers but are mutationally more stable.

Also, the different forms of a characterized single nucleotide polymorphism, such as the biallelic markers of the present invention, are often easier to distinguish and can therefore be typed easily on a routine basis. Biallelic markers have single nucleotide based alleles and they have only two common alleles, which allows highly parallel detection and automated scoring. The biallelic

markers of the present invention offer the possibility of rapid, high throughput genotyping of a large number of individuals.

Biallelic markers are densely spaced in the genome, sufficiently informative and can be assayed in large numbers. The combined effects of these advantages make biallelic markers
5 extremely valuable in genetic studies. Biallelic markers can be used in linkage studies in families, in allele sharing methods, in linkage disequilibrium studies in populations, in association studies of case-control populations or of trait positive and trait negative populations. An important aspect of the present invention is that biallelic markers allow association studies to be performed to identify genes involved in complex traits. Association studies examine the frequency of marker alleles in
10 unrelated case- and control-populations and are generally employed in the detection of polygenic or sporadic traits. Association studies may be conducted within the general population and are not limited to studies performed on related individuals in affected families (linkage studies). Biallelic markers in different genes can be screened in parallel for direct association with disease or response to a treatment. This multiple gene approach is a powerful tool for a variety of human genetic
15 studies as it provides the necessary statistical power to examine the synergistic effect of multiple genetic factors on a particular phenotype, drug response, sporadic trait, or disease state with a complex genetic etiology.

Candidate Gene Of The Present Invention

Different approaches can be employed to perform association studies: genome-wide
20 association studies, candidate region association studies and candidate gene association studies. Genome-wide association studies rely on the screening of genetic markers evenly spaced and covering the entire genome. The candidate gene approach is based on the study of genetic markers specifically located in genes potentially involved in a biological pathway related to the trait of interest. In the present invention, PG-3 is a good candidate gene for cancer. The candidate gene
25 analysis clearly provides a short-cut approach to the identification of genes and gene polymorphisms related to a particular trait when some information concerning the biology of the trait is available. However, it should be noted that all of the biallelic markers disclosed in the instant application can be employed as part of genome-wide association studies or as part of candidate region association studies and such uses are specifically contemplated in the present
30 invention and claims.

PG-3-Related Biallelic Markers And Polynucleotides Related Thereto

The invention also concerns PG-3-related biallelic markers. As used herein the term "PG-3-related biallelic marker" relates to a set of biallelic markers in linkage disequilibrium with the PG-3 gene. The term PG-3-related biallelic marker includes the biallelic markers designated A1 to A80.
35 A portion of the biallelic markers of the present invention are disclosed in Table 2. Their locations in the PG-3 gene are indicated in Table 2 and also as a single base polymorphism in the features of SEQ ID Nos 1 and 2 listed in the accompanying Sequence Listing. The pairs of primers

The invention further concerns a nucleic acid encoding the PG-3 protein, wherein said nucleic acid comprises a polymorphic base of a biallelic marker selected from the group consisting of A1 to A80 and the complements thereof.

The invention also encompasses the use of any polynucleotide for, or any polynucleotide
5 for use in, determining the identity of one or more nucleotides at a PG-3-related biallelic marker. In addition, the polynucleotides of the invention for use in determining the identity of one or more nucleotides at a PG-3-related biallelic marker encompass polynucleotides with any further limitation described in this disclosure, or those following, specified alone or in any combination. Optionally, said PG-3-related biallelic marker is selected from the group consisting of A1 to A80,
10 and the complements thereof, or optionally the biallelic markers in linkage disequilibrium therewith; optionally, said PG-3-related biallelic marker is selected from the group consisting of A1 to A5 and A8 to A80, and the complements thereof, or optionally the biallelic markers in linkage disequilibrium therewith; optionally, said PG-3-related biallelic marker is selected from the group consisting A6 and A7, and the complements thereof, or optionally the biallelic markers in linkage
15 disequilibrium therewith; Optionally, said polynucleotide may comprise a sequence disclosed in the present specification; Optionally, said polynucleotide may comprise, consist of, or consist essentially of any polynucleotide described in the present specification; Optionally, said determining may involve a hybridization assay, sequencing assay, microsequencing assay, or an enzyme-based mismatch detection assay; Optionally, said polynucleotide may be attached to a
20 solid support, array, or addressable array; Optionally, said polynucleotide may be labeled. A preferred polynucleotide may be used in a hybridization assay for determining the identity of the nucleotide at a PG-3-related biallelic marker. Another preferred polynucleotide may be used in a sequencing or microsequencing assay for determining the identity of the nucleotide at a PG-3-related biallelic marker. A third preferred polynucleotide may be used in an enzyme-based
25 mismatch detection assay for determining the identity of the nucleotide at a PG-3-related biallelic marker. A fourth preferred polynucleotide may be used in amplifying a segment of polynucleotides comprising a PG-3-related biallelic marker. Optionally, any of the polynucleotides described above may be attached to a solid support, array, or addressable array; Optionally, said polynucleotide may be labeled.

30 Additionally, the invention encompasses the use of any polynucleotide for, or any polynucleotide for use in amplifying a segment of nucleotides comprising a PG-3-related biallelic marker. In addition, the polynucleotides of the invention for use in amplifying a segment of nucleotides comprising a PG-3-related biallelic marker encompass polynucleotides with any further limitation described in this disclosure, or those following, specified alone or in any combination:
35 Optionally, said PG-3-related biallelic marker is selected from the group consisting of A1 to A80, and the complements thereof, or optionally the biallelic markers in linkage disequilibrium therewith; optionally, said PG-3-related biallelic marker is selected from the group consisting of A1

to A5 and A8 to A80, and the complements thereof, or optionally the biallelic markers in linkage disequilibrium therewith; optionally, said PG-3-related biallelic marker is selected from the group consisting A6 and A7, and the complements thereof, or optionally the biallelic markers in linkage disequilibrium therewith; Optionally, said polynucleotide may comprise a sequence disclosed in the
5 present specification; Optionally, said polynucleotide may comprise, consist of, or consist essentially of any polynucleotide described in the present specification; Optionally, said amplifying may involve PCR or LCR. Optionally, said polynucleotide may be attached to a solid support, array, or addressable array. Optionally, said polynucleotide may be labeled.

The primers for amplification or sequencing reaction of a polynucleotide comprising a
10 biallelic marker of the invention may be designed from the disclosed sequences for any method known in the art. A preferred set of primers are fashioned such that the 3' end of the contiguous span of identity with a sequence selected from the group consisting of SEQ ID Nos 1 and 2 or a sequence complementary thereto or a variant thereof is present at the 3' end of the primer. Such a configuration allows the 3' end of the primer to hybridize to a selected nucleic acid sequence and
15 dramatically increases the efficiency of the primer for amplification or sequencing reactions. Allele specific primers may be designed such that a polymorphic base of a biallelic marker is at the 3' end of the contiguous span and the contiguous span is present at the 3' end of the primer. Such allele specific primers tend to selectively prime an amplification or sequencing reaction so long as they are used with a nucleic acid sample that contains one of the two alleles present at a biallelic marker.
20 The 3' end of the primer of the invention may be located within or at least 2, 4, 6, 8, 10, 12, 15, 18, 20, 25, 50, 100, 250, 500, or 1000 nucleotides upstream of a PG-3-related biallelic marker in said sequence or at any other location which is appropriate for their intended use in sequencing, amplification or the location of novel sequences or markers. Thus, another set of preferred amplification primers comprise an isolated polynucleotide consisting essentially of a contiguous
25 span of at least 8, 10, 12, 15, 18, 20, 25, 30, 35, 40, or 50 nucleotides in length of a sequence selected from the group consisting of SEQ ID Nos 1 and 2 or a sequence complementary thereto or a variant thereof, wherein the 3' end of said contiguous span is located at the 3' end of said polynucleotide, and wherein the 3' end of said polynucleotide is located upstream of a PG-3-related biallelic marker in said sequence. Preferably, those amplification primers comprise a sequence
30 selected from the group consisting of the sequences B1 to B52 and C1 to C52. Primers with their 3' ends located 1 nucleotide upstream of a biallelic marker of PG-3 have a special utility as microsequencing assays. Preferred microsequencing primers are described in Table 4. Optionally, said PG-3-related biallelic marker is selected from the group consisting of A1 to A80, and the complements thereof, or optionally the biallelic markers in linkage disequilibrium therewith;
35 optionally, said PG-3-related biallelic marker is selected from the group consisting of A1 to A5 and A8 to A80, and the complements thereof, or optionally the biallelic markers in linkage disequilibrium therewith; optionally, said PG-3-related biallelic marker is selected from the group

consisting A6 and A7, and the complements thereof, or optionally the biallelic markers in linkage disequilibrium therewith; Optionally, microsequencing primers are selected from the group consisting of the nucleotide sequences of D1 to D4, D6 to D80, E1 to E4 and E6 to E80. More preferred microsequencing primers are selected from the group consisting of the nucleotides
5 sequences of D14, D46, D68, D70, D71, E3, E6, E7, E11, E13, E42, E44, E72 and E75.

The probes of the present invention may be designed from the disclosed sequences for use in any method known in the art, particularly methods for testing if a marker disclosed herein is present in a sample. A preferred set of probes may be designed for use in the hybridization assays of the invention in any manner known in the art such that they selectively bind to one allele of a
10 biallelic marker, but not the other under any particular set of assay conditions. Preferred hybridization probes comprise the polymorphic base of either allele 1 or allele 2 of the relevant biallelic marker. Optionally, said biallelic marker may be within 6, 5, 4, 3, 2, or 1 nucleotides of the center of the hybridization probe or at the center of said probe. In a preferred embodiment, the probes are selected from the group consisting of the sequences P1 to P4 and P6 to P80 and the
15 complementary sequence thereto.

It should be noted that the polynucleotides of the present invention are not limited to having the exact flanking sequences surrounding the polymorphic bases which are enumerated in Sequence Listing. Rather, it will be appreciated that the flanking sequences surrounding the biallelic markers may be lengthened or shortened to any extent compatible with their intended use and the present
20 invention specifically contemplates such sequences. The flanking regions outside of the contiguous span need not be homologous to native flanking sequences which actually occur in human subjects. The addition of any nucleotide sequence which is compatible with the polynucleotide's intended use is specifically contemplated.

Primers and probes may be labeled or immobilized on a solid support as described in the
25 section entitled "Oligonucleotide probes and primers".

The polynucleotides of the invention which are attached to a solid support encompass polynucleotides with any further limitation described in this disclosure, or those following, alone or in any combination: Optionally, said polynucleotides may be attached individually or in groups of at least 2, 5, 8, 10, 12, 15, 20, or 25 distinct polynucleotides of the invention to a single solid support.
30 Optionally, polynucleotides other than those of the invention may attached to the same solid support as polynucleotides of the invention. Optionally, when multiple polynucleotides are attached to a solid support they may be attached at random locations, or in an ordered array. Optionally, said ordered array may be addressable.

The present invention also encompasses diagnostic kits comprising one or more
35 polynucleotides of the invention with a portion or all of the necessary reagents and instructions for genotyping a test subject by determining the identity of a nucleotide at a PG-3-related biallelic marker. The polynucleotides of a kit may optionally be attached to a solid support, or be part of an

array or addressable array of polynucleotides. The kit may provide for the determination of the identity of the nucleotide at a marker position by any method known in the art including, but not limited to, a sequencing assay method, a microsequencing assay method, a hybridization assay method, or an enzyme-based mismatch detection assay method.

5 **METHODS FOR *DE NOVO* IDENTIFICATION OF BIALLELIC MARKERS**

Any of a variety of methods can be used to screen a genomic fragment for single nucleotide polymorphisms, including methods such as differential hybridization with oligonucleotide probes, detection of changes in the mobility measured by gel electrophoresis or direct sequencing of the amplified nucleic acid. A preferred method for identifying biallelic markers involves comparative
10 sequencing of genomic DNA fragments from an appropriate number of unrelated individuals.

In a first embodiment, DNA samples from unrelated individuals are pooled together, following which the genomic DNA of interest is amplified and sequenced. The nucleotide sequences thus obtained are then analyzed to identify significant polymorphisms. One of the major advantages of this method resides in the fact that the pooling of the DNA samples substantially
15 reduces the number of DNA amplification reactions and sequencing reactions, which must be carried out. Moreover, this method is sufficiently sensitive so that a biallelic marker obtained thereby usually demonstrates a sufficient frequency of its less common allele to be useful in conducting association studies.

In a second embodiment, the DNA samples are not pooled and are therefore amplified and
20 sequenced individually. This method is usually preferred when biallelic markers need to be identified in order to perform association studies within candidate genes. Preferably, highly relevant gene regions such as promoter regions or exon regions may be screened for biallelic markers. A biallelic marker obtained using this method may show a lower degree of informativeness for conducting association studies, e.g. if the frequency of its less frequent allele is
25 less than about 10%. Such a biallelic marker will, however, be sufficiently informative to conduct association studies and it will further be appreciated that including less informative biallelic markers in the genetic analysis studies of the present invention, may, in some cases, allow the direct identification of causal mutations, which may, depending on their penetrance, be rare mutations.

The following is a description of the various parameters of a preferred method used by the
30 inventors for the identification of the biallelic markers of the present invention.

Genomic DNA Samples

The genomic DNA samples from which the biallelic markers of the present invention are generated are preferably obtained from unrelated individuals corresponding to a heterogeneous population of known ethnic background. The number of individuals from whom DNA samples are
35 obtained can vary substantially, but is preferably from about 10 to about 1000, or preferably from about 50 to about 200 individuals. It is usually preferred to collect DNA samples from at least

about 100 individuals in order to have sufficient polymorphic diversity in a given population to identify as many markers as possible and to generate statistically significant results.

As for the source of the genomic DNA to be subjected to analysis, any test sample can be foreseen without any particular limitation. These test samples include biological samples, which
5 can be tested by the methods of the present invention described herein, and include human and animal body fluids such as whole blood, serum, plasma, cerebrospinal fluid, urine, lymph fluids, and various external secretions of the respiratory, intestinal and genitourinary tracts, tears, saliva, milk, white blood cells, myelomas and the like; biological fluids such as cell culture supernatants; fixed tissue specimens including tumor and non-tumor tissue and lymph node tissues; bone marrow
10 aspirates and fixed cell specimens. The preferred source of genomic DNA used in the present invention is from peripheral venous blood of each donor. Techniques to prepare genomic DNA from biological samples are well known to the skilled technician. Details of a preferred embodiment are provided in Example 1. The person skilled in the art can choose to amplify pooled or unpooled DNA samples.

15 DNA Amplification

The identification of biallelic markers in a sample of genomic DNA may be facilitated through the use of DNA amplification methods. DNA samples can be pooled or unpooled for the amplification step. DNA amplification techniques are well known to those skilled in the art.

Amplification techniques that can be used in the context of the present invention include,
20 but are not limited to, the ligase chain reaction (LCR) described in EP-A- 320 308, WO 9320227 and EP-A-439 182, the polymerase chain reaction (PCR, RT-PCR) and techniques such as the nucleic acid sequence based amplification (NASBA) described in Guatelli J.C., *et al.*(1990) and in Compton J.(1991), Q-beta amplification as described in European Patent Application No 4544610, strand displacement amplification as described in Walker *et al.*(1996) and EP A 684 315 and, target
25 mediated amplification as described in PCT Publication WO 9322461.

LCR and Gap LCR are exponential amplification techniques, both of which utilize DNA ligase to join adjacent primers annealed to a DNA molecule. In Ligase Chain Reaction (LCR), probe pairs are used which include two primary (first and second) and two secondary (third and fourth) probes, all of which are employed in molar excess to target. The first probe hybridizes to a
30 first segment of the target strand and the second probe hybridizes to a second segment of the target strand, the first and second segments being contiguous so that the primary probes abut one another in 5' phosphate-3'hydroxyl relationship, and so that a ligase can covalently fuse or ligate the two probes into a fused product. In addition, a third (secondary) probe can hybridize to a portion of the first probe and a fourth (secondary) probe can hybridize to a portion of the second probe in a similar
35 abutting fashion. Of course, if the target is initially double stranded, the secondary probes also will hybridize to the target complement in the first instance. Once the ligated strand of primary probes is separated from the target strand, it will hybridize with the third and fourth probes, which can be

ligated to form a complementary, secondary ligated product. It is important to realize that the ligated products are functionally equivalent to either the target or its complement. By repeated cycles of hybridization and ligation, amplification of the target sequence is achieved. A method for multiplex LCR has also been described (WO 9320227). Gap LCR (GLCR) is a version of LCR
5 where the probes are not adjacent but are separated by 2 to 3 bases.

For amplification of mRNAs, it is within the scope of the present invention to reverse transcribe mRNA into cDNA followed by polymerase chain reaction (RT-PCR); or, to use a single enzyme for both steps as described in U.S. Patent No. 5,322,770 or, to use Asymmetric Gap LCR (RT-AGLCR) as described by Marshall *et al.* (1994). AGLCR is a modification of GLCR that
10 allows the amplification of RNA.

The PCR technology is the preferred amplification technique used in the present invention. A variety of PCR techniques are familiar to those skilled in the art. For a review of PCR technology, see White (1992) and the publication entitled "PCR Methods and Applications" (1991, Cold Spring Harbor Laboratory Press). In each of these PCR procedures, PCR primers on either
15 side of the nucleic acid sequences to be amplified are added to a suitably prepared nucleic acid sample along with dNTPs and a thermostable polymerase such as Taq polymerase, Pfu polymerase, or Vent polymerase. The nucleic acid in the sample is denatured and the PCR primers are specifically hybridized to complementary nucleic acid sequences in the sample. The hybridized primers are extended. Thereafter, another cycle of denaturation, hybridization, and extension is
20 initiated. The cycles are repeated multiple times to produce an amplified fragment containing the nucleic acid sequence between the primer sites. PCR has further been described in several patents including US Patents 4,683,195; 4,683,202; and 4,965,188.

The PCR technology is the preferred amplification technique used to identify new biallelic markers. A typical example of a PCR reaction suitable for the purposes of the present invention is
25 provided in Example 2.

One of the aspects of the present invention is a method for the amplification of the human PG-3 gene, particularly of a fragment of the genomic sequence of SEQ ID No 1 or of the cDNA sequence of SEQ ID No 2, or a fragment or a variant thereof in a test sample, preferably using the PCR technology. This method comprises the steps of:

- 30 a) contacting a test sample with amplification reaction reagents comprising a pair of amplification primers as described above which are located on either side of the polynucleotide region to be amplified, and
 b) optionally, detecting the amplification products.

The invention also concerns a kit for the amplification of a PG-3 gene sequence,
35 particularly of a portion of the genomic sequence of SEQ ID No 1 or of the cDNA sequence of SEQ ID No 2, or a variant thereof in a test sample, wherein said kit comprises:

a) a pair of oligonucleotide primers located on either side of the PG-3 region to be amplified;

b) optionally, the reagents necessary for performing the amplification reaction.

In one embodiment of the above amplification method and kit, the amplification product is
5 detected by hybridization with a labeled probe having a sequence which is complementary to the amplified region. In another embodiment of the above amplification method and kit, primers comprise a sequence which is selected from the group consisting of the nucleotide sequences of B1 to B52, C1 to C52, D1 to D4, D6 to D80, E1 to E4, and E6 to E80.

In a first embodiment of the present invention, biallelic markers are identified using
10 genomic sequence information generated by the inventors. Sequenced genomic DNA fragments are used to design primers for the amplification of 500 bp fragments. These 500 bp fragments are amplified from genomic DNA and are scanned for biallelic markers. Primers may be designed using the OSP software (Hillier L. and Green P., 1991). All primers may contain, upstream of the specific target bases, a common oligonucleotide tail that serves as a sequencing primer. Those
15 skilled in the art are familiar with primer extensions, which can be used for these purposes.

Preferred primers, useful for the amplification of genomic sequences encoding the candidate genes, focus on promoters, exons and splice sites of the genes. A biallelic marker presents a higher probability to be a causal mutation if it is located in these functional regions of the gene. Preferred amplification primers of the invention include the nucleotide sequences B1 to B52
20 and C1 to C52, detailed further in Example 2, Table 1.

Sequencing Of Amplified Genomic DNA And Identification Of Single Nucleotide Polymorphisms

The amplification products generated as described above, are then sequenced using any method known and available to the skilled technician. Methods for sequencing DNA using either
25 the dideoxy-mediated method (Sanger method) or the Maxam-Gilbert method are widely known to those of ordinary skill in the art. Such methods are disclosed in Sambrook *et al.* (1989) for example. Alternative approaches include hybridization to high-density DNA probe arrays as described in Chee *et al.* (1996).

Preferably, the amplified DNA is subjected to automated dideoxy terminator sequencing
30 reactions using a dye-primer cycle sequencing protocol. The products of the sequencing reactions are run on sequencing gels and the sequences are determined using gel image analysis. The polymorphism search is based on the presence of superimposed peaks in the electrophoresis pattern resulting from different bases occurring at the same position. Because each dideoxy terminator is labeled with a different fluorescent molecule, the two peaks corresponding to a biallelic site present
35 distinct colors corresponding to two different nucleotides at the same position on the sequence. However, the presence of two peaks can be an artifact due to background noise. To exclude such an

artifact, the two DNA strands are sequenced and a comparison between the peaks is carried out. In order to confirm that a sequence is polymorphic, the polymorphism is detected on both strands.

The above procedure permits those amplification products which contain biallelic markers to be identified. The detection limit for the frequency of biallelic polymorphisms detected by sequencing pools of 100 individuals is approximately 0.1 for the minor allele, as verified by sequencing pools of known allelic frequencies. However, more than 90% of the biallelic polymorphisms detected by the pooling method have a frequency for the minor allele higher than 0.25. Therefore, the biallelic markers selected by this method have a frequency of at least 0.1 for the minor allele and less than 0.9 for the major allele. Preferably, the biallelic markers selected by this method have a frequency of at least 0.2 for the minor allele and less than 0.8 for the major allele, more preferably at least 0.3 for the minor allele and less than 0.7 for the major allele. Thus, the biallelic markers preferably have a heterozygosity rate higher than 0.18, more preferably higher than 0.32, still more preferably higher than 0.42.

In another embodiment, biallelic markers are detected by sequencing individual DNA samples. In some embodiments, the frequency of the minor allele of such a biallelic marker may be less than 0.1.

Validation Of The Biallelic Markers Of The Present Invention

The polymorphisms are evaluated for their usefulness as genetic markers by validating that both alleles are present in a population. Validation of the biallelic markers is accomplished by genotyping a group of individuals by a method of the invention and demonstrating that both alleles are present. Microsequencing is a preferred method of genotyping alleles. The validation by genotyping step may be performed on individual samples derived from each individual in the group or by genotyping a pooled sample derived from more than one individual. The group can be as small as one individual if that individual is heterozygous for the allele in question. Preferably the group contains at least three individuals, more preferably the group contains five or six individuals, so that a single validation test will be more likely to result in the validation of more of the biallelic markers that are being tested. It should be noted, however, that when the validation test is performed on a small group it may result in a false negative result if as a result of sampling error none of the individuals tested carries one of the two alleles. Thus, the validation process is less useful in demonstrating that a particular initial result is an artifact, than it is at demonstrating that there is a *bona fide* biallelic marker at a particular position in a sequence. All of the genotyping, haplotyping, association, and interaction study methods of the invention may optionally be performed solely with validated biallelic markers.

Evaluation Of The Frequency Of The Biallelic Markers Of The Present Invention

The validated biallelic markers are further evaluated for their usefulness as genetic markers by determining the frequency of the least common allele at the biallelic marker site. The higher the frequency of the less common allele the greater the usefulness of the biallelic marker in association

and interaction studies. The identification of the least common allele is accomplished by genotyping a group of individuals by a method of the invention and demonstrating that both alleles are present. The determination of marker frequency by genotyping may be performed using individual samples derived from each individual in the group or by genotyping a pooled sample
5 derived from more than one individual. The group must be large enough to be representative of the population as a whole. Preferably the group contains at least 20 individuals, more preferably the group contains at least 50 individuals, most preferably the group contains at least 100 individuals. Of course the larger the group the greater the accuracy of the frequency determination because of reduced sampling error. A biallelic marker wherein the frequency of the less common allele is 30%
10 or more is termed a "high quality biallelic marker." All of the genotyping, haplotyping, association, and interaction study methods of the invention may optionally be performed solely with high quality biallelic markers.

METHODS FOR GENOTYPING AN INDIVIDUAL FOR BIALLELIC MARKERS

Methods are provided to genotype a biological sample for one or more biallelic markers of
15 the present invention, all of which may be performed *in vitro*. Such methods of genotyping comprise determining the identity of a nucleotide at a PG-3 biallelic marker site by any method known in the art. These methods find use in genotyping case-control populations in association studies as well as individuals in the context of detection of alleles of biallelic markers which are known to be associated with a given trait, in which case both copies of the biallelic marker present
20 in individual's genome are determined so that an individual may be classified as homozygous or heterozygous for a particular allele.

These genotyping methods can be performed on nucleic acid samples derived from a single individual or pooled DNA samples.

Genotyping can be performed using methods similar to those described above for the
25 identification of the biallelic markers, or using other genotyping methods such as those further described below. In preferred embodiments, the comparison of sequences of amplified genomic fragments from different individuals is used to identify new biallelic markers whereas microsequencing is used for genotyping known biallelic markers in diagnostic and association study applications.

30 In one embodiment, the invention encompasses methods of genotyping comprising determining the identity of a nucleotide at a PG-3-related biallelic marker or the complement thereof in a biological sample; optionally, the PG-3-related biallelic marker is selected from the group consisting of A1 to A80, and the complements thereof, or optionally the biallelic markers in linkage disequilibrium therewith; optionally, wherein said PG-3-related biallelic marker is selected
35 from the group consisting of A1 to A5 and A8 to A80, and the complements thereof, or optionally the biallelic markers in linkage disequilibrium therewith; optionally, wherein said PG-3-related biallelic marker is selected from the group consisting of A6 and A7, and the complements thereof,

or optionally the biallelic markers in linkage disequilibrium therewith; optionally, the biological sample is derived from a single subject; optionally, the identity of the nucleotides at said biallelic marker is determined for both copies of said biallelic marker present in said individual's genome; optionally, said biological sample is derived from multiple subjects; Optionally, the genotyping methods of the invention encompass methods with any further limitation described in this disclosure, or those following, alone or in any combination; Optionally, said method is performed *in vitro*; optionally, the method further comprises amplifying a portion of said sequence comprising the biallelic marker prior to said determining step; Optionally, the amplification is performed by PCR, LCR, or replication of a recombinant vector comprising an origin of replication and said fragment in a host cell; optionally, the determination involves a hybridization assay, a sequencing assay, a microsequencing assay, or an enzyme-based mismatch detection assay.

Source of Nucleic Acids for genotyping

Any source of nucleic acids, in purified or non-purified form, can be utilized as the starting nucleic acid, provided it contains or is suspected of containing the specific nucleic acid sequence desired. DNA or RNA may be extracted from cells, tissues, body fluids and the like as described above. While nucleic acids for use in the genotyping methods of the invention can be derived from any mammalian source, the test subjects and individuals from which nucleic acid samples are taken are generally understood to be human.

Amplification Of DNA Fragments Comprising Biallelic Markers

Methods and polynucleotides are provided to amplify a segment of nucleotides comprising one or more biallelic marker of the present invention. It will be appreciated that amplification of DNA fragments comprising biallelic markers may be used in various methods and for various purposes and is not restricted to genotyping. Nevertheless, many genotyping methods, although not all, require the previous amplification of the DNA region carrying the biallelic marker of interest. Such methods specifically increase the concentration or total number of sequences that span the biallelic marker or include that site and sequences located either distal or proximal to it. Diagnostic assays may also rely on amplification of DNA segments carrying a biallelic marker of the present invention. Amplification of DNA may be achieved by any method known in the art. Amplification techniques are described above in the section entitled, "DNA amplification."

Some of these amplification methods are particularly suited for the detection of single nucleotide polymorphisms and allow the simultaneous amplification of a target sequence and the identification of the polymorphic nucleotide as further described below.

The identification of biallelic markers as described above allows the design of appropriate oligonucleotides, which can be used as primers to amplify DNA fragments comprising the biallelic markers of the present invention. Amplification can be performed using the primers initially used to discover new biallelic markers which are described herein or any set of primers allowing the amplification of a DNA fragment comprising a biallelic marker of the present invention.

In some embodiments, the present invention provides primers for amplifying a DNA fragment containing one or more biallelic markers of the present invention. Preferred amplification primers are listed in Example 2. It will be appreciated that the primers listed are merely exemplary and that any other set of primers which produce amplification products containing one or more

5 biallelic markers of the present invention are also of use.

The spacing of the primers determines the length of the segment to be amplified. In the context of the present invention, amplified segments carrying biallelic markers can range in size from at least about 25 bp to 35 kbp. Amplification fragments from 25-3000 bp are typical, fragments from 50-1000 bp are preferred and fragments from 100-600 bp are highly preferred. It

10 will be appreciated that amplification primers for the biallelic markers may be any sequence which allow the specific amplification of any DNA fragment carrying the markers. Amplification primers may be labeled or immobilized on a solid support as described in the section "Oligonucleotide probes and primers".

Methods of Genotyping DNA samples for Biallelic Markers

15 Any method known in the art can be used to identify the nucleotide present at a biallelic marker site. Since the biallelic marker allele to be detected has been identified and specified in the present invention, detection will prove simple for one of ordinary skill in the art by employing any of a number of techniques. Many genotyping methods require the previous amplification of the DNA region carrying the biallelic marker of interest. While the amplification of target or signal is

20 often preferred at present, ultrasensitive detection methods which do not require amplification are also encompassed by the present genotyping methods. Methods well-known to those skilled in the art that can be used to detect biallelic polymorphisms include methods such as, conventional dot blot analyzes, single strand conformational polymorphism analysis (SSCP) described by Orita *et al.*(1989), denaturing gradient gel electrophoresis (DGGE), heteroduplex analysis, mismatch

25 cleavage detection, and other conventional techniques as described in Sheffield *et al.*(1991), White *et al.*(1992), Grompe *et al.*(1989 and 1993). Another method for determining the identity of the nucleotide present at a particular polymorphic site employs a specialized exonuclease-resistant nucleotide derivative as described in US patent 4,656,127.

Preferred methods involve directly determining the identity of the nucleotide present at a

30 biallelic marker site by sequencing assay, enzyme-based mismatch detection assay, or hybridization assay. The following is a description of some preferred methods. A highly preferred method is the microsequencing technique. The term "sequencing" is generally used herein to refer to polymerase extension of duplex primer/template complexes and includes both traditional sequencing and microsequencing.

35 1) Sequencing Assays

The nucleotide present at a polymorphic site can be determined by sequencing methods. In a preferred embodiment, DNA samples are subjected to PCR amplification before sequencing as

described above. DNA sequencing methods are described in the section entitled "Sequencing Of Amplified Genomic DNA And Identification Of Single Nucleotide Polymorphisms".

Preferably, the amplified DNA is subjected to automated dideoxy terminator sequencing reactions using a dye-primer cycle sequencing protocol. Sequence analysis allows the identification of the base present at the biallelic marker site.

2) Microsequencing Assays

In microsequencing methods, the nucleotide at a polymorphic site in a target DNA is detected by a single nucleotide primer extension reaction. This method involves appropriate microsequencing primers which hybridize just upstream of the polymorphic base of interest in the target nucleic acid. A polymerase is used to specifically extend the 3' end of the primer with one single ddNTP (chain terminator) complementary to the nucleotide at the polymorphic site. Next the identity of the incorporated nucleotide is determined in any suitable way.

Typically, microsequencing reactions are carried out using fluorescent ddNTPs and the extended microsequencing primers are analyzed by electrophoresis on ABI 377 sequencing machines to determine the identity of the incorporated nucleotide as described in EP 412 883. Alternatively capillary electrophoresis can be used in order to process a higher number of assays simultaneously. An example of a typical microsequencing procedure that can be used in the context of the present invention is provided in Example 4.

Different approaches can be used for the labeling and detection of ddNTPs. A homogeneous phase detection method based on fluorescence resonance energy transfer has been described by Chen and Kwok (1997) and Chen *et al.* (1997). In this method, amplified genomic DNA fragments containing polymorphic sites are incubated with a 5'-fluorescein-labeled primer in the presence of allelic dye-labeled dideoxyribonucleoside triphosphates and a modified Taq polymerase. The dye-labeled primer is extended one base by the dye-terminator specific for the allele present on the template. At the end of the genotyping reaction, the fluorescence intensities of the two dyes in the reaction mixture are analyzed directly without separation or purification. All these steps can be performed in the same tube and the fluorescence changes can be monitored in real time. Alternatively, the extended primer may be analyzed by MALDI-TOF Mass Spectrometry. The base at the polymorphic site is identified by the mass added onto the microsequencing primer (see Haff and Smirnov, 1997).

Microsequencing may be achieved by the established microsequencing method or by developments or derivatives thereof. Alternative methods include several solid-phase microsequencing techniques. The basic microsequencing protocol is the same as described previously, except that the method is conducted as a heterogeneous phase assay, in which the primer or the target molecule is immobilized or captured onto a solid support. To simplify the primer separation and the terminal nucleotide addition analysis, oligonucleotides are attached to solid supports or are modified in such ways that permit affinity separation as well as polymerase

extension. The 5' ends and internal nucleotides of synthetic oligonucleotides can be modified in a number of different ways to permit different affinity separation approaches, e.g., biotinylation. If a single affinity group is used on the oligonucleotides, the oligonucleotides can be separated from the incorporated terminator reagent. This eliminates the need of physical or size separation. More than one oligonucleotide can be separated from the terminator reagent and analyzed simultaneously if more than one affinity group is used. This permits the analysis of several nucleic acid species or more nucleic acid sequence information per extension reaction. The affinity group need not be on the priming oligonucleotide but could alternatively be present on the template. For example, immobilization can be carried out via an interaction between biotinylated DNA and streptavidin-coated microtitration wells or avidin-coated polystyrene particles. In the same manner, oligonucleotides or templates may be attached to a solid support in a high-density format. In such solid phase microsequencing reactions, incorporated ddNTPs can be radiolabeled (Sylvänen, 1994) or linked to fluorescein (Livak and Hainer, 1994). The detection of radiolabeled ddNTPs can be achieved through scintillation-based techniques. The detection of fluorescein-linked ddNTPs can be based on the binding of anti fluorescein antibody conjugated with alkaline phosphatase, followed by incubation with a chromogenic substrate (such as *p*-nitrophenyl phosphate). Other possible reporter-detection pairs include: ddNTP linked to dinitrophenyl (DNP) and anti-DNP alkaline phosphatase conjugate (Harju *et al.*, 1993) or biotinylated ddNTP and horseradish peroxidase-conjugated streptavidin with *o*-phenylenediamine as a substrate (WO 92/15712). As yet another alternative solid-phase microsequencing procedure, Nyren *et al.* (1993) described a method relying on the detection of DNA polymerase activity by an enzymatic luminometric inorganic pyrophosphate detection assay (ELIDA).

Pastinen *et al.* (1997) describe a method for multiplex detection of single nucleotide polymorphism in which the solid phase minisequencing principle is applied to an oligonucleotide array format. High-density arrays of DNA probes attached to a solid support (DNA chips) are further described below.

In one aspect the present invention provides polynucleotides and methods to genotype one or more biallelic markers of the present invention by performing a microsequencing assay. Preferred microsequencing primers include the nucleotide sequences D1 to D4 and D6 to D80 and E1 to E4 and E6 to E80. It will be appreciated that the microsequencing primers listed in Example 4 are merely exemplary and that any primer having a 3' end immediately adjacent to the polymorphic nucleotide may be used. Similarly, it will be appreciated that microsequencing analysis may be performed for any biallelic marker or any combination of biallelic markers of the present invention. One aspect of the present invention is a solid support which includes one or more microsequencing primers listed in Example 4, or fragments comprising at least 8, 12, 15, 20, 25, 30, 40, or 50 consecutive nucleotides thereof, to the extent that such lengths are consistent with

the primer described, and having a 3' terminus immediately upstream of the corresponding biallelic marker, for determining the identity of a nucleotide at a biallelic marker site.

3) Mismatch detection assays based on polymerases and ligases

In one aspect the present invention provides polynucleotides and methods to determine the
5 allele of one or more biallelic markers of the present invention in a biological sample, by mismatch
detection assays based on polymerases and/or ligases. These assays are based on the specificity of
polymerases and ligases. Polymerization reactions place particularly stringent requirements on
correct base pairing of the 3' end of the amplification primer and the joining of two oligonucleotides
hybridized to a target DNA sequence is quite sensitive to mismatches close to the ligation site,
10 especially at the 3' end. Methods, primers and various parameters to amplify DNA fragments
comprising biallelic markers of the present invention are further described above in the section
entitled "Amplification Of DNA Fragments Comprising Biallelic Markers".

Allele Specific Amplification Primers

Discrimination between the two alleles of a biallelic marker can also be achieved by allele
15 specific amplification, a selective strategy whereby one of the alleles is amplified without
amplification of the other allele. For allele specific amplification, at least one member of the pair of
primers is sufficiently complementary with a region of a PG-3 gene comprising the polymorphic
base of a biallelic marker of the present invention to hybridize therewith and to initiate the
amplification. Such primers are able to discriminate between the two alleles of a biallelic marker.

20 This is accomplished by placing the polymorphic base at the 3' end of one of the
amplification primers. Because the extension progresses from the 3' end of the primer, a mismatch
at or near this position has an inhibitory effect on amplification. Therefore, under appropriate
amplification conditions, these primers only direct amplification on their complementary allele.
Determining the precise location of the mismatch and the corresponding assay conditions are well
25 within the ordinary skill in the art.

Ligation/Amplification Based Methods

The "Oligonucleotide Ligation Assay" (OLA) uses two oligonucleotides which are
designed to be capable of hybridizing to abutting sequences of a single strand of a target molecules.
One of the oligonucleotides is biotinylated, and the other is detectably labeled. If the precise
30 complementary sequence is found in a target molecule, the oligonucleotides will hybridize such that
their termini abut, and create a ligation substrate that can be captured and detected. OLA is capable
of detecting single nucleotide polymorphisms and may be advantageously combined with PCR as
described by Nickerson *et al.* (1990). In this method, PCR is used to achieve the exponential
amplification of target DNA, which is then detected using OLA.

35 Other amplification methods which are particularly suited for the detection of single
nucleotide polymorphism include LCR (ligase chain reaction), Gap LCR (GLCR) which are
described above in the section entitled "DNA Amplification". LCR uses two pairs of probes to

exponentially amplify a specific target. The sequences of each pair of oligonucleotides are selected to permit the pair to hybridize to abutting sequences of the same strand of the target. Such hybridization forms a substrate for a template-dependant ligase. In accordance with the present invention, LCR can be performed with oligonucleotides having the proximal and distal sequences of the same strand of a biallelic marker site. In one embodiment, either oligonucleotide will be designed to include the biallelic marker site. In such an embodiment, the reaction conditions are selected such that the oligonucleotides can be ligated together only if the target molecule either contains or lacks the specific nucleotide that is complementary to the biallelic marker on the oligonucleotide. In an alternative embodiment, the oligonucleotides will not include the biallelic marker, such that when they hybridize to the target molecule, a "gap" is created as described in WO 90/01069. This gap is then "filled" with complementary dNTPs (as mediated by DNA polymerase), or by an additional pair of oligonucleotides. Thus at the end of each cycle, each single strand has a complement capable of serving as a target during the next cycle and exponential allele-specific amplification of the desired sequence is obtained.

Ligase/Polymerase-mediated Genetic Bit AnalysisTM is another method for determining the identity of a nucleotide at a preselected site in a nucleic acid molecule (WO 95/21271). This method involves the incorporation of a nucleoside triphosphate that is complementary to the nucleotide present at the preselected site onto the terminus of a primer molecule, and their subsequent ligation to a second oligonucleotide. The reaction is monitored by detecting a specific label attached to the reaction's solid phase or by detection in solution.

4) Hybridization Assay Methods

A preferred method of determining the identity of the nucleotide present at a biallelic marker site involves nucleic acid hybridization. The hybridization probes, which can be conveniently used in such reactions, preferably include the probes defined herein. Any hybridization assay may be used including Southern hybridization, Northern hybridization, dot blot hybridization and solid-phase hybridization (see Sambrook *et al.*, 1989).

Hybridization refers to the formation of a duplex structure by two single stranded nucleic acids due to complementary base pairing. Hybridization can occur between exactly complementary nucleic acid strands or between nucleic acid strands that contain minor regions of mismatch. Specific probes can be designed that hybridize to one form of a biallelic marker and not to the other and therefore are able to discriminate between different allelic forms. Allele-specific probes are often used in pairs, one member of a pair showing perfect match to a target sequence containing the original allele and the other showing a perfect match to the target sequence containing the alternative allele. Hybridization conditions should be sufficiently stringent that there is a significant difference in hybridization intensity between alleles, and preferably an essentially binary response, whereby a probe hybridizes to only one of the alleles. Stringent, sequence specific hybridization conditions, under which a probe will hybridize only to the exactly complementary target sequence

are well known in the art (Sambrook *et al.*, 1989). Stringent conditions are sequence dependent and will be different in different circumstances. Generally, stringent conditions are selected to be about 5°C lower than the thermal melting point (T_m) for the specific sequence at a defined ionic strength and pH. Although such hybridization can be performed in solution, it is preferred to employ a solid-phase hybridization assay. The target DNA comprising a biallelic marker of the present invention may be amplified prior to the hybridization reaction. The presence of a specific allele in the sample is determined by detecting the presence or the absence of stable hybrid duplexes formed between the probe and the target DNA. The detection of hybrid duplexes can be carried out by a number of methods. Various detection assay formats are well known which utilize detectable labels bound to either the target or the probe to enable detection of the hybrid duplexes. Typically, hybridization duplexes are separated from unhybridized nucleic acids and the labels bound to the duplexes are then detected. Those skilled in the art will recognize that wash steps may be employed to wash away excess target DNA or probe as well as unbound conjugate. Further, standard heterogeneous assay formats are suitable for detecting the hybrids using the labels present on the primers and probes.

Two recently developed assays allow hybridization-based allele discrimination with no need for separations or washes (see Landegren U. *et al.*, 1998). The TaqMan assay takes advantage of the 5' nuclease activity of Taq DNA polymerase to digest a DNA probe annealed specifically to the accumulating amplification product. TaqMan probes are labeled with a donor-acceptor dye pair that interacts via fluorescence energy transfer. Cleavage of the TaqMan probe by the advancing polymerase during amplification dissociates the donor dye from the quenching acceptor dye, greatly increasing the donor fluorescence. All reagents necessary to detect two allelic variants can be assembled at the beginning of the reaction and the results are monitored in real time (see Livak *et al.*, 1995). In an alternative homogeneous hybridization based procedure, molecular beacons are used for allele discriminations. Molecular beacons are hairpin-shaped oligonucleotide probes that report the presence of specific nucleic acids in homogeneous solutions. When they bind to their targets they undergo a conformational reorganization that restores the fluorescence of an internally quenched fluorophore (Tyagi *et al.*, 1998).

The polynucleotides provided herein can be used to produce probes which can be used in hybridization assays for the detection of biallelic marker alleles in biological samples. These probes preferably comprise between 8 and 50 nucleotides and are sufficiently complementary to a sequence comprising a biallelic marker of the present invention to hybridize thereto and preferably sufficiently specific to be able to discriminate the targeted sequence for only one nucleotide variation. A particularly preferred probe is 25 nucleotides in length. Preferably the biallelic marker is within 4 nucleotides of the center of the polynucleotide probe. In particularly preferred probes, the biallelic marker is at the center of said polynucleotide. Preferred probes comprise a nucleotide sequence selected from the group consisting of amplicons listed in Table 1 and the sequences

complementary thereto, or a fragment thereof, said fragment comprising at least about 8 consecutive nucleotides, preferably 10, 15, 20, more preferably 25, 30, 40, 47, or 50 consecutive nucleotides and containing a polymorphic base. Preferred probes comprise a nucleotide sequence selected from the group consisting of P1 to P4 and P6 to P80 and the sequences complementary thereto. In preferred
5 embodiments the polymorphic base(s) are within 5, 4, 3, 2, 1, nucleotides of the center of the said polynucleotide, more preferably at the center of said polynucleotide.

Preferably the probes of the present invention are labeled or immobilized on a solid support. Labels and solid supports are further described in the section entitled "Oligonucleotide Probes and Primers". The probes can be non-extendable as described in the section entitled "Oligonucleotide
10 Probes and Primers".

By assaying the hybridization to an allele specific probe, one can detect the presence or absence of a biallelic marker allele in a given sample. High-Throughput parallel hybridization in array format is specifically encompassed within "hybridization assays" and is described below.

5) Hybridization To Addressable Arrays Of Oligonucleotides

Hybridization assays based on oligonucleotide arrays rely on the differences in
15 hybridization stability of short oligonucleotides to perfectly matched and mismatched target sequence variants. Efficient access to polymorphism information is obtained through a basic structure comprising high-density arrays of oligonucleotide probes attached to a solid support (e.g., the chip) at selected positions. Each DNA chip can contain thousands to millions of individual
20 synthetic DNA probes arranged in a grid-like pattern and miniaturized to the size of a dime.

The chip technology has already been applied with success in numerous cases. For example, the screening of mutations has been undertaken in the BRCA1 gene, in *S. cerevisiae* mutant strains, and in the protease gene of HIV-1 virus (Hacia *et al.*, 1996; Shoemaker *et al.*, 1996; Kozal *et al.*, 1996). Chips of various formats for use in detecting biallelic polymorphisms can be
25 produced on a customized basis by Affymetrix (GeneChip™), Hyseq (HyChip and HyGnostics), and Protogene Laboratories.

In general, these methods employ arrays of oligonucleotide probes that are complementary to target nucleic acid sequence segments from an individual which, target sequences include a polymorphic marker. EP 785280, describes a tiling strategy for the detection of single nucleotide
30 polymorphisms. Briefly, arrays may generally be "tiled" for a large number of specific polymorphisms. By "tiling" is generally meant the synthesis of a defined set of oligonucleotide probes which is made up of a sequence complementary to the target sequence of interest, as well as preselected variations of that sequence, e.g., substitution of one or more given positions with one or more members of the basis set of nucleotides. Tiling strategies are further described in PCT
35 application No. WO 95/11995. In a particular aspect, arrays are tiled for a number of specific, identified biallelic marker sequences. In particular, the array is tiled to include a number of detection blocks, each detection block being specific for a specific biallelic marker or a set of

biallelic markers. For example, a detection block may be tiled to include a number of probes, which span the sequence segment that includes a specific polymorphism. To obtain probes that are complementary to each allele, the probes are synthesized in pairs differing at the biallelic marker. In addition to the probes differing at the polymorphic base, monosubstituted probes are also

5 generally tiled within the detection block. These monosubstituted probes have bases at and up to a certain number of bases in either direction from the polymorphism, substituted with the remaining nucleotides (selected from A, T, G, C and U). Typically the probes in a tiled detection block will include substitutions of the sequence positions up to and including those that are 5 bases away from the biallelic marker. The monosubstituted probes provide internal controls for the tiled array, to

10 distinguish actual hybridization from artefactual cross-hybridization. Upon completion of hybridization with the target sequence and washing of the array, the array is scanned to determine the position on the array to which the target sequence hybridizes. The hybridization data from the scanned array is then analyzed to identify which allele or alleles of the biallelic marker are present in the sample. Hybridization and scanning may be carried out as described in PCT application No.

15 WO 92/10092 and WO 95/11995 and US patent No. 5,424,186.

Thus, in some embodiments, the chips may comprise an array of nucleic acid sequences about 15 nucleotides in length. In further embodiments, the chip may comprise an array including at least one of the sequences selected from the group consisting of amplicons listed in Table 1 and the sequences complementary thereto, or a fragment thereof, said fragment comprising at least

20 about 8 consecutive nucleotides, preferably 10, 15, 20, more preferably 25, 30, 40, 47, or 50 consecutive nucleotides and containing a polymorphic base. In preferred embodiments the polymorphic base is within 5, 4, 3, 2, 1, nucleotides of the center of the said polynucleotide, more preferably at the center of said polynucleotide. In some embodiments, the chip may comprise an array of at least 2, 3, 4, 5, 6, 7, 8 or more of these polynucleotides of the invention. Solid supports

25 and polynucleotides of the present invention attached to solid supports are further described in the section entitled "Oligonucleotide Probes And Primers".

6) Integrated Systems

Another technique, which may be used to analyze polymorphisms, includes multicomponent integrated systems, which miniaturize and compartmentalize processes such as

30 PCR and capillary electrophoresis reactions in a single functional device. An example of such technique is disclosed in US patent 5,589,136, which describes the integration of PCR amplification and capillary electrophoresis in chips.

Integrated systems can be envisaged mainly when microfluidic systems are used. These systems comprise a pattern of microchannels designed onto a glass, silicon, quartz, or plastic wafer

35 included on a microchip. The movements of the samples are controlled by electric, electroosmotic or hydrostatic forces applied across different areas of the microchip to create functional microscopic valves and pumps with no moving parts.

For genotyping biallelic markers, the microfluidic system may integrate nucleic acid amplification, microsequencing, capillary electrophoresis and a detection method such as laser-induced fluorescence detection.

METHODS OF GENETIC ANALYSIS USING THE BIALLELIC MARKERS OF
THE PRESENT INVENTION

5 Different methods are available for the genetic analysis of complex traits (see Lander and Schork, 1994). The search for disease-susceptibility genes is conducted using two main methods: the linkage approach in which evidence is sought for cosegregation between a locus and a putative trait locus using family studies, and the association approach in which evidence is sought for a
10 statistically significant association between an allele and a trait or a trait causing allele (Khoury *et al.*, 1993). In general, the biallelic markers of the present invention find use in any method known in the art to demonstrate a statistically significant correlation between a genotype and a phenotype. The biallelic markers may be used in parametric and non-parametric linkage analysis methods. Preferably, the biallelic markers of the present invention are used to identify genes associated with
15 detectable traits using association studies, an approach which does not require the use of affected families and which permits the identification of genes associated with complex and sporadic traits.

The genetic analysis using the biallelic markers of the present invention may be conducted on any scale. The whole set of biallelic markers of the present invention or any subset of biallelic markers of the present invention corresponding to the candidate gene may be used. Further, any set
20 of genetic markers including a biallelic marker of the present invention may be used. A set of biallelic polymorphisms that could be used as genetic markers in combination with the biallelic markers of the present invention has been described in WO 98/20165. As mentioned above, it should be noted that the biallelic markers of the present invention may be included in any complete or partial genetic map of the human genome. These different uses are specifically contemplated in
25 the present invention and claims.

Linkage Analysis

Linkage analysis is based upon establishing a correlation between the transmission of genetic markers and that of a specific trait throughout generations within a family. Thus, the aim of linkage analysis is to detect marker loci that show cosegregation with a trait of interest in pedigrees.

30 **PARAMETRIC METHODS**

When data are available from successive generations there is the opportunity to study the degree of linkage between pairs of loci. Estimates of the recombination fraction enable loci to be ordered and placed onto a genetic map. With loci that are genetic markers, a genetic map can be established, and then the strength of linkage between markers and traits can be calculated and used
35 to indicate the relative positions of markers and genes affecting those traits (Weir, 1996). The classical method for linkage analysis is the logarithm of odds (lod) score method (see Morton, 1955; Ott, 1991). Calculation of lod scores requires specification of the mode of inheritance for the

disease (parametric method). Generally, the length of the candidate region identified using linkage analysis is between 2 and 20Mb. Once a candidate region is identified as described above, analysis of recombinant individuals using additional markers allows further delineation of the candidate region. Linkage analysis studies have generally relied on the use of a maximum of 5,000
5 microsatellite markers, thus limiting the maximum theoretical attainable resolution of linkage analysis to about 600 kb on average.

Linkage analysis has been successfully applied to map simple genetic traits that show clear Mendelian inheritance patterns and which have a high penetrance (*i.e.*, the ratio between the number of trait positive carriers of allele *a* and the total number of *a* carriers in the population). However,
10 parametric linkage analysis suffers from a variety of drawbacks. First, it is limited by its reliance on the choice of a genetic model suitable for each studied trait. Furthermore, as already mentioned, the resolution attainable using linkage analysis is limited, and complementary studies are required to refine the analysis of the typical 2Mb to 20Mb regions initially identified through linkage analysis. In addition, parametric linkage analysis approaches have proven difficult when applied to complex
15 genetic traits, such as those due to the combined action of multiple genes and/or environmental factors. It is very difficult to model these factors adequately in a lod score analysis. In such cases, too large an effort and cost are needed to recruit the adequate number of affected families required for applying linkage analysis to these situations, as recently discussed by Risch, N. and Merikangas, K. (1996).

20 NON-PARAMETRIC METHODS

The advantage of the so-called non-parametric methods for linkage analysis is that they do not require specification of the mode of inheritance for the disease, they tend to be more useful for the analysis of complex traits. In non-parametric methods, one tries to prove that the inheritance pattern of a chromosomal region is not consistent with random Mendelian segregation by showing
25 that affected relatives inherit identical copies of the region more often than expected by chance. Affected relatives should show excess "allele sharing" even in the presence of incomplete penetrance and polygenic inheritance. In non-parametric linkage analysis the degree of agreement at a marker locus in two individuals can be measured either by the number of alleles identical by state (IBS) or by the number of alleles identical by descent (IBD). Affected sib pair analysis is a
30 well-known special case and is the simplest form of these methods.

The biallelic markers of the present invention may be used in both parametric and non-parametric linkage analysis. Preferably biallelic markers may be used in non-parametric methods which allow the mapping of genes involved in complex traits. The biallelic markers of the present invention may be used in both IBD- and IBS- methods to map genes affecting a complex trait. In
35 such studies, taking advantage of the high density of biallelic markers, several adjacent biallelic marker loci may be pooled to achieve the efficiency attained by multi-allelic markers (Zhao *et al.*, 1998).

Population Association Studies

The present invention comprises methods for detecting an association between the PG-3 gene and a detectable trait using the biallelic markers of the present invention. In one embodiment the present invention comprises methods to detect an association between a biallelic marker allele
5 or a biallelic marker haplotype and a trait. Further, the invention comprises methods to identify a trait causing allele in linkage disequilibrium with any biallelic marker allele of the present invention.

As described above, alternative approaches can be employed to perform association studies: genome-wide association studies, candidate region association studies and candidate gene
10 association studies. In a preferred embodiment, the biallelic markers of the present invention are used to perform candidate gene association studies. The candidate gene analysis clearly provides a short-cut approach to the identification of genes and gene polymorphisms related to a particular trait when some information concerning the biology of the trait is available. Further, the biallelic markers of the present invention may be incorporated in any map of genetic markers of the human
15 genome in order to perform genome-wide association studies. Methods to generate a high-density map of biallelic markers has been described in US Provisional Patent application serial number 60/082,614. The biallelic markers of the present invention may further be incorporated in any map of a specific candidate region of the genome (a specific chromosome or a specific chromosomal segment for example).

20 As mentioned above, association studies may be conducted within the general population and are not limited to studies performed on related individuals in affected families. Association studies are extremely valuable as they permit the analysis of sporadic or multifactor traits. Moreover, association studies represent a powerful method for fine-scale mapping enabling much finer mapping of trait causing alleles than linkage studies. Studies based on pedigrees often only
25 narrow the location of the trait causing allele. Association studies using the biallelic markers of the present invention can therefore be used to refine the location of a trait causing allele in a candidate region identified by Linkage Analysis methods. Moreover, once a chromosome segment of interest has been identified, the presence of a candidate gene such as a candidate gene of the present invention, in the region of interest can provide a shortcut to the identification of the trait causing
30 allele. Biallelic markers of the present invention can be used to demonstrate that a candidate gene is associated with a trait. Such uses are specifically contemplated in the present invention.

Determining The Frequency Of A Biallelic Marker Allele Or Of A Biallelic Marker Haplotype In A Population

Association studies explore the relationships among frequencies for sets of alleles between
35 loci.

DETERMINING THE FREQUENCY OF AN ALLELE IN A POPULATION

Allelic frequencies of the biallelic markers in a populations can be determined using one of the methods described above under the heading "Methods for genotyping an individual for biallelic markers", or any genotyping procedure suitable for this intended purpose. Genotyping pooled samples or individual samples can determine the frequency of a biallelic marker allele in a
5 population. One way to reduce the number of genotypings required is to use pooled samples. A drawback in using pooled samples is in terms of accuracy and reproducibility for determining accurate DNA concentrations in setting up the pools. Genotyping individual samples provides higher sensitivity, reproducibility and accuracy and; is the preferred method used in the present invention. Preferably, each individual is genotyped separately and simple gene counting is applied
10 to determine the frequency of an allele of a biallelic marker or of a genotype in a given population.

The invention also relates to methods of estimating the frequency of an allele in a population comprising: a) genotyping individuals from said population for said biallelic marker according to the method of the present invention; b) determining the proportional representation of said biallelic marker in said population. In addition, the methods of estimating the frequency of an
15 allele in a population of the invention encompass methods with any further limitation described in this disclosure, or those following, specified alone or in any combination; optionally, the PG-3-related biallelic marker is selected from the group consisting of A1 to A80, and the complements thereof, or optionally the biallelic marker is one of the biallelic markers in linkage disequilibrium therewith; optionally, wherein said PG-3-related biallelic marker is selected from the group
20 consisting of A1 to A5 and A8 to A80, and the complements thereof, or optionally the biallelic markers in linkage disequilibrium therewith; optionally, wherein said PG-3-related biallelic marker is selected from the group consisting of A6 and A7, and the complements thereof, or optionally the biallelic markers in linkage disequilibrium therewith; Optionally, the determination of the frequency of a biallelic marker allele in a population may be accomplished by determining the identity of the
25 nucleotides for both copies of said biallelic marker present in the genome of each individual in said population and calculating the proportional representation of said nucleotide at said PG-3-related biallelic marker for the population; Optionally, the determination of the proportional representation may be accomplished by performing a genotyping method of the invention on a pooled biological sample derived from a representative number of individuals, or each individual, in said population,
30 and calculating the proportional amount of said nucleotide compared with the total.

DETERMINING THE FREQUENCY OF A HAPLOTYPE IN A POPULATION

The gametic phase of haplotypes is unknown when diploid individuals are heterozygous at more than one locus. Using genealogical information in families gametic phase can sometimes be inferred (Perlin *et al.*, 1994). When no genealogical information is available different strategies
35 may be used. One possibility is that the multiple-site heterozygous diploids can be eliminated from the analysis, keeping only the homozygotes and the single-site heterozygote individuals, but this approach might lead to a possible bias in the sample composition and the underestimation of low-

frequency haplotypes. Another possibility is that single chromosomes can be studied independently, for example, by asymmetric PCR amplification (see Newton et al, 1989; Wu *et al.*, 1989) or by isolation of single chromosome by limit dilution followed by PCR amplification (see Ruano *et al.*, 1990). Further, a sample may be haplotyped for sufficiently close biallelic markers by

5 double PCR amplification of specific alleles (Sarkar, G. and Sommer S. S., 1991). These approaches are not entirely satisfying either because of their technical complexity, the additional cost they entail, their lack of generalization at a large scale, or the possible biases they introduce. To overcome these difficulties, an algorithm to infer the phase of PCR-amplified DNA genotypes introduced by Clark, A.G.(1990) may be used. Briefly, the principle is to start filling a preliminary

10 list of haplotypes present in the sample by examining unambiguous individuals, that is, the complete homozygotes and the single-site heterozygotes. Then other individuals in the same sample are screened for the possible occurrence of previously recognized haplotypes. For each positive identification, the complementary haplotype is added to the list of recognized haplotypes, until the phase information for all individuals is either resolved or identified as unresolved. This

15 method assigns a single haplotype to each multiheterozygous individual, whereas several haplotypes are possible when there are more than one heterozygous site. Alternatively, one can use methods estimating haplotype frequencies in a population without assigning haplotypes to each individual. Preferably, a method based on an expectation-maximization (EM) algorithm (Dempster *et al.*, 1977) leading to maximum-likelihood estimates of haplotype frequencies under the

20 assumption of Hardy-Weinberg proportions (random mating) is used (see Excoffier L. and Slatkin M., 1995). The EM algorithm is a generalized iterative maximum-likelihood approach to estimation that is useful when data are ambiguous and/or incomplete. The EM algorithm is used to resolve heterozygotes into haplotypes. Haplotype estimations are further described below under the heading "Statistical Methods." Any other method known in the art to determine or to estimate the

25 frequency of a haplotype in a population may be used.

The invention also encompasses methods of estimating the frequency of a haplotype for a set of biallelic markers in a population, comprising the steps of: a) genotyping at least one PG-3-related biallelic marker according to a method of the invention for each individual in said

30 population; b) genotyping a second biallelic marker by determining the identity of the nucleotides at said second biallelic marker for both copies of said second biallelic marker present in the genome of each individual in said population; and c) applying a haplotype determination method to the identities of the nucleotides determined in steps a) and b) to obtain an estimate of said frequency. In addition, the methods of estimating the frequency of a haplotype of the invention encompass methods with any further limitation described in this disclosure, or those following, alone or in any

35 combination: optionally, said PG-3-related biallelic marker is selected from the group consisting of A1 to A80, and the complements thereof, or optionally the biallelic markers in linkage disequilibrium therewith; optionally, wherein said PG-3-related biallelic marker is selected from the

group consisting of A1 to A5 and A8 to A80, and the complements thereof, or optionally the biallelic markers in linkage disequilibrium therewith; optionally, wherein said PG-3-related biallelic marker is selected from the group consisting of A6 and A7, and the complements thereof, or optionally the biallelic markers in linkage disequilibrium therewith; Optionally, said haplotype
5 determination method is performed by asymmetric PCR amplification, double PCR amplification of specific alleles, the Clark algorithm, or an expectation-maximization algorithm.

Linkage Disequilibrium Analysis

Linkage disequilibrium is the non-random association of alleles at two or more loci and represents a powerful tool for mapping genes involved in disease traits (see Ajioka R.S. *et al.*,
10 1997). Biallelic markers, because they are densely spaced in the human genome and can be genotyped in greater numbers than other types of genetic markers (such as RFLP or VNTR markers), are particularly useful in genetic analysis based on linkage disequilibrium.

When a disease mutation is first introduced into a population (by a new mutation or the immigration of a mutation carrier), it necessarily resides on a single chromosome and thus on a
15 single "background" or "ancestral" haplotype of linked markers. Consequently, there is complete disequilibrium between these markers and the disease mutation: one finds the disease mutation only in the presence of a specific set of marker alleles. Through subsequent generations recombination events occur between the disease mutation and these marker polymorphisms, and the disequilibrium gradually dissipates. The pace of this dissipation is a function of the recombination frequency, so
20 the markers closest to the disease gene will manifest higher levels of disequilibrium than those that are further away. When not broken up by recombination, "ancestral" haplotypes and linkage disequilibrium between marker alleles at different loci can be tracked not only through pedigrees but also through populations. Linkage disequilibrium is usually seen as an association between one specific allele at one locus and another specific allele at a second locus.

25 The pattern or curve of disequilibrium between disease and marker loci is expected to exhibit a maximum that occurs at the disease locus. Consequently, the amount of linkage disequilibrium between a disease allele and closely linked genetic markers may yield valuable information regarding the location of the disease gene. For fine-scale mapping of a disease locus, it is useful to have some knowledge of the patterns of linkage disequilibrium that exist between
30 markers in the studied region. As mentioned above the mapping resolution achieved through the analysis of linkage disequilibrium is much higher than that of linkage studies. The high density of biallelic markers combined with linkage disequilibrium analysis provides powerful tools for fine-scale mapping. Different methods to calculate linkage disequilibrium are described below under the heading "Statistical Methods".

35 Population-Based Case-Control Studies Of Trait-Marker Associations

As mentioned above, the occurrence of pairs of specific alleles at different loci on the same chromosome is not random and the deviation from random is called linkage disequilibrium.

Association studies focus on population frequencies and rely on the phenomenon of linkage disequilibrium. If a specific allele in a given gene is directly involved in causing a particular trait, its frequency will be statistically increased in an affected (trait positive) population, when compared to the frequency in a trait negative population or in a random control population. As a consequence of the existence of linkage disequilibrium, the frequency of all other alleles present in the haplotype carrying the trait-causing allele will also be increased in trait positive individuals compared to trait negative individuals or random controls. Therefore, association between the trait and any allele (specifically a biallelic marker allele) in linkage disequilibrium with the trait-causing allele will suffice to suggest the presence of a trait-related gene in that particular region. Case-control populations can be genotyped for biallelic markers to identify associations that narrowly locate a trait causing allele. As any marker in linkage disequilibrium with one given marker associated with a trait will be associated with the trait. Linkage disequilibrium allows the relative frequencies in case-control populations of a limited number of genetic polymorphisms (specifically biallelic markers) to be analyzed as an alternative to screening all possible functional polymorphisms in order to find trait-causing alleles. Association studies compare the frequency of marker alleles in unrelated case-control populations, and represent powerful tools for the dissection of complex traits.

CASE-CONTROL POPULATIONS (INCLUSION CRITERIA)

Population-based association studies do not concern familial inheritance but compare the prevalence of a particular genetic marker, or a set of markers, in case-control populations. They are case-control studies based on comparison of unrelated case (affected or trait positive) individuals and unrelated control (unaffected, trait negative or random) individuals. Preferably the control group is composed of unaffected or trait negative individuals. Further, the control group is ethnically matched to the case population. Moreover, the control group is preferably matched to the case-population for the main known confusion factor for the trait under study (for example age-matched for an age-dependent trait). Ideally, individuals in the two samples are paired in such a way that they are expected to differ only in their disease status. The terms "trait positive population", "case population" and "affected population" are used interchangeably herein.

An important step in the dissection of complex traits using association studies is the choice of case-control populations (see Lander and Schork, 1994). A major step in the choice of case-control populations is the clinical definition of a given trait or phenotype. Any genetic trait may be analyzed by the association method proposed here by carefully selecting the individuals to be included in the trait positive and trait negative phenotypic groups. Four criteria are often useful: clinical phenotype, age at onset, family history and severity. The selection procedure for continuous or quantitative traits (such as blood pressure for example) involves selecting individuals at opposite ends of the phenotype distribution of the trait under study, so as to include in these trait positive and trait negative populations individuals with non-overlapping phenotypes. Preferably, case-control populations consist of phenotypically homogeneous populations. Trait positive and

More particularly, the present invention relates to expression vectors which include nucleic acids encoding a PG-3 protein, preferably the PG-3 protein of the amino acid sequence of SEQ ID No 3 or variants or fragments thereof.

The invention also pertains to a recombinant expression vector useful for the expression of
5 the PG-3 coding sequence, wherein said vector comprises a nucleic acid of SEQ ID No 2.

Recombinant vectors comprising a nucleic acid containing a PG-3-related biallelic marker are also part of the invention. In a preferred embodiment, said biallelic marker is selected from the group consisting of A1 to A80, and the complements thereof.

Some of the elements which can be found in the vectors of the present invention are
10 described in further detail in the following sections.

The present invention also encompasses primary, secondary, and immortalized homologously recombinant host cells of vertebrate origin, preferably mammalian origin and particularly human origin, that have been engineered to: a) insert exogenous (heterologous) polynucleotides into the endogenous chromosomal DNA of a targeted gene, b) delete endogenous
15 chromosomal DNA, and/or c) replace endogenous chromosomal DNA with exogenous polynucleotides. Insertions, deletions, and/or replacements of polynucleotide sequences may be to the coding sequences of the targeted gene and/or to regulatory regions, such as promoter and enhancer sequences, operably associated with the targeted gene.

The present invention further relates to a method of making a homologously recombinant
20 host cell in vitro or in vivo, wherein the expression of a targeted gene not normally expressed in the cell is altered. Preferably the alteration causes expression of the targeted gene under normal growth conditions or under conditions suitable for producing the polypeptide encoded by the targeted gene. The method comprises the steps of: (a) transfecting the cell in vitro or in vivo with a polynucleotide construct, the polynucleotide construct comprising: (i) a targeting sequence; (ii) a regulatory
25 sequence and/or a coding sequence; and (iii) an unpaired splice donor site, if necessary, thereby producing a transfected cell; and (b) maintaining the transfected cell in vitro or in vivo under conditions appropriate for homologous recombination.

The present invention further relates to a method of altering the expression of a targeted gene in a cell in vitro or in vivo wherein the gene is not normally expressed in the cell, comprising
30 the steps of: (a) transfecting the cell in vitro or in vivo with a a polynucleotide construct, the a polynucleotide construct comprising: (i) a targeting sequence; (ii) a regulatory sequence and/or a coding sequence; and (iii) an unpaired splice donor site, if necessary, thereby producing a transfected cell; and (b) maintaining the transfected cell in vitro or in vivo under conditions appropriate for homologous recombination, thereby producing a homologously recombinant cell;
35 and (c) maintaining the homologously recombinant cell in vitro or in vivo under conditions appropriate for expression of the gene.

trait negative populations consist of phenotypically uniform populations of individuals representing each between 1 and 98%, preferably between 1 and 80%, more preferably between 1 and 50%, and more preferably between 1 and 30%, most preferably between 1 and 20% of the total population under study, and preferably selected among individuals exhibiting non-overlapping phenotypes.

- 5 The clearer the difference between the two trait phenotypes, the greater the probability of detecting an association with biallelic markers. The selection of those drastically different but relatively uniform phenotypes enables efficient comparisons in association studies and the possible detection of marked differences at the genetic level, provided that the sample sizes of the populations under study are significant enough.

- 10 In preferred embodiments, a first group of between 50 and 300 trait positive individuals, preferably about 100 individuals, are recruited according to their phenotypes. A similar number of control individuals are included in such studies.

ASSOCIATION ANALYSIS

- The invention also comprises methods of detecting an association between a genotype and a
15 phenotype, comprising the steps of: a) determining the frequency of at least one PG-3-related biallelic marker in a trait positive population according to a genotyping method of the invention; b) determining the frequency of said PG-3-related biallelic marker in a control population according to a genotyping method of the invention; and c) determining whether a statistically significant association exists between said genotype and said phenotype. In addition, the methods of detecting
20 an association between a genotype and a phenotype of the invention encompass methods with any further limitation described in this disclosure, or those following, specified alone or in any combination: optionally, wherein said PG-3-related biallelic marker is selected from the group consisting of A1 to A80, and the complements thereof, or optionally the biallelic markers in linkage disequilibrium therewith; optionally, wherein said PG-3-related biallelic marker is selected from the
25 group consisting of A1 to A5 and A8 to A80, and the complements thereof, or optionally the biallelic markers in linkage disequilibrium therewith; optionally, wherein said PG-3-related biallelic marker is selected from the group consisting of A6 and A7, and the complements thereof, or optionally the biallelic markers in linkage disequilibrium therewith; Optionally, said control population may be a trait negative population, or a random population; Optionally, each of said
30 genotyping steps a) and b) may be performed on a pooled biological sample derived from each of said populations; Optionally, each of said genotyping of steps a) and b) is performed separately on biological samples derived from each individual in said population or a subsample thereof; Optionally, said trait is cancer susceptibility.

- The general strategy to perform association studies using biallelic markers derived from a
35 region carrying a candidate gene is to scan two groups of individuals (case-control populations) in order to measure and statistically compare the allele frequencies of the biallelic markers of the present invention in both groups.

If a statistically significant association with a trait is identified for at least one or more of the analyzed biallelic markers, one can assume that: either the associated allele is directly responsible for causing the trait (*i.e.* the associated allele is the trait causing allele), or more likely the associated allele is in linkage disequilibrium with the trait causing allele. The specific characteristics of the associated allele with respect to the candidate gene function usually give further insight into the relationship between the associated allele and the trait (causal or in linkage disequilibrium). If the evidence indicates that the associated allele within the candidate gene is most probably not the trait causing allele but is in linkage disequilibrium with the real trait causing allele, then the trait causing allele can be found by sequencing the vicinity of the associated marker, and performing further association studies with the polymorphisms that are revealed in an iterative manner.

Association studies are usually run in two successive steps. In a first phase, the frequencies of a reduced number of biallelic markers from the candidate gene are determined in the trait positive and control populations. In a second phase of the analysis, the position of the genetic loci responsible for the given trait is further refined using a higher density of markers from the relevant region. However, if the candidate gene under study is relatively small in length, as is the case for PG-3, a single phase may be sufficient to establish significant associations.

HAPLOTYPE ANALYSIS

As described above, when a chromosome carrying a disease allele first appears in a population as a result of either mutation or migration, the mutant allele necessarily resides on a chromosome having a set of linked markers: the ancestral haplotype. This haplotype can be tracked through populations and its statistical association with a given trait can be analyzed.

Complementing single point (allelic) association studies with multi-point association studies also called haplotype studies increases the statistical power of association studies. Thus, a haplotype association study allows one to define the frequency and the type of the ancestral carrier haplotype. A haplotype analysis is important in that it increases the statistical power of an analysis involving individual markers.

In a first stage of a haplotype frequency analysis, the frequency of the possible haplotypes based on various combinations of the identified biallelic markers of the invention is determined. The haplotype frequency is then compared for distinct populations of trait positive and control individuals. The number of trait positive individuals, which should be, subjected to this analysis to obtain statistically significant results usually ranges between 30 and 300, with a preferred number of individuals ranging between 50 and 150. The same considerations apply to the number of unaffected individuals (or random control) used in the study. The results of this first analysis provide haplotype frequencies in case-control populations, for each evaluated haplotype frequency a p-value and an odd ratio are calculated. If a statistically significant association is found the relative

risk for an individual carrying the given haplotype of being affected with the trait under study can be approximated.

An additional embodiment of the present invention encompasses methods of detecting an association between a haplotype and a phenotype, comprising the steps of: a) estimating the frequency of at least one haplotype in a trait positive population, according to a method of the invention for estimating the frequency of a haplotype; b) estimating the frequency of said haplotype in a control population, according to a method of the invention for estimating the frequency of a haplotype; and c) determining whether a statistically significant association exists between said haplotype and said phenotype. In addition, the methods of detecting an association between a haplotype and a phenotype of the invention encompass methods with any further limitation described in this disclosure, or those following: optionally, said PG-3-related biallelic marker is selected from the group consisting of A1 to A80, and the complements thereof, or optionally the biallelic markers in linkage disequilibrium therewith; optionally, wherein said PG-3-related biallelic marker is selected from the group consisting of A1 to A5 and A8 to A80, and the complements thereof, or optionally the biallelic markers in linkage disequilibrium therewith; optionally, wherein said PG-3-related biallelic marker is selected from the group consisting of A6 and A7, and the complements thereof, or optionally the biallelic markers in linkage disequilibrium therewith; Optionally, said control population is a trait negative population, or a random population. Optionally, said method comprises the additional steps of determining the phenotype in said trait positive and said control populations prior to step c); optionally, said trait is cancer susceptibility.

INTERACTION ANALYSIS

The biallelic markers of the present invention may also be used to identify patterns of biallelic markers associated with detectable traits resulting from polygenic interactions. The analysis of genetic interaction between alleles at unlinked loci requires individual genotyping using the techniques described herein. The analysis of allelic interaction among a selected set of biallelic markers with an appropriate level of statistical significance can be considered as a haplotype analysis. Interaction analysis consists in stratifying the case-control populations with respect to a given haplotype for the first loci and performing a haplotype analysis with the second loci with each subpopulation.

Statistical methods used in association studies are further described below.

Testing For Linkage In The Presence Of Association

The biallelic markers of the present invention may further be used in TDT (transmission/disequilibrium test). TDT tests for both linkage and association and is not affected by population stratification. TDT requires data for affected individuals and their parents or data from unaffected sibs instead of from parents (see Spielmann S. *et al.*, 1993; Schaid D.J. *et al.*, 1996, Spielmann S. and Ewens W.J., 1998). Such combined tests generally reduce the false – positive errors produced by separate analyses.

STATISTICAL METHODS

In general, any method known in the art to test whether a trait and a genotype show a statistically significant correlation may be used.

1) Methods In Linkage Analysis

5 Statistical methods and computer programs useful for linkage analysis are well-known to those skilled in the art (see Terwilliger J.D. and Ott J., 1994; Ott J., 1991).

2) Methods To Estimate Haplotype Frequencies In A Population

As described above, when genotypes are scored, it is often not possible to distinguish heterozygotes so that haplotype frequencies cannot be easily inferred. When the gametic phase is
 10 not known, haplotype frequencies can be estimated from the multilocus genotypic data. Any method known to person skilled in the art can be used to estimate haplotype frequencies (see Lange K., 1997; Weir, B.S., 1996) Preferably, maximum-likelihood haplotype frequencies are computed using an Expectation- Maximization (EM) algorithm (see Dempster *et al.*, 1977; Excoffier L. and Slatkin M., 1995). This procedure is an iterative process aiming at obtaining maximum-likelihood
 15 estimates of haplotype frequencies from multi-locus genotype data when the gametic phase is unknown. Haplotype estimations are usually performed by applying the EM algorithm using for example the EM-HAPLO program (Hawley M. E. *et al.*, 1994) or the Arlequin program (Schneider *et al.*, 1997). The EM algorithm is a generalized iterative maximum likelihood approach to estimation and is briefly described below.

20 Please note that in the present section, "Methods To Estimate Haplotype Frequencies In A Population, ", phenotypes will refer to multi-locus genotypes with unknown haplotypic phase. Genotypes will refer to mutli-locus genotypes with known haplotypic phase.

Suppose one has a sample of N unrelated individuals typed for K markers. The data observed are the unknown-phase K-locus phenotypes that can be categorized with F different
 25 phenotypes. Further, suppose that we have H possible haplotypes (in the case of K biallelic markers, we have for the maximum number of possible haplotypes $H=2^K$).

For phenotype j with c_j possible genotypes, we have:

$$P_j = \sum_{i=1}^{c_j} P(\text{genotype}(i)) = \sum_{i=1}^{c_j} P(h_k, h_l). \quad \text{Equation 1}$$

Here, P_j is the probability of the j^{th} phenotype, and $P(h_k, h_l)$ is the probability of the i^{th}
 30 genotype composed of haplotypes h_k and h_l . Under random mating (*i.e.* Hardy-Weinberg Equilibrium), $P(h_k, h_l)$ is expressed as:

$$\begin{aligned} P(h_k, h_l) &= P(h_k)^2 \text{ for } h_k = h_l, \text{ and} \\ P(h_k, h_l) &= 2P(h_k)P(h_l) \text{ for } h_k \neq h_l. \end{aligned} \quad \text{Equation 2}$$

The E-M algorithm is composed of the following steps: First, the genotype frequencies are
 35 estimated from a set of initial values of haplotype frequencies. These haplotype frequencies are

denoted $P_1^{(0)}, P_2^{(0)}, P_3^{(0)}, \dots, P_H^{(0)}$. The initial values for the haplotype frequencies may be obtained from a random number generator or in some other way well known in the art. This step is referred to the Expectation step. The next step in the method, called the Maximization step, consists of using the estimates for the genotype frequencies to re-calculate the haplotype frequencies. The first iteration haplotype frequency estimates are denoted by $P_1^{(1)}, P_2^{(1)}, P_3^{(1)}, \dots, P_H^{(1)}$. In general, the Expectation step at the s^{th} iteration consists of calculating the probability of placing each phenotype into the different possible genotypes based on the haplotype frequencies of the previous iteration:

$$P(h_k, h_l)^{(s)} = \frac{n_j}{N} \left[\frac{P_j(h_k, h_l)^{(s)}}{P_j} \right], \quad \text{Equation 3}$$

where n_j is the number of individuals with the j^{th} phenotype and $P_j(h_k, h_l)^{(s)}$ is the probability of genotype $h_k h_l$ in phenotype j . In the Maximization step, which is equivalent to the gene-counting method (Smith, 1957), the haplotype frequencies are re-estimated based on the genotype estimates:

$$P_i^{(s+1)} = \frac{1}{2} \sum_{j=1}^F \sum_{i=1}^{c_j} \delta_{ii} P_j(h_k, h_l)^{(s)}. \quad \text{Equation 4}$$

Here, δ_{ii} is an indicator variable which counts the number of occurrences that haplotype i is present in i^{th} genotype; it takes on values 0, 1, and 2.

The E-M iterations cease when the following criterion has been reached. Using Maximum Likelihood Estimation (MLE) theory, one assumes that the phenotypes j are distributed multinomially. At each iteration s , one can compute the likelihood function L . Convergence is achieved when the difference of the log-likelihood between two consecutive iterations is less than some small number, preferably 10^{-7} .

3) Methods To Calculate Linkage Disequilibrium Between Markers

A number of methods can be used to calculate linkage disequilibrium between any two genetic positions, in practice linkage disequilibrium is measured by applying a statistical association test to haplotype data taken from a population.

Linkage disequilibrium between any pair of biallelic markers comprising at least one of the biallelic markers of the present invention (M_i, M_j) having alleles (a_i/b_i) at marker M_i and alleles (a_j/b_j) at marker M_j can be calculated for every allele combination ($a_i, a_j; a_i, b_j; b_i, a_j$ and b_i, b_j), according to the Piazza formula:

$$\Delta_{a_i a_j} = \sqrt{\theta 4} - \sqrt{(\theta 4 + \theta 3)(\theta 4 + \theta 2)}, \text{ where:}$$

$\theta 4 = --$ = frequency of genotypes not having allele a_i at M_i and not having allele a_j at M_j

$\theta 3 = - +$ = frequency of genotypes not having allele a_i at M_i and having allele a_j at M_j

$\theta 2 = + -$ = frequency of genotypes having allele a_i at M_i and not having allele a_j at M_j

Linkage disequilibrium (LD) between pairs of biallelic markers (M_i, M_j) can also be calculated for every allele combination ($a_i, a_j; a_i, b_j; b_i, a_j$ and b_i, b_j), according to the maximum-

likelihood estimate (MLE) for delta (the composite genotypic disequilibrium coefficient), as described by Weir (Weir B. S., 1996). The MLE for the composite linkage disequilibrium is:

$$D_{aiaj} = (2n_1 + n_2 + n_3 + n_4/2)/N - 2(pr(a_i) \cdot pr(a_j))$$

Where $n_1 = \Sigma$ phenotype ($a_i/a_i, a_j/a_j$), $n_2 = \Sigma$ phenotype ($a_i/a_i, a_j/b_j$), $n_3 = \Sigma$ phenotype ($a_i/b_i, a_j/a_j$), $n_4 = \Sigma$ phenotype ($a_i/b_i, a_j/b_j$) and N is the number of individuals in the sample.

This formula allows linkage disequilibrium between alleles to be estimated when only genotype, and not haplotype, data are available.

Another means of calculating the linkage disequilibrium between markers is as follows. For a couple of biallelic markers, $M_i (a_i/b_i)$ and $M_j (a_j/b_j)$, fitting the Hardy-Weinberg equilibrium, one can estimate the four possible haplotype frequencies in a given population according to the approach described above.

The estimation of gametic disequilibrium between a_i and a_j is simply:

$$D_{aiaj} = pr(haplotype(a_i, a_j)) - pr(a_i) \cdot pr(a_j).$$

Where $pr(a_i)$ is the probability of allele a_i and $pr(a_j)$ is the probability of allele a_j and where $pr(haplotype(a_i, a_j))$ is estimated as in Equation 3 above.

For a couple of biallelic marker only one measure of disequilibrium is necessary to describe the association between M_i and M_j .

Then a normalized value of the above is calculated as follows:

$$D'_{aiaj} = D_{aiaj} / \max(-pr(a_i) \cdot pr(a_j), -pr(b_i) \cdot pr(b_j)) \text{ with } D_{aiaj} < 0$$

$$D'_{aiaj} = D_{aiaj} / \max(pr(b_i) \cdot pr(a_j), pr(a_i) \cdot pr(b_j)) \text{ with } D_{aiaj} > 0$$

The skilled person will readily appreciate that other linkage disequilibrium calculation methods can be used.

Linkage disequilibrium among a set of biallelic markers having an adequate heterozygosity rate can be determined by genotyping between 50 and 1000 unrelated individuals, preferably between 75 and 200, more preferably around 100.

4) Testing For Association

Methods for determining the statistical significance of a correlation between a phenotype and a genotype, in this case an allele at a biallelic marker or a haplotype made up of such alleles, may be determined by any statistical test known in the art and with any accepted threshold of statistical significance being required. The application of particular methods and thresholds of significance are well within the skill of the ordinary practitioner of the art.

Testing for association is performed by determining the frequency of a biallelic marker allele in case and control populations and comparing these frequencies with a statistical test to determine if there is a statistically significant difference in frequency which would indicate a correlation between the trait and the biallelic marker allele under study. Similarly, a haplotype analysis is performed by estimating the frequencies of all possible haplotypes for a given set of biallelic markers in case and control populations, and comparing these frequencies with a statistical

test to determine if there is a statistically significant correlation between the haplotype and the phenotype (trait) under study. Any statistical tool useful to test for a statistically significant association between a genotype and a phenotype may be used. Preferably the statistical test employed is a chi-square test with one degree of freedom. A P-value is calculated (the P-value is the probability that a statistic as large or larger than the observed one would occur by chance).

STATISTICAL SIGNIFICANCE

In preferred embodiments, significance for diagnosis purposes, either as a positive basis for further diagnostic tests or as a preliminary starting point for early preventive therapy, the p value related to a biallelic marker association is preferably about 1×10^{-2} or less, more preferably about 1×10^{-4} or less, for a single biallelic marker analysis and about 1×10^{-3} or less, still more preferably 1×10^{-6} or less and most preferably of about 1×10^{-8} or less, for a haplotype analysis involving two or more markers. These values are believed to be applicable to any association studies involving single or multiple marker combinations.

The skilled person can use the range of values set forth above as a starting point in order to carry out association studies with biallelic markers of the present invention. In doing so, significant associations between the biallelic markers of the present invention and a trait can be revealed and used for diagnosis and drug screening purposes.

PHENOTYPIC PERMUTATION

In order to confirm the statistical significance of the first stage haplotype analysis described above, it might be suitable to perform further analyses in which genotyping data from case-control individuals are pooled and randomized with respect to the trait phenotype. Each individual genotyping data is randomly allocated to two groups, which contain the same number of individuals as the case-control populations used to compile the data obtained in the first stage. A second stage haplotype analysis is preferably run on these artificial groups, preferably for the markers included in the haplotype of the first stage analysis showing the highest relative risk coefficient. This experiment is reiterated preferably at least between 100 and 10000 times. The repeated iterations allow the determination of the probability to obtain the tested haplotype by chance.

ASSESSMENT OF STATISTICAL ASSOCIATION

To address the problem of false positives similar analysis may be performed with the same case-control populations in random genomic regions. Results in random regions and the candidate region are compared as described in a co-pending US Provisional Patent Application entitled "Methods, Software And Apparati For Identifying Genomic Regions Harboring A Gene Associated With A Detectable Trait," U.S. Serial Number 60/107,986, filed November 10, 1998, and a second U.S. Provisional Patent Application also entitled "Methods, Software And Apparati For Identifying Genomic Regions Harboring A Gene Associated With A Detectable Trait," U.S. Serial Number 60/140,785, filed June 23, 1999.

5) Evaluation Of Risk Factors

The association between a risk factor (in genetic epidemiology the risk factor is the presence or the absence of a certain allele or haplotype at marker loci) and a disease is measured by the odds ratio (OR) and by the relative risk (RR). If $P(R^+)$ is the probability of developing the disease for individuals with R and $P(R^-)$ is the probability for individuals without the risk factor,

5 then the relative risk is simply the ratio of the two probabilities, that is:

$$RR = P(R^+)/P(R^-)$$

In case-control studies, direct measures of the relative risk cannot be obtained because of the sampling design. However, the odds ratio allows a good approximation of the relative risk for low-incidence diseases and can be calculated:

$$OR = \left[\frac{F^+}{1-F^+} \right] / \left[\frac{F^-}{(1-F^-)} \right]$$

10 $OR = (F^+/(1-F^+))/(F^-/(1-F^-))$

F^+ is the frequency of the exposure to the risk factor in cases and F^- is the frequency of the exposure to the risk factor in controls. F^+ and F^- are calculated using the allelic or haplotype frequencies of the study and further depend on the underlying genetic model (dominant, recessive, additive...).

15 One can further estimate the attributable risk (AR) which describes the proportion of individuals in a population exhibiting a trait due to a given risk factor. This measure is important in quantifying the role of a specific factor in disease etiology and in terms of the public health impact of a risk factor. The public health relevance of this measure lies in estimating the proportion of cases of disease in the population that could be prevented if the exposure of interest were absent.

20 AR is determined as follows:

$$AR = P_E (RR-1) / (P_E (RR-1)+1)$$

AR is the risk attributable to a biallelic marker allele or a biallelic marker haplotype. P_E is the frequency of exposure to an allele or a haplotype within the population at large; and RR is the relative risk which, is approximated with the odds ratio when the trait under study has a relatively

25 low incidence in the general population.

IDENTIFICATION OF BIALLELIC MARKERS IN LINKAGE DISEQUILIBRIUM WITH THE BIALLELIC MARKERS OF THE INVENTION

Once a first biallelic marker has been identified in a genomic region of interest, the practitioner of ordinary skill in the art, using the teachings of the present invention, can easily

30 identify additional biallelic markers in linkage disequilibrium with this first marker. As mentioned before, any marker in linkage disequilibrium with a first marker associated with a trait will be associated with the trait. Therefore, once an association has been demonstrated between a given biallelic marker and a trait, the discovery of additional biallelic markers associated with this trait is of great interest in order to increase the density of biallelic markers in this particular region. The

causal gene or mutation will be found in the vicinity of the marker or set of markers showing the highest correlation with the trait.

Identification of additional markers in linkage disequilibrium with a given marker involves:

- (a) amplifying a genomic fragment comprising a first biallelic marker from a plurality of
5 individuals; (b) identifying of second biallelic markers in the genomic region harboring said first biallelic marker; (c) conducting a linkage disequilibrium analysis between said first biallelic marker and second biallelic markers; and (d) selecting said second biallelic markers as being in linkage disequilibrium with said first marker. Subcombinations comprising steps (b) and (c) are also contemplated.

- 10 Methods to identify biallelic markers and to conduct linkage disequilibrium analysis are described herein and can be carried out by the skilled person without undue experimentation. The present invention then also concerns biallelic markers which are in linkage disequilibrium with the biallelic markers A1 to A80 and which are expected to present similar characteristics in terms of their respective association with a given trait.

15 **IDENTIFICATION OF FUNCTIONAL MUTATIONS**

- Mutations in the PG-3 gene which are responsible for a detectable phenotype or trait may be identified by comparing the sequences of the PG-3 gene from trait positive and control individuals. Once a positive association is confirmed with a biallelic marker of the present invention, the identified locus can be scanned for mutations. In a preferred embodiment, functional
20 regions such as exons and splice sites, promoters and other regulatory regions of the PG-3 gene are scanned for mutations. In a preferred embodiment the sequence of the PG-3 gene is compared in trait positive and control individuals. Preferably, trait positive individuals carry the haplotype shown to be associated with the trait and trait negative individuals do not carry the haplotype or allele associated with the trait. The detectable trait or phenotype may comprise a variety of
25 manifestations of altered PG-3 function.

The mutation detection procedure is essentially similar to that used for biallelic marker identification. The method used to detect such mutations generally comprises the following steps:

- amplification of a region of the PG-3 gene comprising a biallelic marker or a group of biallelic markers associated with the trait from DNA samples of trait positive patients and trait-
30 negative controls;
- sequencing of the amplified region;
- comparison of DNA sequences from trait positive and control individuals;
- determination of mutations specific to trait-positive patients.

- In one embodiment, said biallelic marker is selected from the group consisting of A1 to
35 A80, and the complements thereof. It is preferred that candidate polymorphisms be then verified by screening a larger population of cases and controls by means of any genotyping procedure such as those described herein, preferably using a microsequencing technique in an individual test format.

Polymorphisms are considered as candidate mutations when present in cases and controls at frequencies compatible with the expected association results. Polymorphisms are considered as candidate "trait-causing" mutations when they exhibit a statistically significant correlation with the detectable phenotype.

5

RECOMBINANT VECTORS

The term "vector" is used herein to designate either a circular or a linear DNA or RNA molecule, which is either double-stranded or single-stranded, and which comprise at least one polynucleotide of interest that is sought to be transferred in a cell host or in a unicellular or multicellular host organism.

10

The present invention encompasses a family of recombinant vectors that comprise a regulatory polynucleotide derived from the PG-3 genomic sequence, and/or a coding polynucleotide from either the PG-3 genomic sequence or the cDNA sequence.

Generally, a recombinant vector of the invention may comprise any of the polynucleotides described herein, including regulatory sequences, coding sequences and polynucleotide constructs, as well as any PG-3 primer or probe as defined above. More particularly, the recombinant vectors of the present invention can comprise any of the polynucleotides described in the "Genomic Sequences Of The PG3 Gene" section, the "PG-3 cDNA Sequences" section, the "Coding Regions" section, the "Polynucleotide constructs" section, and the "Oligonucleotide Probes And Primers" section.

20

In a first preferred embodiment, a recombinant vector of the invention is used to amplify the inserted polynucleotide derived from a PG-3 genomic sequence of SEQ ID No 1 or a PG-3 cDNA, for example the cDNA of SEQ ID No 2 in a suitable cell host, this polynucleotide being amplified at every time that the recombinant vector replicates.

A second preferred embodiment of the recombinant vectors according to the invention comprises expression vectors comprising either a regulatory polynucleotide or a coding nucleic acid of the invention, or both. Within certain embodiments, expression vectors are employed to express the PG-3 polypeptide, which can then be purified and, for example be used in ligand screening assays or as an immunogen in order to raise specific antibodies directed against the PG-3 protein. In other embodiments, the expression vectors are used for constructing transgenic animals and also for gene therapy. Expression requires that appropriate signals are provided in the vectors, said signals including various regulatory elements, such as enhancers/promoters from both viral and mammalian sources that drive expression of the genes of interest in host cells. Dominant drug selection markers for establishing permanent, stable cell clones expressing the products are generally included in the expression vectors of the invention, as they are elements that link expression of the drug selection markers to expression of the polypeptide.

The present invention further relates to a method of making a polypeptide of the present invention by altering the expression of a targeted endogenous gene in a cell in vitro or in vivo wherein the gene is not normally expressed in the cell, comprising the steps of: a) transfecting the cell in vitro with a a polynucleotide construct, the a polynucleotide construct comprising: (i) a
5 targeting sequence; (ii) a regulatory sequence and/or a coding sequence; and (iii) an unpaired splice donor site, if necessary, thereby producing a transfected cell; (b) maintaining the transfected cell in vitro or in vivo under conditions appropriate for homologous recombination, thereby producing a homologously recombinant cell; and c) maintaining the homologously recombinant cell in vitro or in vivo under conditions appropriate for expression of the gene thereby making the polypeptide.

10 The present invention further relates to a polynucleotide construct which alters the expression of a targeted gene in a cell type in which the gene is not normally expressed. This occurs when the a polynucleotide construct is inserted into the chromosomal DNA of the target cell, wherein the a polynucleotide construct comprises: a) a targeting sequence; b) a regulatory sequence and/or coding sequence; and c) an unpaired splice-donor site, if necessary. Further included are a
15 polynucleotide constructs, as described above, wherein the construct further comprises a polynucleotide which encodes a polypeptide and is in-frame with the targeted endogenous gene after homologous recombination with chromosomal DNA.

The compositions may be produced, and methods performed, by techniques known in the art, such as those described in U.S. Patent Nos: 6,054,288; 6,048,729; 6,048,724; 6,048,524;
20 5,994,127; 5,968,502; 5,965,125; 5,869,239; 5,817,789; 5,783,385; 5,733,761; 5,641,670; 5,580,734 ; International Publication Nos: WO96/29411, WO 94/12650; and scientific articles including Koller *et al.*, 1989.

1. General features of the expression vectors of the invention

A recombinant vector according to the invention comprises, but is not limited to, a YAC
25 (Yeast Artificial Chromosome), a BAC (Bacterial Artificial Chromosome), a phage, a phagemid, a cosmid, a plasmid or even a linear DNA molecule which may consist of a chromosomal, non-chromosomal, semi-synthetic and synthetic DNA. Such a recombinant vector can comprise a transcriptional unit comprising an assembly of:

(1) a genetic element or elements having a regulatory role in gene expression, for example
30 promoters or enhancers. Enhancers are cis-acting elements of DNA, usually from about 10 to 300 bp in length that act on the promoter to increase the transcription.

(2) a structural or coding sequence which is transcribed into mRNA and eventually translated into a polypeptide, said structural or coding sequence being operably linked to the regulatory elements described in (1); and

35 (3) appropriate transcription initiation and termination sequences. Structural units intended for use in yeast or eukaryotic expression systems preferably include a leader sequence enabling extracellular secretion of translated protein by a host cell. Alternatively, when a recombinant

protein is expressed without a leader or transport sequence, it may include a N-terminal residue. This residue may or may not be subsequently cleaved from the expressed recombinant protein to provide a final product.

Generally, recombinant expression vectors will include origins of replication, selectable
5 markers permitting transformation of the host cell, and a promoter derived from a highly expressed gene to direct transcription of a downstream structural sequence. The heterologous structural sequence is assembled in appropriate phase with translation initiation and termination sequences, and preferably a leader sequence capable of directing secretion of the translated protein into the periplasmic space or the extracellular medium. In a specific embodiment wherein the vector is
10 adapted for transfecting and expressing desired sequences in mammalian host cells, preferred vectors will comprise an origin of replication in the desired host, a suitable promoter and enhancer, and also any necessary ribosome binding sites, polyadenylation signal, splice donor and acceptor sites, transcriptional termination sequences, and 5'-flanking non-transcribed sequences. DNA sequences derived from the SV40 viral genome, for example SV40 origin, early promoter,
15 enhancer, splice and polyadenylation signals may be used to provide the required non-transcribed genetic elements.

The *in vivo* expression of a PG-3 polypeptide of SEQ ID No 3 or fragments or variants thereof may be useful in order to correct a genetic defect related to the expression of the native gene in a host organism or to the production of a biologically inactive PG-3 protein.

20 Consequently, the present invention also deals with recombinant expression vectors mainly designed for the *in vivo* production of the PG-3 polypeptide of SEQ ID No 3 or fragments or variants thereof by the introduction of the appropriate genetic material in the organism of the patient to be treated. This genetic material may be introduced *in vitro* in a cell that has been previously extracted from the organism, the modified cell being subsequently reintroduced in the said
25 organism, directly *in vivo* into the appropriate tissue.

2. Regulatory Elements

PROMOTERS

The suitable promoter regions used in the expression vectors according to the present invention are chosen taking into account the cell host in which the heterologous gene has to be
30 expressed. The particular promoter employed to control the expression of a nucleic acid sequence of interest is not believed to be important, so long as it is capable of directing the expression of the nucleic acid in the targeted cell. Thus, where a human cell is targeted, it is preferable to position the nucleic acid coding region adjacent to and under the control of a promoter that is capable of being expressed in a human cell, such as, for example, a human or a viral promoter.

35 A suitable promoter may be heterologous with respect to the nucleic acid for which it controls the expression or alternatively can be endogenous to the native polynucleotide containing the coding sequence to be expressed. Additionally, the promoter is generally heterologous with

respect to the recombinant vector sequences within which the construct promoter/coding sequence has been inserted.

Promoter regions can be selected from any desired gene using, for example, CAT (chloramphenicol transferase) vectors and more preferably pKK232-8 and pCM7 vectors.

5 Preferred bacterial promoters are the LacI, LacZ, the T3 or T7 bacteriophage RNA polymerase promoters, the gpt, lambda PR, PL and trp promoters (EP 0036776), the polyhedrin promoter, or the p10 protein promoter from baculovirus (Kit Novagen) (Smith *et al.*, 1983; O'Reilly *et al.*, 1992), the lambda PR promoter or also the trc promoter.

Eukaryotic promoters include CMV immediate early, HSV thymidine kinase, early and late SV40, LTRs from retrovirus, and mouse metallothionein-L. Selection of a convenient vector and promoter is well within the level of ordinary skill in the art.

The choice of a promoter is well within the ability of a person skilled in the field of genetic engineering. For example, one may refer to the book of Sambrook *et al.* (1989) or also to the procedures described by Fuller *et al.* (1996).

15 OTHER REGULATORY ELEMENTS

Where a cDNA insert is employed, one will typically desire to include a polyadenylation signal to effect proper polyadenylation of the gene transcript. The nature of the polyadenylation signal is not believed to be crucial to the successful practice of the invention, and any such sequence may be employed such as human growth hormone and SV40 polyadenylation signals. Also contemplated as an element of the expression cassette is a terminator. These elements can serve to enhance message levels and to minimize read through from the cassette into other sequences.

3. Selectable Markers

Such markers would confer an identifiable change to the cell permitting easy identification of cells containing the expression construct. The selectable marker genes for selection of transformed host cells are preferably dihydrofolate reductase or neomycin resistance for eukaryotic cell culture, TRP1 for *S. cerevisiae* or tetracycline, rifampicin or ampicillin resistance in *E. coli*, or levan saccharase for mycobacteria, this latter marker being a negative selection marker.

4. Preferred Vectors.

BACTERIAL VECTORS

30 As a representative but non-limiting example, useful expression vectors for bacterial use can comprise a selectable marker and a bacterial origin of replication derived from commercially available plasmids comprising genetic elements of pBR322 (ATCC 37017). Such commercial vectors include, for example, pKK223-3 (Pharmacia, Uppsala, Sweden), and GEM1 (Promega Biotec, Madison, WI, USA).

35 Large numbers of other suitable vectors are known to those of skill in the art, and commercially available, such as the following bacterial vectors: pQE70, pQE60, pQE-9 (Qiagen), pbs, pD10, phagescript, psiX174, pbluescript SK, pbsks, pNH8A, pNH16A, pNH18A, pNH46A

(Stratagene); ptrc99a, pKK223-3, pKK233-3, pDR540, pRIT5 (Pharmacia); pWLNEO, pSV2CAT, pOG44, pXT1, pSG (Stratagene); pSVK3, pBPV, pMSG, pSVL (Pharmacia); pQE-30 (QIAexpress).

BACTERIOPHAGE VECTORS

5 The P1 bacteriophage vector may contain large inserts ranging from about 80 to about 100 kb.

The construction of P1 bacteriophage vectors such as p158 or p158/neo8 are notably described by Sternberg (1992, 1994). Recombinant P1 clones comprising PG-3 nucleotide sequences may be designed for inserting large polynucleotides of more than 40 kb (Linton *et al.*, 10 1993). To generate P1 DNA for transgenic experiments, a preferred protocol is the protocol described by McCormick *et al.* (1994). Briefly, *E. coli* (preferably strain NS3529) harboring the P1 plasmid are grown overnight in a suitable broth medium containing 25 µg/ml of kanamycin. The P1 DNA is prepared from the *E. coli* by alkaline lysis using the Qiagen Plasmid Maxi kit (Qiagen, Chatsworth, CA, USA), according to the manufacturer's instructions. The P1 DNA is purified from 15 the bacterial lysate on two Qiagen-tip 500 columns, using the washing and elution buffers contained in the kit. A phenol/chloroform extraction is then performed before precipitating the DNA with 70% ethanol. After solubilizing the DNA in TE (10 mM Tris-HCl, pH 7.4, 1 mM EDTA), the concentration of the DNA is assessed by spectrophotometry.

When the goal is to express a P1 clone comprising PG-3 nucleotide sequences in a 20 transgenic animal, typically in transgenic mice, it is desirable to remove vector sequences from the P1 DNA fragment, for example by cleaving the P1 DNA at rare-cutting sites within the P1 polylinker (*SfiI*, *NotI* or *SaI*). The P1 insert is then purified from vector sequences on a pulsed-field agarose gel, using methods similar using methods similar to those originally reported for the isolation of DNA from YACs (Schedl *et al.*, 1993a; Peterson *et al.*, 1993). At this stage, the 25 resulting purified insert DNA can be concentrated, if necessary, on a Millipore Ultrafree-MC Filter Unit (Millipore, Bedford, MA, USA – 30,000 molecular weight limit) and then dialyzed against microinjection buffer (10 mM Tris-HCl, pH 7.4; 250 µM EDTA) containing 100 mM NaCl, 30 µM spermine, 70 µM spermidine on a microdialysis membrane (type VS, 0.025 µM from Millipore). The intactness of the purified P1 DNA insert is assessed by electrophoresis on 1% agarose (Sea 30 Kem GTG; FMC Bio-products) pulse-field gel and staining with ethidium bromide.

BACULOVIRUS VECTORS

A suitable vector for the expression of the PG-3 polypeptide of SEQ ID No 3 or fragments or variants thereof is a baculovirus vector that can be propagated in insect cells and in insect cell lines. A specific suitable host vector system is the pVL1392/1393 baculovirus transfer vector 35 (Pharmingen) that is used to transfect the SF9 c II line (ATCC N^oCRL 1711) which is derived from *Spodoptera frugiperda*.

Other suitable vectors for the expression of the PG-3 polypeptide of SEQ ID No 3 or fragments or variants thereof in a baculovirus expression system include those described by Chai *et al.* (1993), Vlasak *et al.* (1983) and Lenhard *et al.* (1996).

VIRAL VECTORS

5 In one specific embodiment, the vector is derived from an adenovirus. Preferred adenovirus vectors according to the invention are those described by Feldman and Steg (1996) or Ohno *et al.* (1994). Another preferred recombinant adenovirus according to this specific embodiment of the present invention is the human adenovirus type 2 or 5 (Ad 2 or Ad 5) or an adenovirus of animal origin (French patent application N° FR-93.05954).

10 Retrovirus vectors and adeno-associated virus vectors are generally understood to be the recombinant gene delivery systems of choice for the transfer of exogenous polynucleotides *in vivo*, particularly to mammals, including humans. These vectors provide efficient delivery of genes into cells, and the transferred nucleic acids are stably integrated into the chromosomal DNA of the host.

Particularly preferred retroviruses for the preparation or construction of retroviral *in vitro* or
15 *in vitro* gene delivery vehicles of the present invention include retroviruses selected from the group consisting of Mink-Cell Focus Inducing Virus, Murine Sarcoma Virus, Reticuloendotheliosis virus and Rous Sarcoma virus. Particularly preferred Murine Leukemia Viruses include the 4070A and the 1504A viruses, Abelson (ATCC No VR-999), Friend (ATCC No VR-245), Gross (ATCC No VR-590), Rauscher (ATCC No VR-998) and Moloney Murine Leukemia Virus (ATCC No VR-
20 190; PCT Application No WO 94/24298). Particularly preferred Rous Sarcoma Viruses include Bryan high titer (ATCC Nos VR-334, VR-657, VR-726, VR-659 and VR-728). Other preferred retroviral vectors are those described in Roth *et al.* (1996), PCT Application No WO 93/25234, PCT Application No WO 94/06920, Roux *et al.*, 1989, Julan *et al.*, 1992 and Neda *et al.*, 1991.

Yet another viral vector system that is contemplated by the invention consists in the adeno-
25 associated virus (AAV). The adeno-associated virus is a naturally occurring defective virus that requires another virus, such as an adenovirus or a herpes virus, as a helper virus for efficient replication and a productive life cycle (Muzyczka *et al.*, 1992). It is also one of the few viruses that may integrate its DNA into non-dividing cells, and exhibits a high frequency of stable integration (Flotte *et al.*, 1992; Samulski *et al.*, 1989; McLaughlin *et al.*, 1989). One advantageous feature of
30 AAV derives from its reduced efficacy for transducing primary cells relative to transformed cells.

BAC VECTORS

The bacterial artificial chromosome (BAC) cloning system (Shizuya *et al.*, 1992) has been developed to stably maintain large fragments of genomic DNA (100-300 kb) in *E. coli*. A preferred BAC vector consists of pBeloBAC11 vector that has been described by Kim *et al.* (1996).
35 BAC libraries are prepared with this vector using size-selected genomic DNA that has been partially digested using enzymes that permit ligation into either the *Bam* HI or *Hind*III sites in the vector. Flanking these cloning sites are T7 and SP6 RNA polymerase transcription initiation sites

that can be used to generate end probes by either RNA transcription or PCR methods. After the construction of a BAC library in *E. coli*, BAC DNA is purified from the host cell as a supercoiled circle. Converting these circular molecules into a linear form precedes both size determination and introduction of the BACs into recipient cells. The cloning site is flanked by two *Not* I sites,

- 5 permitting cloned segments to be excised from the vector by *Not* I digestion. Alternatively, the DNA insert contained in the pBeloBAC11 vector may be linearized by treatment of the BAC vector with the commercially available enzyme lambda terminase that leads to the cleavage at the unique *cos*N site, but this cleavage method results in a full length BAC clone containing both the insert DNA and the BAC sequences.

10 5. Delivery Of The Recombinant Vectors

In order to effect expression of the polynucleotides and polynucleotide constructs of the invention, these constructs must be delivered into a cell. This delivery may be accomplished *in vitro*, as in laboratory procedures for transforming cell lines, or *in vivo* or *ex vivo*, as in the treatment of certain diseases states.

- 15 One mechanism is viral infection where the expression construct is encapsulated in an infectious viral particle.

Several non-viral methods for the transfer of polynucleotides into cultured mammalian cells are also contemplated by the present invention, and include, without being limited to, calcium phosphate precipitation (Graham *et al.*, 1973; Chen *et al.*, 1987;), DEAE-dextran (Gopal, 1985),
20 electroporation (Tur-Kaspa *et al.*, 1986; Potter *et al.*, 1984), direct microinjection (Harland *et al.*, 1985), DNA-loaded liposomes (Nicolau *et al.*, 1982; Fraley *et al.*, 1979), and receptor-mediated transfection (Wu and Wu, 1987; 1988). Some of these techniques may be successfully adapted for *in vivo* or *ex vivo* use.

- Once the expression polynucleotide has been delivered into the cell, it may be stably
25 integrated into the genome of the recipient cell. This integration may be in the cognate location and orientation via homologous recombination (gene replacement) or it may be integrated in a random, non specific location (gene augmentation). In yet further embodiments, the nucleic acid may be stably maintained in the cell as a separate, episomal segment of DNA. Such nucleic acid segments or "episomes" encode sequences sufficient to permit maintenance and replication independent of or
30 in synchronization with the host cell cycle.

- One specific embodiment for a method for delivering a protein or peptide to the interior of a cell of a vertebrate *in vivo* comprises the step of introducing a preparation comprising a physiologically acceptable carrier and a naked polynucleotide operatively coding for the polypeptide of interest into the interstitial space of a tissue comprising the cell, whereby the naked
35 polynucleotide is taken up into the interior of the cell and has a physiological effect. This is particularly applicable for transfer *in vitro* but it may be applied to *in vivo* as well.

Compositions for use *in vitro* and *in vivo* comprising a "naked" polynucleotide are described in PCT application N° WO 90/11092 (Vical Inc.), and also in PCT application No. WO 95/11307 (Institut Pasteur, INSERM, Université d'Ottawa), as well as in the articles of Tacson *et al.* (1996) and of Huygen *et al.* (1996).

5 In still another embodiment of the invention, the transfer of a naked polynucleotide of the invention, including a polynucleotide construct of the invention, into cells may be proceeded with a particle bombardment (biolistic), said particles being DNA-coated microprojectiles accelerated to a high velocity allowing them to pierce cell membranes and enter cells without killing them, such as described by Klein *et al.* (1987).

10 In a further embodiment, the polynucleotide of the invention may be entrapped in a liposome (Ghosh and Bacchawat, 1991; Wong *et al.*, 1980; Nicolau *et al.*, 1987)

In a specific embodiment, the invention provides a composition for the *in vivo* production of the PG-3 protein or polypeptide described herein. It comprises a naked polynucleotide operatively coding for this polypeptide, in solution in a physiologically acceptable carrier, and
15 suitable for introduction into a tissue to cause cells of the tissue to express the said protein or polypeptide.

The amount of vector to be injected to the desired host organism varies according to the site of injection. As an indicative dose, it will be injected between 0,1 and 100 µg of the vector in an animal body, preferably a mammal body, for example a mouse body.

20 In another embodiment of the vector according to the invention, it may be introduced *in vitro* in a host cell, preferably in a host cell previously harvested from the animal to be treated and more preferably a somatic cell such as a muscle cell. In a subsequent step, the cell that has been transformed with the vector coding for the desired PG-3 polypeptide or the desired fragment thereof is reintroduced into the animal body in order to deliver the recombinant protein within the body
25 either locally or systemically.

CELL HOSTS

Another object of the invention consists of a host cell that has been transformed or transfected with one of the polynucleotides described herein, and in particular a polynucleotide either comprising a PG-3 regulatory polynucleotide or the coding sequence for the PG-3
30 polypeptide in a polynucleotide selected from the group consisting of SEQ ID Nos 1 and 2 or a fragment or a variant thereof. Also included are host cells that are transformed (prokaryotic cells) or that are transfected (eukaryotic cells) with a recombinant vector such as one of those described above. More particularly, the cell hosts of the present invention can comprise any of the polynucleotides described in the "Genomic Sequences Of The PG3 Gene" section, the "PG-3 cDNA
35 Sequences" section, the "Coding Regions" section, the "Polynucleotide constructs" section, and the "Oligonucleotide Probes And Primers" section.

A further recombinant cell host according to the invention comprises a polynucleotide containing a biallelic marker selected from the group consisting of A1 to A80, and the complements thereof.

An additional recombinant cell host according to the invention comprises any of the vectors
5 described herein, more particularly any of the vectors described in the "Recombinant Vectors" section.

Preferred host cells used as recipients for the expression vectors of the invention are the following:

a) Prokaryotic host cells: *Escherichia coli* strains (*I.E.*DH5- α strain), *Bacillus*
10 *subtilis*, *Salmonella typhimurium*, and strains from species like *Pseudomonas*, *Streptomyces* and *Staphylococcus*.

b) Eukaryotic host cells: HeLa cells (ATCC N°CCL2; N°CCL2.1; N°CCL2.2), Cv
1 cells (ATCC N°CCL70), COS cells (ATCC N°CRL1650; N°CRL1651), Sf-9 cells
(ATCC N°CRL1711), C127 cells (ATCC N° CRL-1804), 3T3 (ATCC N° CRL-6361),
15 CHO (ATCC N° CCL-61), human kidney 293. (ATCC N° 45504; N° CRL-1573) and
BHK (ECACC N° 84100501; N° 84111301).

c) Other mammalian host cells.

The PG-3 gene expression in mammalian, and typically human, cells may be rendered defective, or alternatively expression may be provided by the insertion of a PG-3 genomic or cDNA
20 sequence with the replacement of the PG-3 gene counterpart in the genome of an animal cell by a PG-3 polynucleotide according to the invention. These genetic alterations may be generated by homologous recombination events using specific DNA constructs that have been previously described.

One kind of cell hosts that may be used are mammalian zygotes, such as murine zygotes.
25 For example, murine zygotes may undergo microinjection with a purified DNA molecule of interest, for example a purified DNA molecule that has previously been adjusted to a concentration range from 1 ng/ml –for BAC inserts- 3 ng/ μ l –for P1 bacteriophage inserts- in 10 mM Tris-HCl, pH 7.4, 250 μ M EDTA containing 100 mM NaCl, 30 μ M spermine, and 70 μ M spermidine. When the DNA to be microinjected has a large size, polyamines and high salt concentrations can be used
30 in order to avoid mechanical breakage of this DNA, as described by Schedl et al (1993b).

Anyone of the polynucleotides of the invention, including the DNA constructs described herein, may be introduced in an embryonic stem (ES) cell line, preferably a mouse ES cell line. ES cell lines are derived from pluripotent, uncommitted cells of the inner cell mass of pre-implantation blastocysts. Preferred ES cell lines are the following: ES-E14TG2a (ATCC n° CRL-1821), ES-D3
35 (ATCC n° CRL1934 and n° CRL-11632), YS001 (ATCC n° CRL-11776), 36.5 (ATCC n° CRL-11116). To maintain ES cells in an uncommitted state, they are cultured in the presence of growth inhibited feeder cells which provide the appropriate signals to preserve this embryonic phenotype

and serve as a matrix for ES cell adherence. Preferred feeder cells consist of primary embryonic fibroblasts that are established from tissue of day 13- day 14 embryos of virtually any mouse strain, that are maintained in culture, such as described by Abbondanzo *et al.*(1993) and are inhibited in growth by irradiation, such as described by Robertson (1987), or by the presence of an inhibitory
5 concentration of LIF, such as described by Pease and Williams (1990).

The constructs in the host cells can be used in a conventional manner to produce the gene product encoded by the recombinant sequence.

Following transformation of a suitable host and growth of the host to an appropriate cell density, the selected promoter is induced by appropriate means, such as temperature shift or
10 chemical induction, and cells are cultivated for an additional period.

Cells are typically harvested by centrifugation, disrupted by physical or chemical means, and the resulting crude extract retained for further purification.

Microbial cells employed in the expression of proteins can be disrupted by any convenient method, including freeze-thaw cycling, sonication, mechanical disruption, or use of cell lysing
15 agents. Such methods are well known by the skill artisan.

TRANSGENIC ANIMALS

The terms "transgenic animals" or "host animals" are used herein designate animals that have their genome genetically and artificially manipulated so as to include one of the nucleic acids according to the invention. Preferred animals are non-human mammals and include those
20 belonging to a genus selected from *Mus* (e.g. mice), *Rattus* (e.g. rats) and *Oryctogalus* (e.g. rabbits) which have their genome artificially and genetically altered by the insertion of a nucleic acid according to the invention. In one embodiment, the invention encompasses non-human host mammals and animals comprising a recombinant vector of the invention or a PG-3 gene disrupted by homologous recombination with a knock out vector.

25 The transgenic animals of the invention all include within a plurality of their cells a cloned recombinant or synthetic DNA sequence, more specifically one of the purified or isolated nucleic acids comprising a PG-3 coding sequence, a PG-3 regulatory polynucleotide, a polynucleotide construct, or a DNA sequence encoding an antisense polynucleotide such as described in the present specification.

30 Generally, a transgenic animal according the present invention comprises any one of the polynucleotides, the recombinant vectors and the cell hosts described in the present invention. More particularly, the transgenic animals of the present invention can comprise any of the polynucleotides described in the "Genomic Sequences Of The PG3 Gene" section, the "PG-3 cDNA Sequences" section, the "Coding Regions" section, the "Polynucleotide constructs" section, the
35 "Oligonucleotide Probes And Primers" section, the "Recombinant Vectors" section and the "Cell Hosts" section.

A further transgenic animals according to the invention contains in their somatic cells and/or in their germ line cells a polynucleotide comprising a biallelic marker selected from the group consisting of A1 to A80, and the complements thereof.

In a first preferred embodiment, these transgenic animals may be good experimental models
5 in order to study the diverse pathologies related to cell differentiation, in particular concerning the transgenic animals within the genome of which has been inserted one or several copies of a polynucleotide encoding a native PG-3 protein, or alternatively a mutant PG-3 protein.

In a second preferred embodiment, these transgenic animals may express a desired polypeptide of interest under the control of the regulatory polynucleotides of the PG-3 gene, leading
10 to good yields in the synthesis of this protein of interest, and eventually a tissue specific expression of this protein of interest.

The design of the transgenic animals of the invention may be made according to the conventional techniques well known from the one skilled in the art. For more details regarding the production of transgenic animals, and specifically transgenic mice, it may be referred to US Patents
15 Nos 4,873,191, issued Oct. 10, 1989; 5,464,764 issued Nov 7, 1995; and 5,789,215, issued Aug 4, 1998; these documents disclosing methods producing transgenic mice.

Transgenic animals of the present invention are produced by the application of procedures which result in an animal with a genome that has incorporated exogenous genetic material. The procedure involves obtaining the genetic material, or a portion thereof, which encodes either a PG-3
20 coding sequence, a PG-3 regulatory polynucleotide or a DNA sequence encoding a PG-3 antisense polynucleotide such as described in the present specification.

A recombinant polynucleotide of the invention is inserted into an embryonic or ES stem cell line. The insertion is preferably made using electroporation, such as described by Thomas *et al.*(1987). The cells subjected to electroporation are screened (e.g. by selection via selectable
25 markers, by PCR or by Southern blot analysis) to find positive cells which have integrated the exogenous recombinant polynucleotide into their genome, preferably via an homologous recombination event. An illustrative positive-negative selection procedure that may be used according to the invention is described by Mansour *et al.*(1988).

Then, the positive cells are isolated, cloned and injected into 3.5 days old blastocysts from
30 mice, such as described by Bradley (1987). The blastocysts are then inserted into a female host animal and allowed to grow to term.

Alternatively, the positive ES cells are brought into contact with embryos at the 2.5 days old 8-16 cell stage (morulae) such as described by Wood *et al.*(1993) or by Nagy *et al.*(1993), the ES cells being internalized to colonize extensively the blastocyst including the cells which will give
35 rise to the germ line.

The offspring of the female host are tested to determine which animals are transgenic e.g. include the inserted exogenous DNA sequence and which are wild-type.

Thus, the present invention also concerns a transgenic animal containing a nucleic acid, a recombinant expression vector or a recombinant host cell according to the invention.

Recombinant Cell Lines Derived From The Transgenic Animals Of The Invention.

A further object of the invention consists of recombinant host cells obtained from a
5 transgenic animal described herein. In one embodiment the invention encompasses cells derived from non-human host mammals and animals comprising a recombinant vector of the invention or a PG-3 gene disrupted by homologous recombination with a knock out vector.

Recombinant cell lines may be established *in vitro* from cells obtained from any tissue of a transgenic animal according to the invention, for example by transfection of primary cell cultures
10 with vectors expressing *onc*-genes such as SV40 large T antigen, as described by Chou (1989) and Shay *et al.* (1991).

**METHODS FOR SCREENING SUBSTANCES INTERACTING WITH A PG-3
POLYPEPTIDE**

For the purpose of the present invention, a ligand means a molecule, such as a protein, a
15 peptide, an antibody or any synthetic chemical compound capable of binding to the PG-3 protein or one of its fragments or variants or to modulate the expression of the polynucleotide coding for PG-3 or a fragment or variant thereof. These molecules may be used in therapeutic compositions, preferably therapeutic compositions acting against cancer.

In the ligand screening method according to the present invention, a biological sample or a
20 defined molecule to be tested as a putative ligand of the PG-3 protein is brought into contact with the corresponding purified PG-3 protein, for example the corresponding purified recombinant PG-3 protein produced by a recombinant cell host as described hereinbefore, in order to form a complex between this protein and the putative ligand molecule to be tested.

As an illustrative example, to study the interaction of the PG-3 protein, or a fragment
25 comprising a contiguous span of at least 6 amino acids, preferably at least 8 to 10 amino acids, more preferably at least 12, 15, 20, 25, 30, 40, 50, or 100 amino acids of SEQ ID No 3, with drugs or small molecules, such as molecules generated through combinatorial chemistry approaches, the microdialysis coupled to HPLC method described by Wang *et al.* (1997) or the affinity capillary electrophoresis method described by Bush *et al.* (1997).

30 In further methods, peptides, drugs, fatty acids, lipoproteins, or small molecules which interact with the PG-3 protein, or a fragment comprising a contiguous span of at least 6 amino acids, preferably at least 8 to 10 amino acids, more preferably at least 12, 15, 20, 25, 30, 40, 50, or 100 amino acids of SEQ ID No 3 may be identified using assays such as the following. The molecule to be tested for binding is labeled with a detectable label, such as a fluorescent, radioactive, or
35 enzymatic tag and placed in contact with immobilized PG-3 protein, or a fragment thereof under conditions which permit specific binding to occur. After removal of non-specifically bound molecules, bound molecules are detected using appropriate means.

Another object of the present invention consists of methods and kits for the screening of candidate substances that interact with PG-3 polypeptide.

The present invention pertains to methods for screening substances of interest that interact with a PG-3 protein or one fragment or variant thereof. By their capacity to bind covalently or non-
5 covalently to a PG-3 protein or to a fragment or variant thereof, these substances or molecules may be advantageously used both *in vitro* and *in vivo*.

In vitro, said interacting molecules may be used as detection means in order to identify the presence of a PG-3 protein in a sample, preferably a biological sample.

A method for the screening of a candidate substance comprises the following steps :

- 10 a) providing a polypeptide consisting of a PG-3 protein or a fragment comprising a contiguous span of at least 6 amino acids, preferably at least 8 to 10 amino acids, more preferably at least 12, 15, 20, 25, 30, 40, 50, or 100 amino acids of SEQ ID No 3;
- b) obtaining a candidate substance;
- c) bringing into contact said polypeptide with said candidate substance;
- 15 d) detecting the complexes formed between said polypeptide and said candidate substance.

The invention further concerns a kit for the screening of a candidate substance interacting with the PG-3 polypeptide, wherein said kit comprises:

- a) a PG-3 protein having an amino acid sequence selected from the group
20 consisting of the amino acid sequences of SEQ ID No 3 or a peptide fragment comprising a contiguous span of at least 6 amino acids, preferably at least 8 to 10 amino acids, more preferably at least 12, 15, 20, 25, 30, 40, 50, or 100 amino acids of SEQ ID No 3;
- b) optionally means useful to detect the complex formed between the PG-3 protein or a peptide fragment or a variant thereof and the candidate substance.

25 In a preferred embodiment of the kit described above, the detection means consist in monoclonal or polyclonal antibodies directed against the PG-3 protein or a peptide fragment or a variant thereof.

 Various candidate substances or molecules can be assayed for interaction with a PG-3 polypeptide. These substances or molecules include, without being limited to, natural or synthetic
30 organic compounds or molecules of biological origin such as polypeptides. When the candidate substance or molecule consists of a polypeptide, this polypeptide may be the resulting expression product of a phage clone belonging to a phage-based random peptide library, or alternatively the polypeptide may be the resulting expression product of a cDNA library cloned in a vector suitable for performing a two-hybrid screening assay.

35 The invention also pertains to kits useful for performing the hereinbefore described screening method. Preferably, such kits comprise a PG-3 polypeptide or a fragment or a variant thereof, and optionally means useful to detect the complex formed between the PG-3 polypeptide or

its fragment or variant and the candidate substance. In a preferred embodiment the detection means consist in monoclonal or polyclonal antibodies directed against the corresponding PG-3 polypeptide or a fragment or a variant thereof.

A. Candidate ligands obtained from random peptide libraries

5 In a particular embodiment of the screening method, the putative ligand is the expression product of a DNA insert contained in a phage vector (Parmley and Smith, 1988). Specifically, random peptide phages libraries are used. The random DNA inserts encode for peptides of 8 to 20 amino acids in length (Oldenburg K.R. *et al.*, 1992; Valadon P., *et al.*, 1996; Lucas A.H., 1994; Westerink M.A.J., 1995; Felici F. *et al.*, 1991). According to this particular embodiment, the
10 recombinant phages expressing a protein that binds to the immobilized PG-3 protein is retained and the complex formed between the PG-3 protein and the recombinant phage may be subsequently immunoprecipitated by a polyclonal or a monoclonal antibody directed against the PG-3 protein.

Once the ligand library in recombinant phages has been constructed, the phage population is brought into contact with the immobilized PG-3 protein. Then the preparation of complexes is
15 washed in order to remove the non-specifically bound recombinant phages. The phages that bind specifically to the PG-3 protein are then eluted by a buffer (acid pH) or immunoprecipitated by the monoclonal antibody produced by the hybridoma anti-PG-3, and this phage population is subsequently amplified by an over-infection of bacteria (for example *E. coli*). The selection step may be repeated several times, preferably 2-4 times, in order to select the more specific
20 recombinant phage clones. The last step consists in characterizing the peptide produced by the selected recombinant phage clones either by expression in infected bacteria and isolation, expressing the phage insert in another host-vector system, or sequencing the insert contained in the selected recombinant phages.

B. Candidate ligands obtained by competition experiments.

25 Alternatively, peptides, drugs or small molecules which bind to the PG-3 protein, or a fragment comprising a contiguous span of at least 6 amino acids, preferably at least 8 to 10 amino acids, more preferably at least 12, 15, 20, 25, 30, 40, 50, or 100 amino acids of SEQ ID No 3, may be identified in competition experiments. In such assays, the PG-3 protein, or a fragment thereof, is immobilized to a surface, such as a plastic plate. Increasing amounts of the peptides, drugs or small
30 molecules are placed in contact with the immobilized PG-3 protein, or a fragment thereof, in the presence of a detectable labeled known PG-3 protein ligand. For example, the PG-3 ligand may be detectably labeled with a fluorescent, radioactive, or enzymatic tag. The ability of the test molecule to bind the PG-3 protein, or a fragment thereof, is determined by measuring the amount of detectably labeled known ligand bound in the presence of the test molecule. A decrease in the
35 amount of known ligand bound to the PG-3 protein, or a fragment thereof, when the test molecule is present indicated that the test molecule is able to bind to the PG-3 protein, or a fragment thereof.

C. Candidate ligands obtained by affinity chromatography.

Proteins or other molecules interacting with the PG-3 protein, or a fragment comprising a contiguous span of at least 6 amino acids, preferably at least 8 to 10 amino acids, more preferably at least 12, 15, 20, 25, 30, 40, 50, or 100 amino acids of SEQ ID No 3, can also be found using affinity columns which contain the PG-3 protein, or a fragment thereof. The PG-3 protein, or a fragment thereof, may be attached to the column using conventional techniques including chemical coupling to a suitable column matrix such as agarose, Affi Gel®, or other matrices familiar to those of skill in art. In some embodiments of this method, the affinity column contains chimeric proteins in which the PG-3 protein, or a fragment thereof, is fused to glutathion S transferase (GST). A mixture of cellular proteins or pool of expressed proteins as described above is applied to the affinity column. Proteins or other molecules interacting with the PG-3 protein, or a fragment thereof, attached to the column can then be isolated and analyzed on 2-D electrophoresis gel as described in Ramussen *et al.* (1997). Alternatively, the proteins retained on the affinity column can be purified by electrophoresis based methods and sequenced. The same method can be used to isolate antibodies, to screen phage display products, or to screen phage display human antibodies.

D. Candidate ligands obtained by optical biosensor methods

Proteins interacting with the PG-3 protein, or a fragment comprising a contiguous span of at least 6 amino acids, preferably at least 8 to 10 amino acids, more preferably at least 12, 15, 20, 25, 30, 40, 50, or 100 amino acids of SEQ ID No 3, can also be screened by using an Optical Biosensor as described in Edwards and Leatherbarrow (1997) and also in Szabo *et al.* (1995). This technique permits the detection of interactions between molecules in real time, without the need of labeled molecules. This technique is based on the surface plasmon resonance (SPR) phenomenon. Briefly, the candidate ligand molecule to be tested is attached to a surface (such as a carboxymethyl dextran matrix). A light beam is directed towards the side of the surface that does not contain the sample to be tested and is reflected by said surface. The SPR phenomenon causes a decrease in the intensity of the reflected light with a specific association of angle and wavelength. The binding of candidate ligand molecules cause a change in the refraction index on the surface, which change is detected as a change in the SPR signal. For screening of candidate ligand molecules or substances that are able to interact with the PG-3 protein, or a fragment thereof, the PG-3 protein, or a fragment thereof, is immobilized onto a surface. This surface consists of one side of a cell through which flows the candidate molecule to be assayed. The binding of the candidate molecule on the PG-3 protein, or a fragment thereof, is detected as a change of the SPR signal. The candidate molecules tested may be proteins, peptides, carbohydrates, lipids, or small molecules generated by combinatorial chemistry. This technique may also be performed by immobilizing eukaryotic or prokaryotic cells or lipid vesicles exhibiting an endogenous or a recombinantly expressed PG-3 protein at their surface.

The main advantage of the method is that it allows the determination of the association rate between the PG-3 protein and molecules interacting with the PG-3 protein. It is thus possible to

select specifically ligand molecules interacting with the PG-3 protein, or a fragment thereof, through strong or conversely weak association constants.

E. Candidate ligands obtained through a two-hybrid screening assay.

The yeast two-hybrid system is designed to study protein-protein interactions *in vivo* (Fields
5 and Song, 1989), and relies upon the fusion of a bait protein to the DNA binding domain of the yeast Gal4 protein. This technique is also described in the US Patent N° US 5,667,973 and the US Patent N° 5,283,173.

The general procedure of library screening by the two-hybrid assay may be performed as described by Harper *et al.* (1993) or as described by Cho *et al.* (1998) or also Fromont-Racine *et al.*
10 (1997).

The bait protein or polypeptide consists of a PG-3 polypeptide or a fragment comprising a contiguous span of at least 6 amino acids, preferably at least 8 to 10 amino acids, more preferably at least 12, 15, 20, 25, 30, 40, 50, or 100 amino acids of SEQ ID No 3.

More precisely, the nucleotide sequence encoding the PG-3 polypeptide or a fragment or
15 variant thereof is fused to a polynucleotide encoding the DNA binding domain of the GAL4 protein, the fused nucleotide sequence being inserted in a suitable expression vector, for example pAS2 or pM3.

Then, a human cDNA library is constructed in a specially designed vector, such that the human cDNA insert is fused to a nucleotide sequence in the vector that encodes the transcriptional
20 domain of the GAL4 protein. Preferably, the vector used is the pACT vector. The polypeptides encoded by the nucleotide inserts of the human cDNA library are termed "prey" polypeptides.

A third vector contains a detectable marker gene, such as beta galactosidase gene or CAT gene that is placed under the control of a regulation sequence that is responsive to the binding of a complete Gal4 protein containing both the transcriptional activation domain and the DNA binding
25 domain. For example, the vector pG5EC may be used.

Two different yeast strains are also used. As an illustrative but non limiting example the two different yeast strains may be the followings :

- Y190, the phenotype of which is (*MATa, Leu2-3, 112 ura3-12, trp1-901, his3-D200, ade2-101, gal4Dgal180D URA3 GAL-LacZ, LYS GAL-HIS3, cyh*);
- 30 - Y187, the phenotype of which is (*MATa gal4 gal80 his3 trp1-901 ade2-101 ura3-52 leu2-3, -112 URA3 GAL-lacZmet*), which is the opposite mating type of Y190.

Briefly, 20 µg of pAS2/PG-3 and 20 µg of pACT-cDNA library are co-transformed into yeast strain Y190. The transformants are selected for growth on minimal media lacking histidine, leucine and tryptophan, but containing the histidine synthesis inhibitor 3-AT (50 mM). Positive
35 colonies are screened for beta galactosidase by filter lift assay. The double positive colonies (*His⁺, beta-gal⁺*) are then grown on plates lacking histidine, leucine, but containing tryptophan and cycloheximide (10 mg/ml) to select for loss of pAS2/PG-3 plasmids but retention of pACT-cDNA

library plasmids. The resulting Y190 strains are mated with Y187 strains expressing PG-3 or non-related control proteins; such as cyclophilin B, lamin, or SNF1, as *Gal4* fusions as described by Harper *et al.* (1993) and by Bram *et al.* (1993), and screened for beta galactosidase by filter lift assay. Yeast clones that are *beta gal-* after mating with the control *Gal4* fusions are considered
5 false positives.

In another embodiment of the two-hybrid method according to the invention, interaction between the PG-3 or a fragment or variant thereof with cellular proteins may be assessed using the Matchmaker Two Hybrid System 2 (Catalog No. K1604-1, Clontech). As described in the manual accompanying the Matchmaker Two Hybrid System 2 (Catalog No. K1604-1, Clontech), nucleic acids
10 encoding the PG-3 protein or a portion thereof, are inserted into an expression vector such that they are in frame with DNA encoding the DNA binding domain of the yeast transcriptional activator GAL4. A desired cDNA, preferably human cDNA, is inserted into a second expression vector such that they are in frame with DNA encoding the activation domain of GAL4. The two expression plasmids are transformed into yeast and the yeast are plated on selection medium which selects for expression of
15 selectable markers on each of the expression vectors as well as GAL4 dependent expression of the HIS3 gene. Transformants capable of growing on medium lacking histidine are screened for GAL4 dependent lacZ expression. Those cells which are positive in both the histidine selection and the lacZ assay contain interaction between PG-3 and the protein or peptide encoded by the initially selected cDNA insert.

20 **METHOD FOR SCREENING SUBSTANCES INTERACTING WITH THE
REGULATORY SEQUENCES OF THE PG-3 GENE.**

The present invention also concerns a method for screening substances or molecules that are able to interact with the regulatory sequences of the PG-3 gene, such as for example promoter or enhancer sequences.

25 Nucleic acids encoding proteins which are able to interact with the regulatory sequences of the PG-3 gene, more particularly a nucleotide sequence selected from the group consisting of the polynucleotides of the 5' and 3' regulatory region or a fragment or variant thereof, and preferably a variant comprising one of the biallelic markers of the invention, may be identified by using a one-hybrid system, such as that described in the booklet enclosed in the Matchmaker One-Hybrid
30 System kit from Clontech (Catalog Ref. n° K1603-1). Briefly, the target nucleotide sequence is cloned upstream of a selectable reporter sequence and the resulting DNA construct is integrated in the yeast genome (*Saccharomyces cerevisiae*). The yeast cells containing the reporter sequence in their genome are then transformed with a library consisting of fusion molecules between cDNAs encoding candidate proteins for binding onto the regulatory sequences of the PG-3 gene and
35 sequences encoding the activator domain of a yeast transcription factor such as GAL4. The recombinant yeast cells are plated in a culture broth for selecting cells expressing the reporter sequence. The recombinant yeast cells thus selected contain a fusion protein that is able to bind

onto the target regulatory sequence of the PG-3 gene. Then, the cDNAs encoding the fusion proteins are sequenced and may be cloned into expression or transcription vectors *in vitro*. The binding of the encoded polypeptides to the target regulatory sequences of the PG-3 gene may be confirmed by techniques familiar to the one skilled in the art, such as gel retardation assays or

5 DNase protection assays.

Gel retardation assays may also be performed independently in order to screen candidate molecules that are able to interact with the regulatory sequences of the PG-3 gene, such as described by Fried and Crothers (1981), Garner and Revzin (1981) and Dent and Latchman (1993). These techniques are based on the principle according to which a DNA fragment which is bound to a

10 protein migrates slower than the same unbound DNA fragment. Briefly, the target nucleotide sequence is labeled. Then the labeled target nucleotide sequence is brought into contact with either a total nuclear extract from cells containing transcription factors, or with different candidate molecules to be tested. The interaction between the target regulatory sequence of the PG-3 gene and the candidate molecule or the transcription factor is detected after gel or capillary

15 electrophoresis through a retardation in the migration.

METHOD FOR SCREENING LIGANDS THAT MODULATE THE EXPRESSION OF THE PG-3 GENE.

Another subject of the present invention is a method for screening molecules that modulate the expression of the PG-3 protein. Such a screening method comprises the steps of:

- 20 a) cultivating a prokaryotic or an eukaryotic cell that has been transfected with a nucleotide sequence encoding the PG-3 protein or a variant or a fragment thereof, placed under the control of its own promoter;
- b) bringing into contact the cultivated cell with a molecule to be tested;
- c) quantifying the expression of the PG-3 protein or a variant or a fragment thereof.

25 In an embodiment, the nucleotide sequence encoding the PG-3 protein or a variant or a fragment thereof comprises an allele of at least one of the biallelic markers A1 to A80, and the complements thereof.

Using DNA recombination techniques well known by the one skill in the art, the PG-3 protein encoding DNA sequence is inserted into an expression vector, downstream from its

30 promoter sequence. As an illustrative example, the promoter sequence of the PG-3 gene is contained in the nucleic acid of the 5' regulatory region.

The quantification of the expression of the PG-3 protein may be realized either at the mRNA level or at the protein level. In the latter case, polyclonal or monoclonal antibodies may be used to quantify the amounts of the PG-3 protein that have been produced, for example in an ELISA

35 or a RIA assay.

In a preferred embodiment, the quantification of the PG-3 mRNA is realized by a quantitative PCR amplification of the cDNA obtained by a reverse transcription of the total mRNA of the cultivated PG-3 -transfected host cell, using a pair of primers specific for PG-3.

The present invention also concerns a method for screening substances or molecules that
5 are able to increase, or in contrast to decrease, the level of expression of the PG-3 gene. Such a method may allow the one skilled in the art to select substances exerting a regulating effect on the expression level of the PG-3 gene and which may be useful as active ingredients included in pharmaceutical compositions for treating patients suffering from cancer.

Thus, another aspect of the present invention is a method for screening a candidate
10 substance or molecule for the ability to modulate the expression of the PG-3 gene, comprising the following steps:

- a) providing a recombinant cell host containing a nucleic acid, wherein said nucleic acid comprises a nucleotide sequence of the 5' regulatory region or a biologically active fragment or variant thereof located upstream of a polynucleotide encoding a detectable protein;
- 15 b) obtaining a candidate substance; and
- c) determining the ability of the candidate substance to modulate the expression levels of the polynucleotide encoding the detectable protein.

In a further embodiment, the nucleic acid comprising the nucleotide sequence of the 5' regulatory region or a biologically active fragment or variant thereof also includes a 5'UTR region
20 of the PG-3 cDNA of SEQ ID No 2, or one of its biologically active fragments or variants thereof.

Among the preferred polynucleotides encoding a detectable protein, there may be cited polynucleotides encoding beta galactosidase, green fluorescent protein (GFP) and chloramphenicol acetyl transferase (CAT).

The invention also pertains to kits useful for performing the herein described screening
25 method. Preferably, such kits comprise a recombinant vector that allows the expression of a nucleotide sequence of the 5' regulatory region or a biologically active fragment or variant thereof located upstream and operably linked to a polynucleotide encoding a detectable protein or the PG-3 protein or a fragment or a variant thereof.

In another embodiment of a method for the screening of a candidate substance or molecule
30 for the ability to modulate the expression of the PG-3 gene, the method comprises the following steps:

- a) providing a recombinant host cell containing a nucleic acid, wherein said nucleic acid comprises a 5'UTR sequence of the PG-3 cDNA of SEQ ID No 2, or one of its biologically active fragments or variants, the 5'UTR sequence or its biologically active fragment or variant being
35 operably linked to a polynucleotide encoding a detectable protein;
- b) obtaining a candidate substance; and

c) determining the ability of the candidate substance to modulate the expression levels of the polynucleotide encoding the detectable protein.

In a specific embodiment of the above screening method, the nucleic acid that comprises a nucleotide sequence selected from the group consisting of the 5'UTR sequence of the PG-3 cDNA of SEQ ID No 2 or one of its biologically active fragments or variants, includes a promoter sequence which is endogenous with respect to the PG-3 5'UTR sequence.

In another specific embodiment of the above screening method, the nucleic acid that comprises a nucleotide sequence selected from the group consisting of the 5'UTR sequence of the PG-3 cDNA of SEQ ID No 2 or one of its biologically active fragments or variants, includes a promoter sequence which is exogenous with respect to the PG-3 5'UTR sequence defined therein.

In a further preferred embodiment, the nucleic acid comprising the 5'-UTR sequence of the PG-3 cDNA or SEQ ID No 2 or the biologically active fragments thereof includes a biallelic marker selected from the group consisting of A1 to A80 or the complements thereof.

The invention further encompasses a kit for the screening of a candidate substance for the ability to modulate the expression of the PG-3 gene, wherein said kit comprises a recombinant vector that comprises a nucleic acid including a 5'UTR sequence of the PG-3 cDNA of SEQ ID No 2, or one of their biologically active fragments or variants, the 5'UTR sequence or its biologically active fragment or variant being operably linked to a polynucleotide encoding a detectable protein.

For the design of suitable recombinant vectors useful for performing the screening methods described above, the section of the present specification wherein the preferred recombinant vectors of the invention are detailed is pertinent.

Expression levels and patterns of PG-3 may be analyzed by solution hybridization with long probes as described in International Patent Application No. WO 97/05277. Briefly, the PG-3 cDNA or the PG-3 genomic DNA described above, or fragments thereof, is inserted at a cloning site immediately downstream of a bacteriophage (T3, T7 or SP6) RNA polymerase promoter to produce antisense RNA. Preferably, the PG-3 insert comprises at least 100 or more consecutive nucleotides of the genomic DNA sequence or the cDNA sequences. The plasmid is linearized and transcribed in the presence of ribonucleotides comprising modified ribonucleotides (*i.e.* biotin-UTP and DIG-UTP). An excess of this doubly labeled RNA is hybridized in solution with mRNA isolated from cells or tissues of interest. The hybridization is performed under standard stringent conditions (40-50°C for 16 hours in an 80% formamide, 0.4 M NaCl buffer, pH 7-8). The unhybridized probe is removed by digestion with ribonucleases specific for single-stranded RNA (*i.e.* RNases CL3, T1, Phy M, U2 or A). The presence of the biotin-UTP modification enables capture of the hybrid on a microtitration plate coated with streptavidin. The presence of the DIG modification enables the hybrid to be detected and quantified by ELISA using an anti-DIG antibody coupled to alkaline phosphatase.

Quantitative analysis of PG-3 gene expression may also be performed using arrays. As used herein, the term array means a one dimensional, two dimensional, or multidimensional arrangement of a plurality of nucleic acids of sufficient length to permit specific detection of expression of mRNAs capable of hybridizing thereto. For example, the arrays may contain a
5 plurality of nucleic acids derived from genes whose expression levels are to be assessed. The arrays may include the PG-3 genomic DNA, the PG-3 cDNA sequences or the sequences complementary thereto or fragments thereof, particularly those comprising at least one of the biallelic markers according the present invention, preferably at least one of the biallelic markers A1 to A80. Preferably, the fragments are at least 15 nucleotides in length. In other embodiments, the fragments
10 are at least 25 nucleotides in length. In some embodiments, the fragments are at least 50 nucleotides in length. More preferably, the fragments are at least 100 nucleotides in length. In another preferred embodiment, the fragments are more than 100 nucleotides in length. In some embodiments the fragments may be more than 500 nucleotides in length.

For example, quantitative analysis of PG-3 gene expression may be performed with a
15 complementary DNA microarray as described by Schena *et al.* (1995 and 1996). Full length PG-3 cDNAs or fragments thereof are amplified by PCR and arrayed from a 96-well microtiter plate onto silylated microscope slides using high-speed robotics. Printed arrays are incubated in a humid chamber to allow rehydration of the array elements and rinsed, once in 0. 2% SDS for 1 min, twice in water for 1 min and once for 5 min in sodium borohydride solution. The arrays are submerged in
20 water for 2 min at 95°C, transferred into 0. 2% SDS for 1 min, rinsed twice with water, air dried and stored in the dark at 25°C.

Cell or tissue mRNA is isolated or commercially obtained and probes are prepared by a single round of reverse transcription. Probes are hybridized to 1 cm² microarrays under a 14 x 14 mm glass coverslip for 6-12 hours at 60°C. Arrays are washed for 5 min at 25°C in low stringency
25 wash buffer (1X SSC/0. 2% SDS), then for 10 min at room temperature in high stringency wash buffer (0. 1X SSC/0. 2% SDS). Arrays are scanned in 0. 1X SSC using a fluorescence laser scanning device fitted with a custom filter set. Accurate differential expression measurements are obtained by taking the average of the ratios of two independent hybridizations.

Quantitative analysis of PG-3 gene expression may also be performed with full length PG-3
30 cDNAs or fragments thereof in complementary DNA arrays as described by Pietu *et al.* (1996). The full length PG-3 cDNA or fragments thereof is PCR amplified and spotted on membranes. Then, mRNAs originating from various tissues or cells are labeled with radioactive nucleotides. After hybridization and washing in controlled conditions, the hybridized mRNAs are detected by phospho-imaging or autoradiography. Duplicate experiments are performed and a quantitative
35 analysis of differentially expressed mRNAs is then performed.

Alternatively, expression analysis using the PG-3 genomic DNA, the PG-3 cDNA, or fragments thereof can be done through high density nucleotide arrays as described by Lockhart *et*

al.(1996) and Sosnowski *et al.*(1997). Oligonucleotides of 15-50 nucleotides from the sequences of the PG-3 genomic DNA, the PG-3 cDNA sequences particularly those comprising at least one of biallelic markers according the present invention, preferably at least one biallelic marker selected from the group consisting of A1 to A80, or the sequences complementary thereto, are synthesized
5 directly on the chip (Lockhart *et al.*, supra) or synthesized and then addressed to the chip (Sosnowski *et al.*, supra). Preferably, the oligonucleotides are about 20 nucleotides in length.

PG-3 cDNA probes labeled with an appropriate compound, such as biotin, digoxigenin or fluorescent dye, are synthesized from the appropriate mRNA population and then randomly fragmented to an average size of 50 to 100 nucleotides. The said probes are then hybridized to the
10 chip. After washing as described in Lockhart *et al.*, supra and application of different electric fields (Sosnowski *et al.*, 1997), the dyes or labeling compounds are detected and quantified. Duplicate hybridizations are performed. Comparative analysis of the intensity of the signal originating from cDNA probes on the same target oligonucleotide in different cDNA samples indicates a differential expression of PG-3 mRNA.

15 METHODS FOR INHIBITING THE EXPRESSION OF A PG-3 GENE

Other therapeutic compositions according to the present invention comprise advantageously an oligonucleotide fragment of the nucleic sequence of PG-3 as an antisense tool or a triple helix tool that inhibits the expression of the corresponding PG-3 gene. A preferred fragment of the nucleic sequence of PG-3 comprises an allele of at least one of the biallelic markers A1 to A80.

20 **Antisense Approach**

Preferred methods using antisense polynucleotide according to the present invention are the procedures described by Sczakiel *et al.*(1995).

Preferably, the antisense tools are chosen among the polynucleotides (15-200 bp long) that are complementary to the 5'end of the PG-3 mRNA. In another embodiment, a combination of
25 different antisense polynucleotides complementary to different parts of the desired targeted gene are used.

Preferred antisense polynucleotides according to the present invention are complementary to a sequence of the mRNAs of PG-3 that contains either the translation initiation codon ATG or a splicing donor or acceptor site.

30 The antisense nucleic acids should have a length and melting temperature sufficient to permit formation of an intracellular duplex having sufficient stability to inhibit the expression of the PG-3 mRNA in the duplex. Strategies for designing antisense nucleic acids suitable for use in gene therapy are disclosed in Green *et al.*, (1986) and Izant and Weintraub, (1984).

In some strategies, antisense molecules are obtained by reversing the orientation of the PG-
35 3 coding region with respect to a promoter so as to transcribe the opposite strand from that which is normally transcribed in the cell. The antisense molecules may be transcribed using in vitro transcription systems such as those which employ T7 or SP6 polymerase to generate the transcript.

Another approach involves transcription of PG-3 antisense nucleic acids in vivo by operably linking DNA containing the antisense sequence to a promoter in a suitable expression vector.

Alternatively, suitable antisense strategies are those described by Rossi *et al.* (1991), in the International Applications Nos. WO 94/23026, WO 95/04141, WO 92/18522 and in the European
5 Patent Application No. EP 0 572 287 A2.

An alternative to the antisense technology that is used according to the present invention consists in using ribozymes that will bind to a target sequence via their complementary polynucleotide tail and that will cleave the corresponding RNA by hydrolyzing its target site (namely "hammerhead ribozymes"). Briefly, the simplified cycle of a hammerhead ribozyme
10 consists of (1) sequence specific binding to the target RNA via complementary antisense sequences; (2) site-specific hydrolysis of the cleavable motif of the target strand; and (3) release of cleavage products, which gives rise to another catalytic cycle. Indeed, the use of long-chain antisense polynucleotide (at least 30 bases long) or ribozymes with long antisense arms are advantageous. A preferred delivery system for antisense ribozyme is achieved by covalently linking these antisense
15 ribozymes to lipophilic groups or to use liposomes as a convenient vector. Preferred antisense ribozymes according to the present invention are prepared as described by Sczakiel *et al.* (1995).

Triple Helix Approach

The PG-3 genomic DNA may also be used to inhibit the expression of the PG-3 gene based on intracellular triple helix formation.

20 Triple helix oligonucleotides are used to inhibit transcription from a genome. They are particularly useful for studying alterations in cell activity when it is associated with a particular gene.

Similarly, a portion of the PG-3 genomic DNA can be used to study the effect of inhibiting PG-3 transcription within a cell. Traditionally, homopurine sequences were considered the most
25 useful for triple helix strategies. However, homopyrimidine sequences can also inhibit gene expression. Such homopyrimidine oligonucleotides bind to the major groove at homopurine:homopyrimidine sequences. Thus, both types of sequences from the PG-3 genomic DNA are contemplated within the scope of this invention.

To carry out gene therapy strategies using the triple helix approach, the sequences of the
30 PG-3 genomic DNA are first scanned to identify 10-mer to 20-mer homopyrimidine or homopurine stretches which could be used in triple-helix based strategies for inhibiting PG-3 expression. Following identification of candidate homopyrimidine or homopurine stretches, their efficiency in inhibiting PG-3 expression is assessed by introducing varying amounts of oligonucleotides containing the candidate sequences into tissue culture cells which express the PG-3 gene.

35 The oligonucleotides can be introduced into the cells using a variety of methods known to those skilled in the art, including but not limited to calcium phosphate precipitation, DEAE-Dextran, electroporation, liposome-mediated transfection or native uptake.

Treated cells are monitored for altered cell function or reduced PG-3 expression using techniques such as Northern blotting, RNase protection assays, or PCR based strategies to monitor the transcription levels of the PG-3 gene in cells which have been treated with the oligonucleotide.

The oligonucleotides which are effective in inhibiting gene expression in tissue culture cells may then be introduced in vivo using the techniques described above in the antisense approach at a dosage calculated based on the in vitro results, as described in antisense approach.

In some embodiments, the natural (beta) anomers of the oligonucleotide units can be replaced with alpha anomers to render the oligonucleotide more resistant to nucleases. Further, an intercalating agent such as ethidium bromide, or the like, can be attached to the 3' end of the alpha oligonucleotide to stabilize the triple helix. For information on the generation of oligonucleotides suitable for triple helix formation see Griffin *et al.* (1989), which is hereby incorporated by this reference.

COMPUTER-RELATED EMBODIMENTS

As used herein the term "nucleic acid codes of the invention" encompass the nucleotide sequences comprising, consisting essentially of, or consisting of any one of the following: a) a contiguous span of at least 12, 15, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 500, or 1000 nucleotides of SEQ ID No 1, wherein said contiguous span comprises at least 1, 2, 3, 5, or 10 of the following nucleotide positions of SEQ ID No 1: 1-97921, 98517-103471, 103603-108222, 108390-109221, 109324-114409, 114538-115723, 115957-122102, 122225-126876, 127033-157212, 157808-240825; b) a contiguous span of at least 12, 15, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 500, or 1000 nucleotides of SEQ ID No 2 or the complements thereof; and, c) a nucleotide sequence complementary to any one of the preceding nucleotide sequences.

The "nucleic acid codes of the invention" further encompass nucleotide sequences homologous to: a) a contiguous span of at least 12, 15, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 500, or 1000 nucleotides of SEQ ID No 1, wherein said contiguous span comprises at least 1, 2, 3, 5, or 10 of the following nucleotide positions of SEQ ID No 1: 1-97921, 98517-103471, 103603-108222, 108390-109221, 109324-114409, 114538-115723, 115957-122102, 122225-126876, 127033-157212, 157808-240825; b) a contiguous span of at least 12, 15, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 500, or 1000 nucleotides of SEQ ID No 2 or the complements thereof; and, c) sequences complementary to all of the preceding sequences.

Homologous sequences refer to a sequence having at least 99%, 98%, 97%, 96%, 95%, 90%, 85%, 80%, or 75% homology to these contiguous spans. Homology may be determined using any method described herein, including BLAST2N with the default parameters or with any modified parameters. Homologous sequences also may include RNA sequences in which uridines replace the thymines in the nucleic acid codes of the invention. It will be appreciated that the nucleic acid codes of the invention can be represented in the traditional single character format (See the inside back cover of Stryer,

Lubert. 1995) or in any other format or code which records the identity of the nucleotides in a sequence.

As used herein the term "polypeptide codes of the invention" encompass the polypeptide sequences comprising a contiguous span of at least 6, 8, 10, 12, 15, 20, 25, 30, 40, 50, or 100 amino acids of SEQ ID No 3. It will be appreciated that the polypeptide codes of the invention can be represented in the traditional single character format or three letter format (See the inside back cover of Stryer, Lubert.) or in any other format or code which records the identity of the polypeptides in a sequence.

It will be appreciated by those skilled in the art that the nucleic acid codes of the invention and polypeptide codes of the invention can be stored, recorded, and manipulated on any medium which can be read and accessed by a computer. As used herein, the words "recorded" and "stored" refer to a process for storing information on a computer medium. A skilled artisan can readily adopt any of the presently known methods for recording information on a computer readable medium to generate manufactures comprising one or more of the nucleic acid codes of the invention, or one or more of the polypeptide codes of the invention. Another aspect of the present invention is a computer readable medium having recorded thereon at least 2, 5, 10, 15, 20, 25, 30, or 50 nucleic acid codes of the invention. Another aspect of the present invention is a computer readable medium having recorded thereon at least 2, 5, 10, 15, 20, 25, 30, or 50 polypeptide codes of the invention.

Computer readable media include magnetically readable media, optically readable media, electronically readable media and magnetic/optical media. For example, the computer readable media may be a hard disk, a floppy disk, a magnetic tape, CD-ROM, Digital Versatile Disk (DVD), Random Access Memory (RAM), or Read Only Memory (ROM) as well as other types of other media known to those skilled in the art.

Embodiments of the present invention include systems, particularly computer systems which store and manipulate the sequence information described herein. One example of a computer system 100 is illustrated in block diagram form in Figure 1. As used herein, "a computer system" refers to the hardware components, software components, and data storage components used to analyze the nucleotide sequences of the nucleic acid codes of the invention or the amino acid sequences of the polypeptide codes of the invention. In one embodiment, the computer system 100 is a Sun Enterprise 1000 server (Sun Microsystems, Palo Alto, CA). The computer system 100 preferably includes a processor for processing, accessing and manipulating the sequence data. The processor 105 can be any well-known type of central processing unit, such as the Pentium III from Intel Corporation, or similar processor from Sun, Motorola, Compaq or International Business Machines.

Preferably, the computer system 100 is a general purpose system that comprises the processor 105 and one or more internal data storage components 110 for storing data, and one or more data retrieving devices for retrieving the data stored on the data storage components. A skilled artisan can readily appreciate that any one of the currently available computer systems are suitable.

In one particular embodiment, the computer system 100 includes a processor 105 connected to a bus which is connected to a main memory 115 (preferably implemented as RAM) and one or more internal data storage devices 110, such as a hard drive and/or other computer readable media having data recorded thereon. In some embodiments, the computer system 100 further includes one or more
5 data retrieving device 118 for reading the data stored on the internal data storage devices 110.

The data retrieving device 118 may represent, for example, a floppy disk drive, a compact disk drive, a magnetic tape drive, etc. In some embodiments, the internal data storage device 110 is a removable computer readable medium such as a floppy disk, a compact disk, a magnetic tape, etc. containing control logic and/or data recorded thereon. The computer system 100 may advantageously
10 include or be programmed by appropriate software for reading the control logic and/or the data from the data storage component once inserted in the data retrieving device.

The computer system 100 includes a display 120 which is used to display output to a computer user. It should also be noted that the computer system 100 can be linked to other computer systems 125a-c in a network or wide area network to provide centralized access to the computer system 100.

15 Software for accessing and processing the nucleotide sequences of the nucleic acid codes of the invention or the amino acid sequences of the polypeptide codes of the invention (such as search tools, compare tools, and modeling tools etc.) may reside in main memory 115 during execution.

In some embodiments, the computer system 100 may further comprise a sequence comparer for comparing the above-described nucleic acid codes of the invention or the polypeptide codes of the
20 invention stored on a computer readable medium to reference nucleotide or polypeptide sequences stored on a computer readable medium. A "sequence comparer" refers to one or more programs which are implemented on the computer system 100 to compare a nucleotide or polypeptide sequence with other nucleotide or polypeptide sequences and/or compounds including but not limited to peptides, peptidomimetics, and chemicals stored within the data storage means. For example, the sequence
25 comparer may compare the nucleotide sequences of nucleic acid codes of the invention or the amino acid sequences of the polypeptide codes of the invention stored on a computer readable medium to reference sequences stored on a computer readable medium to identify homologies, motifs implicated in biological function, or structural motifs. The various sequence comparer programs identified elsewhere in this patent specification are particularly contemplated for use in this aspect of the
30 invention.

Figure 2 is a flow diagram illustrating one embodiment of a process 200 for comparing a new nucleotide or protein sequence with a database of sequences in order to determine the homology levels between the new sequence and the sequences in the database. The database of sequences can be a private database stored within the computer system 100, or a public database such as GENBANK, PIR
35 OR SWISSPROT that is available through the Internet.

99-12767	A38	99-12767-189	Intron J-K	C	T	123277		
99-12767	A39	99-12767-380	Intron J-K	A	G	123468		
4-80	A40	4-80-328	Intron J-K	C	T	126738		
4-36	A41	4-36-384	Intron J-K	G	C	128210		
4-36	A42	4-36-264	Intron J-K	A	G	128330		
4-36	A43	4-36-261	Intron J-K	A	C	128333		
4-35	A44	4-35-333	Intron J-K	A	C	128594		
4-35	A45	4-35-240	Intron J-K	G	C	128687		
4-35	A46	4-35-173	Intron J-K	A	T	128754		
4-35	A47	4-35-133	Intron J-K	C	T	128794		
99-12771	A48	99-12771-59	Intron J-K	G	T	130805		
99-12774	A49	99-12774-334	Intron J-K	A	C	133206		
99-12776	A50	99-12776-358	Intron J-K	A	G	135386		
99-12781	A51	99-12781-113	Intron J-K	A	G	139389		
4-104	A52	4-104-298	Intron J-K	G	C	157535		
4-104	A53	4-104-254	Intron J-K	A	G	157579		
4-104	A54	4-104-250	Intron J-K	C	T	157583		
4-104	A55	4-104-214	Intron J-K	A	G	157619		
99-12818	A56	99-12818-289	Intron J-K	C	T	172980		
99-24807	A57	99-24807-271	Intron J-K	C	T	180622		
99-24807	A58	99-24807-84	Intron J-K	A	G	180809		
99-12831	A59	99-12831-157	Intron J-K	A	G	190334		
99-12831	A60	99-12831-241	Intron J-K	C	T	190418		
99-12832	A61	99-12832-387	Intron J-K	C	T	191397		
99-12836	A62	99-12836-30	Intron J-K	G	C	195128		
99-12844	A63	99-12844-262	Intron J-K	G	C	203846		
4-24	A64	4-24-74	Intron J-K	C	T	210151		
4-24	A65	4-24-246	Intron J-K	C	T	210321		
4-24	A66	4-24-314	Intron J-K	G	C	210389		
4-27	A67	4-27-190	Intron J-K	A	G	211168		
5-400	A68	5-400-145	Intron J-K	A	G	215996		
5-400	A69	5-400-149	Intron J-K	G	C	216000		
5-400	A70	5-400-175	Exon K	C	T	216026	2283	742 = S
5-400	A71	5-400-231	Exon K	C	T	216082	2339	761 = A or V
5-400	A72	5-400-367	Exon K	A	C	216218	2475	806 = A
99-12852	A73	99-12852-110	Intron K-L	G	T	216322		
99-12852	A74	99-12852-325	Intron K-L	A	G	216537		
4-37	A75	4-37-326	Intron K-L	A	C	221649		
4-37	A76	4-37-107	Intron K-L	A	G	221867		
5-270	A77	5-270-92	Intron K-L	G	C	225645		
99-12860	A78	99-12860-47	Intron K-L	A	G	229387		
99-12860	A79	99-12860-57	Intron K-L	A	T	229397		
5-402	A80	5-402-144	Exon L	C	T	237555	2539	828 = P or S

BM refers to "biallelic marker". All1 and all2 refer respectively to allele 1 and allele 2 of the biallelic marker.

The process 200 begins at a start state 201 and then moves to a state 202 wherein the new sequence to be compared is stored to a memory in a computer system 100. As discussed above, the memory could be any type of memory, including RAM or an internal storage device.

The process 200 then moves to a state 204 wherein a database of sequences is opened for
5 analysis and comparison. The process 200 then moves to a state 206 wherein the first sequence stored in the database is read into a memory on the computer. A comparison is then performed at a state 210 to determine if the first sequence is the same as the second sequence. It is important to note that this step is not limited to performing an exact comparison between the new sequence and the first sequence in the database. Well-known methods are known to those of skill in the art for comparing two
10 nucleotide or protein sequences, even if they are not identical. For example, gaps can be introduced into one sequence in order to raise the homology level between the two tested sequences. The parameters that control whether gaps or other features are introduced into a sequence during comparison are normally entered by the user of the computer system.

Once a comparison of the two sequences has been performed at the state 210, a determination
15 is made at a decision state 210 whether the two sequences are the same. Of course, the term "same" is not limited to sequences that are absolutely identical. Sequences that are within the homology parameters entered by the user will be marked as "same" in the process 200.

If a determination is made that the two sequences are the same, the process 200 moves to a state 214 wherein the name of the sequence from the database is displayed to the user. This state
20 notifies the user that the sequence with the displayed name fulfills the homology constraints that were entered. Once the name of the stored sequence is displayed to the user, the process 200 moves to a decision state 218 wherein a determination is made whether more sequences exist in the database. If no more sequences exist in the database, then the process 200 terminates at an end state 220. However, if more sequences do exist in the database, then the process 200 moves to a state 224 wherein a pointer is
25 moved to the next sequence in the database so that it can be compared to the new sequence. In this manner, the new sequence is aligned and compared with every sequence in the database.

It should be noted that if a determination had been made at the decision state 212 that the sequences were not homologous, then the process 200 would move immediately to the decision state 218 in order to determine if any other sequences were available in the database for comparison.

30 Accordingly, one aspect of the present invention is a computer system comprising a processor, a data storage device having stored thereon a nucleic acid code of the invention or a polypeptide code of the invention, a data storage device having retrievably stored thereon reference nucleotide sequences or polypeptide sequences to be compared to the nucleic acid code of the invention or polypeptide code of the invention and a sequence comparer for conducting the
35 comparison. The sequence comparer may indicate a homology level between the sequences compared or identify structural motifs in the nucleic acid code of the invention and polypeptide codes of the invention or it may identify structural motifs in sequences which are compared to these

nucleic acid codes and polypeptide codes. In some embodiments, the data storage device may have stored thereon the sequences of at least 2, 5, 10, 15, 20, 25, 30, or 50 of the nucleic acid codes of the invention or polypeptide codes of the invention.

Another aspect of the present invention is a method for determining the level of homology
5 between a nucleic acid code of the invention and a reference nucleotide sequence, comprising the steps of reading the nucleic acid code and the reference nucleotide sequence through the use of a computer program which determines homology levels and determining homology between the nucleic acid code and the reference nucleotide sequence with the computer program. The computer program may be any of a number of computer programs for determining homology levels, including those
10 specifically enumerated herein, including BLAST2N with the default parameters or with any modified parameters. The method may be implemented using the computer systems described above. The method may also be performed by reading 2, 5, 10, 15, 20, 25, 30, or 50 of the above described nucleic acid codes of the invention through the use of the computer program and determining homology between the nucleic acid codes and reference nucleotide sequences.

15 Figure 3 is a flow diagram illustrating one embodiment of a process 250 in a computer for determining whether two sequences are homologous. The process 250 begins at a start state 252 and then moves to a state 254 wherein a first sequence to be compared is stored to a memory. The second sequence to be compared is then stored to a memory at a state 256. The process 250 then moves to a state 260 wherein the first character in the first sequence is read and then to a state 262
20 wherein the first character of the second sequence is read. It should be understood that if the sequence is a nucleotide sequence, then the character would normally be either A, T, C, G or U. If the sequence is a protein sequence, then it should be in the single letter amino acid code so that the first and sequence sequences can be easily compared.

A determination is then made at a decision state 264 whether the two characters are the
25 same. If they are the same, then the process 250 moves to a state 268 wherein the next characters in the first and second sequences are read. A determination is then made whether the next characters are the same. If they are, then the process 250 continues this loop until two characters are not the same. If a determination is made that the next two characters are not the same, the process 250 moves to a decision state 274 to determine whether there are any more characters either sequence to
30 read.

If there aren't any more characters to read, then the process 250 moves to a state 276 wherein the level of homology between the first and second sequences is displayed to the user. The level of homology is determined by calculating the proportion of characters between the sequences that were the same out of the total number of sequences in the first sequence. Thus, if every
35 character in a first 100 nucleotide sequence aligned with a every character in a second sequence, the homology level would be 100%.

Alternatively, the computer program may be a computer program which compares the nucleotide sequences of the nucleic acid codes of the present invention, to reference nucleotide sequences in order to determine whether the nucleic acid code of the invention differs from a reference nucleic acid sequence at one or more positions. Optionally such a program records the length and
5 identity of inserted, deleted or substituted nucleotides with respect to the sequence of either the reference polynucleotide or the nucleic acid code of the invention. In one embodiment, the computer program may be a program which determines whether the nucleotide sequences of the nucleic acid codes of the invention contain one or more single nucleotide polymorphisms (SNP) with respect to a reference nucleotide sequence. These single nucleotide polymorphisms may each comprise a single
10 base substitution, insertion, or deletion.

Another aspect of the present invention is a method for determining the level of homology between a polypeptide code of the invention and a reference polypeptide sequence, comprising the steps of reading the polypeptide code of the invention and the reference polypeptide sequence through use of a computer program which determines homology levels and determining homology between the
15 polypeptide code and the reference polypeptide sequence using the computer program.

Accordingly, another aspect of the present invention is a method for determining whether a nucleic acid code of the invention differs at one or more nucleotides from a reference nucleotide sequence comprising the steps of reading the nucleic acid code and the reference nucleotide sequence through use of a computer program which identifies differences between nucleic acid
20 sequences and identifying differences between the nucleic acid code and the reference nucleotide sequence with the computer program. In some embodiments, the computer program is a program which identifies single nucleotide polymorphisms. The method may be implemented by the computer systems described above and the method illustrated in Figure 3. The method may also be performed by reading at least 2, 5, 10, 15, 20, 25, 30, or 50 of the nucleic acid codes of the
25 invention and the reference nucleotide sequences through the use of the computer program and identifying differences between the nucleic acid codes and the reference nucleotide sequences with the computer program.

In other embodiments the computer based system may further comprise an identifier for identifying features within the nucleotide sequences of the nucleic acid codes of the invention or the
30 amino acid sequences of the polypeptide codes of the invention.

An "identifier" refers to one or more programs which identifies certain features within the above-described nucleotide sequences of the nucleic acid codes of the invention or the amino acid sequences of the polypeptide codes of the invention. In one embodiment, the identifier may comprise a program which identifies an open reading frame in the cDNAs codes of the invention.

35 Figure 4 is a flow diagram illustrating one embodiment of an identifier process 300 for detecting the presence of a feature in a sequence. The process 300 begins at a start state 302 and then moves to a state 304 wherein a first sequence that is to be checked for features is stored to a

memory 115 in the computer system 100. The process 300 then moves to a state 306 wherein a database of sequence features is opened. Such a database would include a list of each feature's attributes along with the name of the feature. For example, a feature name could be "Initiation Codon" and the attribute would be "ATG". Another example would be the feature name

5 "TAATAA Box" and the feature attribute would be "TAATAA". An example of such a database is produced by the University of Wisconsin Genetics Computer Group (www.gcg.com).

Once the database of features is opened at the state 306, the process 300 moves to a state 308 wherein the first feature is read from the database. A comparison of the attribute of the first feature with the first sequence is then made at a state 310. A determination is then made at a
10 decision state 316 whether the attribute of the feature was found in the first sequence. If the attribute was found, then the process 300 moves to a state 318 wherein the name of the found feature is displayed to the user.

The process 300 then moves to a decision state 320 wherein a determination is made whether more features exist in the database. If no more features do exist, then the process 300
15 terminates at an end state 324. However, if more features do exist in the database, then the process 300 reads the next sequence feature at a state 326 and loops back to the state 310 wherein the attribute of the next feature is compared against the first sequence.

It should be noted, that if the feature attribute is not found in the first sequence at the decision state 316, the process 300 moves directly to the decision state 320 in order to determine if
20 any more features exist in the database.

In another embodiment, the identifier may comprise a molecular modeling program which determines the 3-dimensional structure of the polypeptides codes of the invention. In some embodiments, the molecular modeling program identifies target sequences that are most compatible with profiles representing the structural environments of the residues in known three-dimensional
25 protein structures. (See, e.g., Eisenberg *et al.*, U.S. Patent No. 5,436,850 issued July 25, 1995). In another technique, the known three-dimensional structures of proteins in a given family are superimposed to define the structurally conserved regions in that family. This protein modeling technique also uses the known three-dimensional structure of a homologous protein to approximate the structure of the polypeptide codes of the invention. (See e.g., Srinivasan, *et al.*, U.S. Patent
30 No. 5,557,535 issued September 17, 1996). Conventional homology modeling techniques have been used routinely to build models of proteases and antibodies. (Sowdhamini *et al.*, (1997)). Comparative approaches can also be used to develop three-dimensional protein models when the protein of interest has poor sequence identity to template proteins. In some cases, proteins fold into similar three-dimensional structures despite having very weak sequence identities. For example, the
35 three-dimensional structures of a number of helical cytokines fold in similar three-dimensional topology in spite of weak sequence homology.

The recent development of threading methods now enables the identification of likely folding patterns in a number of situations where the structural relatedness between target and template(s) is not detectable at the sequence level. Hybrid methods, in which fold recognition is performed using Multiple Sequence Threading (MST), structural equivalencies are deduced from the threading output using a distance geometry program DRAGON to construct a low resolution model, and a full-atom representation is constructed using a molecular modeling package such as QUANTA.

According to this 3-step approach, candidate templates are first identified by using the novel fold recognition algorithm MST, which is capable of performing simultaneous threading of multiple aligned sequences onto one or more 3-D structures. In a second step, the structural equivalencies obtained from the MST output are converted into interresidue distance restraints and fed into the distance geometry program DRAGON, together with auxiliary information obtained from secondary structure predictions. The program combines the restraints in an unbiased manner and rapidly generates a large number of low resolution model confirmations. In a third step, these low resolution model confirmations are converted into full-atom models and subjected to energy minimization using the molecular modeling package QUANTA. (See e.g., Aszódi *et al.*, (1997)).

The results of the molecular modeling analysis may then be used in rational drug design techniques to identify agents which modulate the activity of the polypeptide codes of the invention.

Accordingly, another aspect of the present invention is a method of identifying a feature within the nucleic acid codes of the invention or the polypeptide codes of the invention comprising reading the nucleic acid code(s) or the polypeptide code(s) through the use of a computer program which identifies features therein and identifying features within the nucleic acid code(s) or polypeptide code(s) with the computer program. In one embodiment, computer program comprises a computer program which identifies open reading frames. In a further embodiment, the computer program identifies structural motifs in a polypeptide sequence. In another embodiment, the computer program comprises a molecular modeling program. The method may be performed by reading a single sequence or at least 2, 5, 10, 15, 20, 25, 30, or 50 of the nucleic acid codes of the invention or the polypeptide codes of the invention through the use of the computer program and identifying features within the nucleic acid codes or polypeptide codes with the computer program.

The nucleic acid codes of the invention or the polypeptide codes of the invention may be stored and manipulated in a variety of data processor programs in a variety of formats. For example, they may be stored as text in a word processing file, such as MicrosoftWORD or WORDPERFECT or as an ASCII file in a variety of database programs familiar to those of skill in the art, such as DB2, SYBASE, or ORACLE. In addition, many computer programs and databases may be used as sequence comparers, identifiers, or sources of reference nucleotide or polypeptide sequences to be compared to the nucleic acid codes of the invention or the polypeptide codes of the invention. The following list is intended not to limit the invention but to provide guidance to programs and databases which are

useful with the nucleic acid codes of the invention or the polypeptide codes of the invention. The programs and databases which may be used include, but are not limited to: MacPattern (EMBL), DiscoveryBase (Molecular Applications Group), GeneMine (Molecular Applications Group), Look (Molecular Applications Group), MacLook (Molecular Applications Group), BLAST and BLAST2 (NCBI), BLASTN and BLASTX (Altschul et al, 1990), FASTA (Pearson and Lipman, 1988), FASTDB (Brutlag *et al.*, 1990), Catalyst (Molecular Simulations Inc.), Catalyst/SHAPE (Molecular Simulations Inc.), Cerius².DBAccess (Molecular Simulations Inc.), HypoGen (Molecular Simulations Inc.), Insight II, (Molecular Simulations Inc.), Discover (Molecular Simulations Inc.), CHARMM (Molecular Simulations Inc.), Felix (Molecular Simulations Inc.), DelPhi, (Molecular Simulations Inc.), QuanteMM, (Molecular Simulations Inc.), Homology (Molecular Simulations Inc.), Modeler (Molecular Simulations Inc.), ISIS (Molecular Simulations Inc.), Quanta/Protein Design (Molecular Simulations Inc.), WebLab (Molecular Simulations Inc.), WebLab Diversity Explorer (Molecular Simulations Inc.), Gene Explorer (Molecular Simulations Inc.), SeqFold (Molecular Simulations Inc.), the EMBL/Swissprotein database, the MDL Available Chemicals Directory database, the MDL Drug Data Report data base, the Comprehensive Medicinal Chemistry database, Derwent's World Drug Index database, the BioByteMasterFile database, the Genbank database, the Genseqn database and the Genseqp databases. Many other programs and data bases would be apparent to one of skill in the art given the present disclosure.

Motifs which may be detected using the above programs include sequences encoding leucine zippers, helix-turn-helix motifs, glycosylation sites, ubiquitination sites, alpha helices, and beta sheets, signal sequences encoding signal peptides which direct the secretion of the encoded proteins, sequences implicated in transcription regulation such as homeoboxes, acidic stretches, enzymatic active sites, substrate binding sites, and enzymatic cleavage sites.

Throughout this application, various publications, patents and published patent applications are cited. The disclosures of these publications, patents and published patent specification referenced in this application are hereby incorporated by reference into the present disclosure to more fully describe the state of the art to which this invention pertains.

EXAMPLES

EXAMPLE 1

IDENTIFICATION OF BIALLELIC MARKERS - DNA EXTRACTION

Donors were unrelated and healthy. They presented a sufficient diversity for being representative of a French heterogeneous population. The DNA from 100 individuals was extracted and tested for the detection of the biallelic markers.

30 ml of peripheral venous blood were taken from each donor in the presence of EDTA. Cells (pellet) were collected after centrifugation for 10 minutes at 2000 rpm. Red cells were lysed by a lysis solution (50 ml final volume: 10 mM Tris pH7.6; 5 mM MgCl₂; 10 mM NaCl). The

solution was centrifuged (10 minutes, 2000 rpm) as many times as necessary to eliminate the residual red cells present in the supernatant, after resuspension of the pellet in the lysis solution.

The pellet of white cells was lysed overnight at 42°C with 3.7 ml of lysis solution composed of:

- 5 - 3 ml TE 10-2 (Tris-HCl 10 mM, EDTA 2 mM) / NaCl 0.4 M
- 200 µl SDS 10%
- 500 µl K-proteinase (2 mg K-proteinase in TE 10-2 / NaCl 0.4 M).

For the extraction of proteins, 1 ml saturated NaCl (6M) (1/3.5 v/v) was added. After
10 vigorous agitation, the solution was centrifuged for 20 minutes at 10000 rpm.

For the precipitation of DNA, 2 to 3 volumes of 100% ethanol were added to the previous supernatant, and the solution was centrifuged for 30 minutes at 2000 rpm. The DNA solution was rinsed three times with 70% ethanol to eliminate salts, and centrifuged for 20 minutes at 2000 rpm. The pellet was dried at 37°C, and resuspended in 1 ml TE 10-1 or 1 ml water. The DNA
15 concentration was evaluated by measuring the OD at 260 nm (1 unit OD = 50 µg/ml DNA).

To determine the presence of proteins in the DNA solution, the OD 260 / OD 280 ratio was determined. Only DNA preparations having a OD 260 / OD 280 ratio between 1.8 and 2 were used in the subsequent examples described below.

The pool was constituted by mixing equivalent quantities of DNA from each individual.
20

EXAMPLE 2

IDENTIFICATION OF BIALLELIC MARKERS: AMPLIFICATION OF GENOMIC DNA BY PCR

The amplification of specific genomic sequences of the DNA samples of example 1 was carried out on the pool of DNA obtained previously. In addition, 50 individual samples were
25 similarly amplified.

PCR assays were performed using the following protocol:

	Final volume	25 µl
	DNA	2 ng/µl
	MgCl ₂	2 mM
30	dNTP (each)	200 µM
	primer (each)	2.9 ng/µl
	Ampli Taq Gold DNA polymerase	0.05 unit/µl
	PCR buffer (10x = 0.1 M TrisHCl pH8.3 0.5M KCl)	1x

35 Each pair of first primers was designed using the sequence information of the PG-3 gene disclosed herein and the OSP software (Hillier & Green, 1991). This first pair of primers was about

20 nucleotides in length and had the sequences disclosed in Table 1 in the columns labeled PU and RP.

Table 1

Amplicon	Position range of the amplicon in SEQ ID No:1		PU primer name	Position range of amplification primer in SEQ ID No:1		RP primer name	Complementary position range of amplification primer in SEQ ID No:1	
5-390	1823	2125	B1	1823	1840	C1	2108	2125
5-391	4559	4908	B2	4559	4577	C2	4891	4908
5-392	10007	10430	B3	10007	10025	C3	10411	10430
4-59	39556	39970	B4	39556	39574	C4	39953	39970
4-58	39877	40259	B5	39877	39896	C5	40242	40259
4-54	41137	41581	B6	41137	41154	C6	41564	41581
4-51	42122	42543	B7	42122	42141	C7	42526	42543
99-86	67289	67741	B8	67289	67309	C8	67724	67741
4-88	69182	69626	B9	69182	69200	C9	69609	69626
5-397	72698	73117	B10	72698	72715	C10	73099	73117
5-398	75858	76306	B11	75858	75877	C11	76289	76306
99-12738	81006	81485	B12	81006	81025	C12	81466	81485
99-109	83564	84007	B13	83564	83582	C13	83990	84007
99-12749	91743	92142	B14	91743	91763	C14	92123	92142
4-21	95196	95619	B15	95196	95214	C15	95600	95619
4-23	95865	96229	B16	95865	95882	C16	96210	96229
99-12753	97261	97747	B17	97261	97278	C17	97728	97747
5-364	97831	98275	B18	97831	97849	C18	98256	98275
99-12755	98638	99131	B19	98638	98656	C19	99111	99131
4-87	103376	103818	B20	103376	103395	C20	103801	103818
99-12757	104081	104636	B21	104081	104100	C21	104619	104636
99-12758	106272	106799	B22	106272	106291	C22	106780	106799
4-105	108200	108412	B23	108200	108218	C23	108390	108412
4-45	108223	108520	B24	108223	108246	C24	108499	108520
4-44	109123	109471	B25	109123	109142	C25	109454	109471
4-86	114217	114663	B26	114217	114234	C26	114646	114663
4-84	115630	116049	B27	115630	115647	C27	116031	116049
99-78	121991	122401	B28	121991	122011	C28	122384	122401
99-12767	123089	123583	B29	123089	123106	C29	123565	123583
4-80	126711	127065	B30	126711	126729	C30	127048	127065
4-36	128162	128590	B31	128162	128179	C31	128573	128590
4-35	128480	128926	B32	128480	128497	C32	128909	128926
99-12771	130747	131273	B33	130747	130764	C33	131254	131273
99-12774	132873	133325	B34	132873	132892	C34	133305	133325
99-12776	135029	135478	B35	135029	135048	C35	135458	135478
99-12781	139277	139742	B36	139277	139296	C36	139724	139742
4-104	157181	157832	B37	157181	157199	C37	157814	157832
99-12818	172692	173091	B38	172692	172709	C38	173072	173091

99-24807	180248	180892	B39	180248	180268	C39	180874	180892
99-12827	184662	185156	B40	184662	184680	C40	185138	185156
99-12831	190178	190663	B41	190178	190196	C41	190643	190663
99-12832	191011	191460	B42	191011	191030	C42	191441	191460
99-12836	195099	195587	B43	195099	195116	C43	195568	195587
99-12844	203585	204115	B44	203585	203602	C44	204095	204115
4-24	210079	210495	B45	210079	210096	C45	210476	210495
4-27	210979	211401	B46	210979	210996	C46	211382	211401
5-400	215852	216271	B47	215852	215870	C47	216253	216271
99-12852	216213	216728	B48	216213	216231	C48	216708	216728
4-37	221530	221973	B49	221530	221549	C49	221956	221973
5-270	225554	225845	B50	225554	225572	C50	225827	225845
99-12860	229341	229790	B51	229341	229359	C51	229770	229790
5-402	237412	237766	B52	237412	237429	C52	237747	237766

Preferably, the primers contained a common oligonucleotide tail upstream of the specific bases targeted for amplification which was useful for sequencing.

Primers PU contain the following additional PU 5' sequence:

- 5 TGTAACGACGGCCAGT; primers RP contain the following RP 5' sequence: CAGGAAACAGCTATGACC. The primer containing the additional PU 5' sequence is listed in SEQ ID No 4. The primer containing the additional RP 5' sequence is listed in SEQ ID No 5.

The synthesis of these primers was performed following the phosphoramidite method, on a GENSET UFPS 24.1 synthesizer.

- 10 DNA amplification was performed on a Genius II thermocycler. After heating at 95°C for 10 min, 40 cycles were performed. Each cycle comprised: 30 sec at 95°C, 54°C for 1 min, and 30 sec at 72°C. For final elongation, 10 min at 72°C ended the amplification. The quantities of the amplification products obtained were determined on 96-well microtiter plates, using a fluorometer and Picogreen as intercalant agent (Molecular Probes).

15 EXAMPLE 3

IDENTIFICATION OF BIALLELIC MARKERS - SEQUENCING OF AMPLIFIED GENOMIC DNA AND IDENTIFICATION OF POLYMORPHISMS

- The sequencing of the amplified DNA obtained in example 2 was carried out on ABI 377 sequencers. The sequences of the amplification products were determined using automated dideoxy terminator sequencing reactions with a dye terminator cycle sequencing protocol. The products of the sequencing reactions were run on sequencing gels and the sequences were determined using gel image analysis (ABI Prism DNA Sequencing Analysis software (2.1.2 version)).

- 25 The sequence data were further evaluated to detect the presence of biallelic markers within the amplified fragments. The polymorphism search was based on the presence of superimposed peaks in the electrophoresis pattern resulting from different bases occurring at the same position as described previously.

In the 52 fragments of amplification, 80 biallelic markers were detected. The localization of these biallelic markers are as shown in Table 2.

Table 2

Amplicon	BM	Marker name	Localization in PG-3 gene	Polymorphism		BM position in SEQ ID		Position of amino acid in SEQ ID No:3
				all1	all2	No:1	No:2	
5-390	A1	5-390-177	5' regulatory	G	C	1999		
5-391	A2	5-391-43	Intron A-B	A	G	4601		
5-392	A3	5-392-222	Exon C	G	T	10228	285	76 = V
5-392	A4	5-392-280	Intron C-D	G	T	10286		
5-392	A5	5-392-364	Intron C-D	G	-	10370		
4-59	A6	4-58-318	Exon T	G	T	39944	968	304 = R or I
4-58	A7	4-58-289	Exon T	G	C	39973	997	314 = H or D
4-54	A8	4-54-199	Intron T-G	A	C	41385		
4-54	A9	4-54-180	Intron T-G	A	C	41404		
4-51	A10	4-51-312	Intron T-G	G	C	42232		
99-86	A11	99-86-266	Intron G-H	A	G	67475		
4-88	A12	4-88-107	Intron G-H	A	G	69521		
5-397	A13	5-397-141	Intron G-H	G	T	72838		
5-398	A14	5-398-203	Exon I	A	C	76060	2102	682 = T or N
99-12738	A15	99-12738-248	Intron I-J	A	C	81253		
99-109	A16	99-109-358	Intron I-J	A	C	83921		
99-12749	A17	99-12749-175	Intron I-J	C	T	91917		
4-21	A18	4-21-154	Intron J-K	C	T	95349		
4-21	A19	4-21-317	Intron J-K	G	T	95511		
4-23	A20	4-23-326	Intron J-K	A	G	96190		
99-12753	A21	99-12753-34	Intron J-K	A	T	97294		
5-364	A22	5-364-252	Intron J-K	G	T	98024		
99-12755	A23	99-12755-280	Intron J-K	A	G	98914		
99-12755	A24	99-12755-329	Intron J-K	A	C	98963		
4-87	A25	4-87-212	Intron J-K	A	G	103593		
99-12757	A26	99-12757-318	Intron J-K	C	T	104398		
99-12758	A27	99-12758-102	Intron J-K	A	G	106373		
99-12758	A28	99-12758-136	Intron J-K	C	T	106407		
4-105	A29	4-105-98	Intron J-K	A	G	108315		
4-105	A30	4-105-86	Intron J-K	A	G	108327		
4-45	A31	4-45-49	Intron J-K	C	T	108472		
4-44	A32	4-44-277	Intron J-K	C	T	109196		
4-86	A33	4-86-60	Intron J-K	G	C	114604		
4-84	A34	4-84-334	Intron J-K	A	G	115716		
99-78	A35	99-78-321	Intron J-K	A	T	122083		
99-12767	A36	99-12767-36	Intron J-K	G	C	123124		
99-12767	A37	99-12767-143	Intron J-K	C	T	123231		

Table 3

BM	Marker name	Position range of probes in SEQ ID No 1		Probes
A1	5-390-177	1987	2011	P1
A2	5-391-43	4589	4613	P2
A3	5-392-222	10216	10240	P3
A4	5-392-280	10274	10298	P4
A6	4-58-318	39932	39956	P6
A7	4-58-289	39961	39985	P7
A8	4-54-199	41373	41397	P8
A9	4-54-180	41392	41416	P9
A10	4-51-312	42220	42244	P10
A11	99-86-266	67463	67487	P11
A12	4-88-107	69509	69533	P12
A13	5-397-141	72826	72850	P13
A14	5-398-203	76048	76072	P14
A15	99-12738-248	81241	81265	P15
A16	99-109-358	83909	83933	P16
A17	99-12749-175	91905	91929	P17
A18	4-21-154	95337	95361	P18
A19	4-21-317	95499	95523	P19
A20	4-23-326	96178	96202	P20
A21	99-12753-34	97282	97306	P21
A22	5-364-252	98012	98036	P22
A23	99-12755-280	98902	98926	P23
A24	99-12755-329	98951	98975	P24
A25	4-87-212	103581	103605	P25
A26	99-12757-318	104386	104410	P26
A27	99-12758-102	106361	106385	P27
A28	99-12758-136	106395	106419	P28
A29	4-105-98	108303	108327	P29
A30	4-105-86	108315	108339	P30
A31	4-45-49	108460	108484	P31
A32	4-44-277	109184	109208	P32
A33	4-86-60	114592	114616	P33
A34	4-84-334	115704	115728	P34
A35	99-78-321	122071	122095	P35
A36	99-12767-36	123112	123136	P36
A37	99-12767-143	123219	123243	P37
A38	99-12767-189	123265	123289	P38
A39	99-12767-380	123456	123480	P39
A40	4-80-328	126726	126750	P40
A41	4-36-384	128198	128222	P41
A42	4-36-264	128318	128342	P42
A43	4-36-261	128321	128345	P43
A44	4-35-333	128582	128606	P44

A45	4-35-240	128675	128699	P45
A46	4-35-173	128742	128766	P46
A47	4-35-133	128782	128806	P47
A48	99-12771-59	130793	130817	P48
A49	99-12774-334	133194	133218	P49
A50	99-12776-358	135374	135398	P50
A51	99-12781-113	139377	139401	P51
A52	4-104-298	157523	157547	P52
A53	4-104-254	157567	157591	P53
A54	4-104-250	157571	157595	P54
A55	4-104-214	157607	157631	P55
A56	99-12818-289	172968	172992	P56
A57	99-24807-271	180610	180634	P57
A58	99-24807-84	180797	180821	P58
A59	99-12831-157	190322	190346	P59
A60	99-12831-241	190406	190430	P60
A61	99-12832-387	191385	191409	P61
A62	99-12836-30	195116	195140	P62
A63	99-12844-262	203834	203858	P63
A64	4-24-74	210139	210163	P64
A65	4-24-246	210309	210333	P65
A66	4-24-314	210377	210401	P66
A67	4-27-190	211156	211180	P67
A68	5-400-145	215984	216008	P68
A69	5-400-149	215988	216012	P69
A70	5-400-175	216014	216038	P70
A71	5-400-231	216070	216094	P71
A72	5-400-367	216206	216230	P72
A73	99-12852-110	216310	216334	P73
A74	99-12852-325	216525	216549	P74
A75	4-37-326	221637	221661	P75
A76	4-37-107	221855	221879	P76
A77	5-270-92	225633	225657	P77
A78	99-12860-47	229375	229399	P78
A79	99-12860-57	229385	229409	P79
A80	5-402-144	237543	237567	P80

EXAMPLE 4

VALIDATION OF THE POLYMORPHISMS THROUGH MICROSEQUENCING

The biallelic markers identified in example 3 were further confirmed and their respective frequencies were determined through microsequencing. Microsequencing was carried out for each individual DNA sample described in Example 1.

Amplification from genomic DNA of individuals was performed by PCR as described above for the detection of the biallelic markers with the same set of PCR primers (Table 1).

The preferred primers used in microsequencing were about 19 nucleotides in length and hybridized just upstream of the considered polymorphic base. According to the invention, the primers used in microsequencing are detailed in Table 4.

Table 4

Marker name	BM	Mis 1	Position range of microsequencing primer mis 1 in SEQ ID No 1		Mis 2	Complementary position range of microsequencing primer mis. 2 in SEQ ID No 1	
5-390-177	A1	D1	1980	1998	E1	2000	2018
5-391-43	A2	D2	4582	4600	E2	4602	4620
5-392-222	A3	D3	10209	10227	E3	10229	10247
5-392-280	A4	D4	10267	10285	E4	10287	10305
4-58-318	A6	D6	39925	39943	E6	39945	39963
4-58-289	A7	D7	39954	39972	E7	39974	39992
4-54-199	A8	D8	41366	41384	E8	41386	41404
4-54-180	A9	D9	41385	41403	E9	41405	41423
4-51-312	A10	D10	42213	42231	E10	42233	42251
99-86-266	A11	D11	67456	67474	E11	67476	67494
4-88-107	A12	D12	69502	69520	E12	69522	69540
5-397-141	A13	D13	72819	72837	E13	72839	72857
5-398-203	A14	D14	76041	76059	E14	76061	76079
99-12738-248	A15	D15	81234	81252	E15	81254	81272
99-109-358	A16	D16	83902	83920	E16	83922	83940
99-12749-175	A17	D17	91898	91916	E17	91918	91936
4-21-154	A18	D18	95330	95348	E18	95350	95368
4-21-317	A19	D19	95492	95510	E19	95512	95530
4-23-326	A20	D20	96171	96189	E20	96191	96209
99-12753-34	A21	D21	97275	97293	E21	97295	97313
5-364-252	A22	D22	98005	98023	E22	98025	98043
99-12755-280	A23	D23	98895	98913	E23	98915	98933
99-12755-329	A24	D24	98944	98962	E24	98964	98982
4-87-212	A25	D25	103574	103592	E25	103594	103612
99-12757-318	A26	D26	104379	104397	E26	104399	104417
99-12758-102	A27	D27	106354	106372	E27	106374	106392
99-12758-136	A28	D28	106388	106406	E28	106408	106426
4-105-98	A29	D29	108296	108314	E29	108316	108334

4-105-86	A30	D30	108308	108326	E30	108328	108346
4-45-49	A31	D31	108453	108471	E31	108473	108491
4-44-277	A32	D32	109177	109195	E32	109197	109215
4-86-60	A33	D33	114585	114603	E33	114605	114623
4-84-334	A34	D34	115697	115715	E34	115717	115735
99-78-321	A35	D35	122064	122082	E35	122084	122102
99-12767-36	A36	D36	123105	123123	E36	123125	123143
99-12767-143	A37	D37	123212	123230	E37	123232	123250
99-12767-189	A38	D38	123258	123276	E38	123278	123296
99-12767-380	A39	D39	123449	123467	E39	123469	123487
4-80-328	A40	D40	126719	126737	E40	126739	126757
4-36-384	A41	D41	128191	128209	E41	128211	128229
4-36-264	A42	D42	128311	128329	E42	128331	128349
4-36-261	A43	D43	128314	128332	E43	128334	128352
4-35-333	A44	D44	128575	128593	E44	128595	128613
4-35-240	A45	D45	128668	128686	E45	128688	128706
4-35-173	A46	D46	128735	128753	E46	128755	128773
4-35-133	A47	D47	128775	128793	E47	128795	128813
99-12771-59	A48	D48	130786	130804	E48	130806	130824
99-12774-334	A49	D49	133187	133205	E49	133207	133225
99-12776-358	A50	D50	135367	135385	E50	135387	135405
99-12781-113	A51	D51	139370	139388	E51	139390	139408
4-104-298	A52	D52	157516	157534	E52	157536	157554
4-104-254	A53	D53	157560	157578	E53	157580	157598
4-104-250	A54	D54	157564	157582	E54	157584	157602
4-104-214	A55	D55	157600	157618	E55	157620	157638
99-12818-289	A56	D56	172961	172979	E56	172981	172999
99-24807-271	A57	D57	180603	180621	E57	180623	180641
99-24807-84	A58	D58	180790	180808	E58	180810	180828
99-12831-157	A59	D59	190315	190333	E59	190335	190353
99-12831-241	A60	D60	190399	190417	E60	190419	190437
99-12832-387	A61	D61	191378	191396	E61	191398	191416
99-12836-30	A62	D62	195109	195127	E62	195129	195147
99-12844-262	A63	D63	203827	203845	E63	203847	203865
4-24-74	A64	D64	210132	210150	E64	210152	210170
4-24-246	A65	D65	210302	210320	E65	210322	210340
4-24-314	A66	D66	210370	210388	E66	210390	210408
4-27-190	A67	D67	211149	211167	E67	211169	211187
5-400-145	A68	D68	215977	215995	E68	215997	216015
5-400-149	A69	D69	215981	215999	E69	216001	216019
5-400-175	A70	D70	216007	216025	E70	216027	216045
5-400-231	A71	D71	216063	216081	E71	216083	216101
5-400-367	A72	D72	216199	216217	E72	216219	216237
99-12852-110	A73	D73	216303	216321	E73	216323	216341
99-12852-325	A74	D74	216518	216536	E74	216538	216556
4-37-326	A75	D75	221630	221648	E75	221650	221668

4-37-107	A76	D76	221848	221866	E76	221868	221886
5-270-92	A77	D77	225626	225644	E77	225646	225664
99-12860-47	A78	D78	229368	229386	E78	229388	229406
99-12860-57	A79	D79	229378	229396	E79	229398	229416
5-402-144	A80	D80	237536	237554	E80	237556	237574

Mis 1 and Mis 2 respectively refer to microsequencing primers which hybridized with the non-coding strand of the PG-3 gene or with the coding strand of the PG-3 gene.

The microsequencing reaction was performed as follows :

- 5 After purification of the amplification products, the microsequencing reaction mixture was prepared by adding, in a 20µl final volume: 10 pmol microsequencing oligonucleotide, 1 U Thermosequenase (Amersham E79000G), 1.25 µl Thermosequenase buffer (260 mM Tris HCl pH 9.5, 65 mM MgCl₂), and the two appropriate fluorescent ddNTPs (Perkin Elmer, Dye Terminator Set 401095) complementary to the nucleotides at the polymorphic site of each biallelic marker
- 10 tested, following the manufacturer's recommendations. After 4 minutes at 94°C, 20 PCR cycles of 15 sec at 55°C, 5 sec at 72°C, and 10 sec at 94°C were carried out in a Tetrad PTC-225 thermocycler (MJ Research). The unincorporated dye terminators were then removed by ethanol precipitation. Samples were finally resuspended in formamide-EDTA loading buffer and heated for 2 min at 95°C before being loaded on a polyacrylamide sequencing gel. The data were collected by
- 15 an ABI PRISM 377 DNA sequencer and processed using the GENESCAN software (Perkin Elmer).

Following gel analysis, data were automatically processed with software that allows the determination of the alleles of biallelic markers present in each amplified fragment.

- The software evaluates such factors as whether the intensities of the signals resulting from the above microsequencing procedures are weak, normal, or saturated, or whether the signals are
- 20 ambiguous. In addition, the software identifies significant peaks (according to shape and height criteria). Among the significant peaks, peaks corresponding to the targeted site are identified based on their position. When two significant peaks are detected for the same position, each sample is categorized classification as homozygous or heterozygous type based on the height ratio.

EXAMPLE 5

- 25 PREPARATION OF ANTIBODY COMPOSITIONS TO THE PG-3 PROTEIN

- Substantially pure protein or polypeptide is isolated from transfected or transformed cells containing an expression vector encoding the PG-3 protein or a portion thereof. The concentration of protein in the final preparation is adjusted, for example, by concentration on an Amicon filter device, to the level of a few micrograms/ml. Monoclonal or polyclonal antibody to the protein can then be
- 30 prepared as follows:

A. Monoclonal Antibody Production by Hybridoma Fusion

Monoclonal antibody to epitopes in the PG-3 protein or a portion thereof can be prepared from murine hybridomas according to the classical method of Kohler, G. and Milstein, C., (1975) or derivative methods thereof. Also see Harlow, E., and D. Lane. 1988.

Briefly, a mouse is repetitively inoculated with a few micrograms of the PG-3 protein or a portion thereof over a period of a few weeks. The mouse is then sacrificed, and the antibody producing cells of the spleen isolated. The spleen cells are fused by means of polyethylene glycol with mouse myeloma cells, and the excess unfused cells destroyed by growth of the system on selective media comprising aminopterin (HAT media). The successfully fused cells are diluted and aliquots of the dilution placed in wells of a microtiter plate where growth of the culture is continued. Antibody-producing clones are identified by detection of antibody in the supernatant fluid of the wells by immunoassay procedures, such as ELISA, as originally described by Engvall, (1980), and derivative methods thereof. Selected positive clones can be expanded and their monoclonal antibody product harvested for use. Detailed procedures for monoclonal antibody production are described in Davis, L. *et al.* (1986).

15 B. Polyclonal Antibody Production by Immunization

Polyclonal antiserum containing antibodies to heterogeneous epitopes in the PG-3 protein or a portion thereof can be prepared by immunizing suitable non-human animal with the PG-3 protein or a portion thereof, which can be unmodified or modified to enhance immunogenicity. A suitable non-human animal is preferably a non-human mammal is selected, usually a mouse, rat, rabbit, goat, or horse. Alternatively, a crude preparation which has been enriched for PG-3 concentration can be used to generate antibodies. Such proteins, fragments or preparations are introduced into the non-human mammal in the presence of an appropriate adjuvant (e.g. aluminum hydroxide, RIBI, etc.) which is known in the art. In addition the protein, fragment or preparation can be pretreated with an agent which will increase antigenicity, such agents are known in the art and include, for example, methylated bovine serum albumin (mBSA), bovine serum albumin (BSA), Hepatitis B surface antigen, and keyhole limpet hemocyanin (KLH). Serum from the immunized animal is collected, treated and tested according to known procedures. If the serum contains polyclonal antibodies to undesired epitopes, the polyclonal antibodies can be purified by immunoaffinity chromatography.

Effective polyclonal antibody production is affected by many factors related both to the antigen and the host species. Also, host animals vary in response to site of inoculations and dose, with both inadequate or excessive doses of antigen resulting in low titer antisera. Small doses (ng level) of antigen administered at multiple intradermal sites appears to be most reliable. Techniques for producing and processing polyclonal antisera are known in the art, see for example, Mayer and Walker (1987). An effective immunization protocol for rabbits can be found in Vaitukaitis, J. *et al.* (1971).

Booster injections can be given at regular intervals, and antiserum harvested when antibody titer thereof, as determined semi-quantitatively, for example, by double immunodiffusion in agar against known concentrations of the antigen, begins to fall. See, for example, Ouchterlony, O. *et al.*, (1973). Plateau concentration of antibody is usually in the range of 0.1 to 0.2 mg/ml of serum
5 (about 12 μ M). Affinity of the antisera for the antigen is determined by preparing competitive binding curves, as described, for example, by Fisher, D., (1980).

Antibody preparations prepared according to either the monoclonal or the polyclonal protocol are useful in quantitative immunoassays which determine concentrations of antigen-bearing substances in biological samples; they are also used semi-quantitatively or qualitatively to identify the presence of
10 antigen in a biological sample. The antibodies may also be used in therapeutic compositions for killing cells expressing the protein or reducing the levels of the protein in the body.

While the preferred embodiment of the invention has been illustrated and described, it will be appreciated that various changes can be made therein by the one skilled in the art without
15 departing from the spirit and scope of the invention.

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- 30

SEQUENCE LISTING FREE TEXT

The following free text appears in the accompanying Sequence Listing :

5' regulatory region

3' regulatory region

35 polymorphic base

or

complement

probe

sequencing oligonucleotide primer

insertion of

exon

WHAT IS CLAIMED:

1. An isolated, purified, or recombinant polynucleotide comprising a contiguous span of at least 15 nucleotides of SEQ ID No 1 or the complements thereof, wherein said contiguous span comprises at least one of the following nucleotide positions of SEQ ID No 1: 1-97921, 98517-
5 103471, 103603-108222, 108390-109221, 109324-114409, 114538-115723, 115957-122102, 122225-126876, 127033-157212, 157808-240825.
2. An isolated, purified, or recombinant polynucleotide comprising a contiguous span of at least 15 nucleotides of SEQ ID No 2 or the complements thereof.
- 10 3. An isolated, purified, or recombinant polynucleotide consisting essentially of a contiguous span of at least 15 nucleotides of anyone of SEQ ID Nos 1 and 2 or the complement thereof, wherein said span includes a PG-3-related biallelic marker in said sequence.
- 15 4. A polynucleotide according to claim 3, wherein said PG-3-related biallelic marker is selected from the group consisting of A1 to A80, and the complements thereof.
5. A polynucleotide according to claim 3, wherein said PG-3-related biallelic marker is selected from the group consisting of A1 to A5 and A8 to A80, and the complements thereof.
- 20 6. A polynucleotide according to claim 3, wherein said PG-3-related biallelic marker is selected from the group consisting of A6 and A7, and the complements thereof.
7. A polynucleotide according to claim 3, wherein said contiguous span is 18 to 35
25 nucleotides in length and said biallelic marker is within 4 nucleotides of the center of said polynucleotide.
8. A polynucleotide according to claim 7, wherein said polynucleotide consists of said contiguous span and said contiguous span is 25 nucleotides in length and said biallelic marker is at
30 the center of said polynucleotide.
9. A polynucleotide according to claim 7, wherein said polynucleotide consists essentially of a sequence selected from the following sequences: P1 to P4 and P6 to P80, and the complementary sequences thereto.
- 35 10. A polynucleotide according to any one of claims 1, 2 or 3, wherein the 3' end of said contiguous span is present at the 3' end of said polynucleotide.

11. A polynucleotide according to claim 3, wherein the 3' end of said contiguous span is located at the 3' end of said polynucleotide and said biallelic marker is present at the 3' end of said polynucleotide.

5

12. An isolated, purified, or recombinant polynucleotide consisting essentially of a contiguous span of at least 15 nucleotides of anyone of SEQ ID No 1,2 or the complements thereof, wherein the 3' end of said contiguous span is located at the 3' end of said polynucleotide, and wherein the 3' end of said polynucleotide is located within 20 nucleotides upstream of a PG-3-
10 related biallelic marker in said sequence.

13. A polynucleotide according to claim 12, wherein the 3' end of said polynucleotide is located one nucleotide upstream of said PG-3-related biallelic marker in said sequence.

14. A polynucleotide according to claim 13, wherein said polynucleotide consists essentially of a sequence selected from the following sequences: D1 to D4, D6 to D80, E1 to E4, and E6 to E80.

15. An isolated, purified, or recombinant polynucleotide consisting essentially of a
20 sequence selected from the following sequences: B1 to B52 and C1 to C52.

16. An isolated, purified, or recombinant polynucleotide which encodes a polypeptide comprising a contiguous span of at least 6 amino acids of SEQ ID No 3.

17. A polynucleotide according to any one of claims 1-16 attached to a solid support.

18. An array of polynucleotides comprising at least one polynucleotide according to claim
17.

19. An array according to claim 18, wherein said array is addressable.

20. A polynucleotide according to any one of claims 1-16 further comprising a label.

21. A recombinant vector comprising a polynucleotide according to any one of claims 1-
35 16.

22. A host cell comprising a recombinant vector according to claim 21.

23. A non-human host animal or mammal comprising a recombinant vector according to claim 21.

5 24. A mammalian host cell comprising a PG-3 gene disrupted by homologous recombination with a knock out vector, comprising a polynucleotide according to any one of claims 1- 16.

25. A non-human host mammal comprising a PG-3 gene disrupted by homologous
10 recombination with a knock out vector, comprising a polynucleotide according to any one of claims 1-16.

26. A method of genotyping comprising determining the identity of a nucleotide at a PG-3-
related biallelic marker or the complement thereof in a biological sample.
15

27. A method according to claim 26, wherein said biological sample is derived from a single subject.

28. A method according to claim 27, wherein the identity of the nucleotides at said biallelic
20 marker is determined for both copies of said biallelic marker present in said individual's genome.

29. A method according to claim 26, wherein said biological sample is derived from multiple subjects.

25 30. A method according to claim 26, further comprising amplifying a portion of said sequence comprising the biallelic marker prior to said determining step.

31. A method according to claim 30, wherein said amplifying is performed by PCR.

30 32. A method according to claim 26, wherein said determining is performed by a hybridization assay.

33. A method according to claim 26, wherein said determining is performed by a sequencing assay.
35

34. A method according to claim 26, wherein said determining is performed by a microsequencing assay.

35. A method according to claim 26, wherein said determining is performed by an enzyme-based mismatch detection assay.

5 36. A method of estimating the frequency of an allele of a PG-3-related biallelic marker in a population comprising:

 a) genotyping individuals from said population for said biallelic marker according to the method of claim 26; and

 b) determining the proportional representation of said biallelic marker in said population.
10

37. A method of detecting an association between a genotype and a trait, comprising the steps of:

 a) determining the frequency of at least one PG-3-related biallelic marker in trait positive population according to the method of claim 36;

15 b) determining the frequency of at least one PG-3-related biallelic marker in a control population according to the method of claim 36; and

 c) determining whether a statistically significant association exists between said genotype and said trait.

20 38. A method of estimating the frequency of a haplotype for a set of biallelic markers in a population, comprising:

 a) genotyping at least one PG-3-related biallelic marker according to claim 27 for each individual in said population;

 b) genotyping a second biallelic marker by determining the identity of the nucleotides at
25 said second biallelic marker for both copies of said second biallelic marker present in the genome of each individual in said population; and

 c) applying a haplotype determination method to the identities of the nucleotides determined in steps a) and b) to obtain an estimate of said frequency.

30 39. A method according to claim 38, wherein said haplotype determination method is selected from the group consisting of asymmetric PCR amplification, double PCR amplification of specific alleles, the Clark algorithm, or an expectation-maximization algorithm.

40. A method of detecting an association between a haplotype and a trait, comprising the
35 steps of:

 a) estimating the frequency of at least one haplotype in a trait positive population according to the method of claim 38;

b) estimating the frequency of said haplotype in a control population according to the method of claim 38; and

c) determining whether a statistically significant association exists between said haplotype and said trait.

5

41. A method according to claim 37, wherein said genotyping steps a) and b) are performed on a single pooled biological sample derived from each of said populations.

42. A method according to claim 37, wherein said genotyping steps a) and b) performed
10 separately on biological samples derived from each individual in said populations.

43. A method according to either claim 37 or 40, wherein said trait is cancer susceptibility.

44. A method according to either claim 37 or 40, wherein said control population is a trait
15 negative population.

45. A method according to either claim 37 or 40, wherein said case control population is a random population.

20 46. Use of a polynucleotide comprising a contiguous span of at least 15 nucleotides of a sequence selected from the group consisting of the SEQ ID Nos 1, 2, amplicons 5-390, 5-391, 5-392, 4-59, 4-58, 4-54, 4-51, 99-86, 4-88, 5-397, 5-398, 99-12738, 99-109, 99-12749, 4-21, 4-23, 99-12753, 5-364, 99-12755, 4-87, 99-12757, 99-12758, 4-105, 4-45, 4-44, 4-86, 4-84, 99-78, 99-12767, 4-80, 4-36, 4-35, 99-12771, 99-12774, 99-12776, 99-12781, 4-104, 99-12818, 99-24807, 99-
25 12827, 99-12831, 99-12832, 99-12836, 99-12844, 4-24, 4-27, 5-400, 99-12852, 4-37, 5-270, 99-12860, and 5-402 or the complementary sequence thereto for determining the identity of the nucleotide at a PG-3-related biallelic marker

30 47. Use according to claim 46 in a microsequencing assay, wherein the 3' end of said contiguous span is located at the 3' end of said polynucleotide and wherein the 3' end of said polynucleotide is located 1 nucleotide upstream of said PG-3-related biallelic marker in said sequence.

35 48. Use according to claim 46 in a hybridization assay, wherein said c ntiguous span includes said PG-3-related biallelic marker.

49. Use according to claim 46 in a specific amplification assay, wherein the 3' end of said contiguous span is located at the 3' end of said polynucleotide and said biallelic marker is present at the 3' end of said polynucleotide.

50. Use according to claim 46 in a sequencing assay, wherein the 3' end of said contiguous span is located at the 3' end of said polynucleotide.

51. Use according to any one of claims 46-50, wherein said PG-3-related biallelic is a biallelic marker selected from the group consisting of A1 to A80.

10

52. An isolated, purified, or recombinant polypeptide comprising a contiguous span of at least 6 amino acids of SEQ ID No 3.

53. An isolated or purified antibody composition capable of selectively binding to an epitope-containing fragment of a polypeptide according to claim 52.

15

54. A method according to any one of claims 26-45, wherein said PG-3-related biallelic marker is selected from the group consisting of A1 to A80 and the complements thereof.

55. A diagnostic kit comprising a polynucleotide according to any one of claims 3-15.

20

56. A computer readable medium having stored thereon a sequence selected from the group consisting of a nucleic acid code comprising one of the following:

a) a contiguous span of at least 15 nucleotides of SEQ ID No 1, wherein said contiguous span comprises at least one of the following nucleotide positions of SEQ ID No 1: 1-97921, 98517-103471, 103603-108222, 108390-109221, 109324-114409, 114538-115723, 115957-122102, 122225-126876, 127033-157212, 157808-240825;

25

b) a contiguous span of at least 15 nucleotides of SEQ ID No 2 or the complements thereof; and

c) a nucleotide sequence complementary to any one of the preceding nucleotide sequences.

30

57. A computer readable medium having stored thereon a sequence consisting of a polypeptide code comprising a contiguous span of at least 6 amino acids of SEQ ID No 3.

58. A computer system comprising a processor and a data storage device wherein said data storage device is a computer readable medium according to claim 56 or 57.

35

59. A computer system according to claim 58, further comprising a sequence comparer and a data storage device having reference sequences stored thereon.

60. A computer system of Claim 59 wherein said sequence comparer comprises a computer
5 program which indicates polymorphisms.

61. A computer system of Claim 58 further comprising an identifier which identifies features in said sequence.

10 62. A method for comparing a first sequence to a reference sequence, comprising the steps of:

reading said first sequence and said reference sequence through use of a computer program which compares sequences; and

determining differences between said first sequence and said reference sequence with said
15 computer program,

wherein said first sequence is selected from the group consisting of a nucleic acid code comprising one of the following:

a) a contiguous span of at least 15 nucleotides of SEQ ID No 1, wherein said contiguous span comprises at least one of the following nucleotide positions of SEQ ID No 1: 1-97921, 98517-
20 103471, 103603-108222, 108390-109221, 109324-114409, 114538-115723, 115957-122102, 122225-126876, 127033-157212, 157808-240825;

b) a contiguous span of at least 15 nucleotides of SEQ ID No 2 or the complements thereof;
and

c) a nucleotide sequence complementary to any one of the preceding nucleotide sequences;
25 and,

d) a polypeptide code comprising a contiguous span of at least 6 amino acids of SEQ ID No
3.

AMENDED CLAIMS

[received by the International Bureau on 25 January 2001 (25.01.01);
original claims 1,2 and 56 amended; remaining claims unchanged (2 pages)]

1. An isolated, purified, or recombinant polynucleotide comprising a contiguous span of at least 200 nucleotides of SEQ ID No 1 or the complements thereof, wherein said contiguous span comprises at least one of the following nucleotide positions of SEQ ID No 1: 1-97921, 98517-
5 103471, 103603-108222, 108390-109221, 109324-114409, 114538-115723, 115957-122102, 122225-126876, 127033-157212, 157808-240825.

2. An isolated, purified, or recombinant polynucleotide comprising a contiguous span of at least 200 nucleotides of SEQ ID No 2 or the complements thereof.

10

3. An isolated, purified, or recombinant polynucleotide consisting essentially of a contiguous span of at least 15 nucleotides of anyone of SEQ ID Nos 1 and 2 or the complement thereof, wherein said span includes a PG-3-related biallelic marker in said sequence.

15 4. A polynucleotide according to claim 3, wherein said PG-3-related biallelic marker is selected from the group consisting of A1 to A80, and the complements thereof.

5. A polynucleotide according to claim 3, wherein said PG-3-related biallelic marker is selected from the group consisting of A1 to A5 and A8 to A80, and the complements thereof.

20

6. A polynucleotide according to claim 3, wherein said PG-3-related biallelic marker is selected from the group consisting of A6 and A7, and the complements thereof.

7. A polynucleotide according to claim 3, wherein said contiguous span is 18 to 35
25 nucleotides in length and said biallelic marker is within 4 nucleotides of the center of said polynucleotide.

8. A polynucleotide according to claim 7, wherein said polynucleotide consists of said contiguous span and said contiguous span is 25 nucleotides in length and said biallelic marker is at
30 the center of said polynucleotide.

9. A polynucleotide according to claim 7, wherein said polynucleotide consists essentially of a sequence selected from the following sequences: P1 to P4 and P6 to P80, and the complementary sequences thereto.

35

10. A polynucleotide according to any one of claims 1, 2 or 3, wherein the 3' end of said contiguous span is present at the 3' end of said polynucleotide.

49. Use according to claim 46 in a specific amplification assay, wherein the 3' end of said contiguous span is located at the 3' end of said polynucleotide and said biallelic marker is present at the 3' end of said polynucleotide.

5 50. Use according to claim 46 in a sequencing assay, wherein the 3' end of said contiguous span is located at the 3' end of said polynucleotide.

51. Use according to any one of claims 46-50, wherein said PG-3-related biallelic is a biallelic marker selected from the group consisting of A1 to A80.

10

52. An isolated, purified, or recombinant polypeptide comprising a contiguous span of at least 6 amino acids of SEQ ID No 3.

53. An isolated or purified antibody composition capable of selectively binding to an epitope-containing fragment of a polypeptide according to claim 52.

15

54. A method according to any one of claims 26-45, wherein said PG-3-related biallelic marker is selected from the group consisting of A1 to A80 and the complements thereof.

20 55. A diagnostic kit comprising a polynucleotide according to any one of claims 3-15.

56. A computer readable medium having stored thereon at least 2 nucleic acid code sequences comprising any one of the following:

25 a) a contiguous span of at least 200 nucleotides of SEQ ID No 1, wherein said contiguous span comprises at least one of the following nucleotide positions of SEQ ID No 1: 1-97921, 98517-103471, 103603-108222, 108390-109221, 109324-114409, 114538-115723, 115957-122102, 122225-126876, 127033-157212, 157808-240825;

b) a contiguous span of at least 200 nucleotides of SEQ ID No 2 or the complements thereof; and

30 c) a nucleotide sequence complementary to any one of the preceding nucleotide sequences.

57. A computer readable medium having stored thereon a sequence consisting of a polypeptide code comprising a contiguous span of at least 6 amino acids of SEQ ID No 3.

35 58. A computer system comprising a processor and a data storage device wherein said data storage device is a computer readable medium according to claim 56 or 57.

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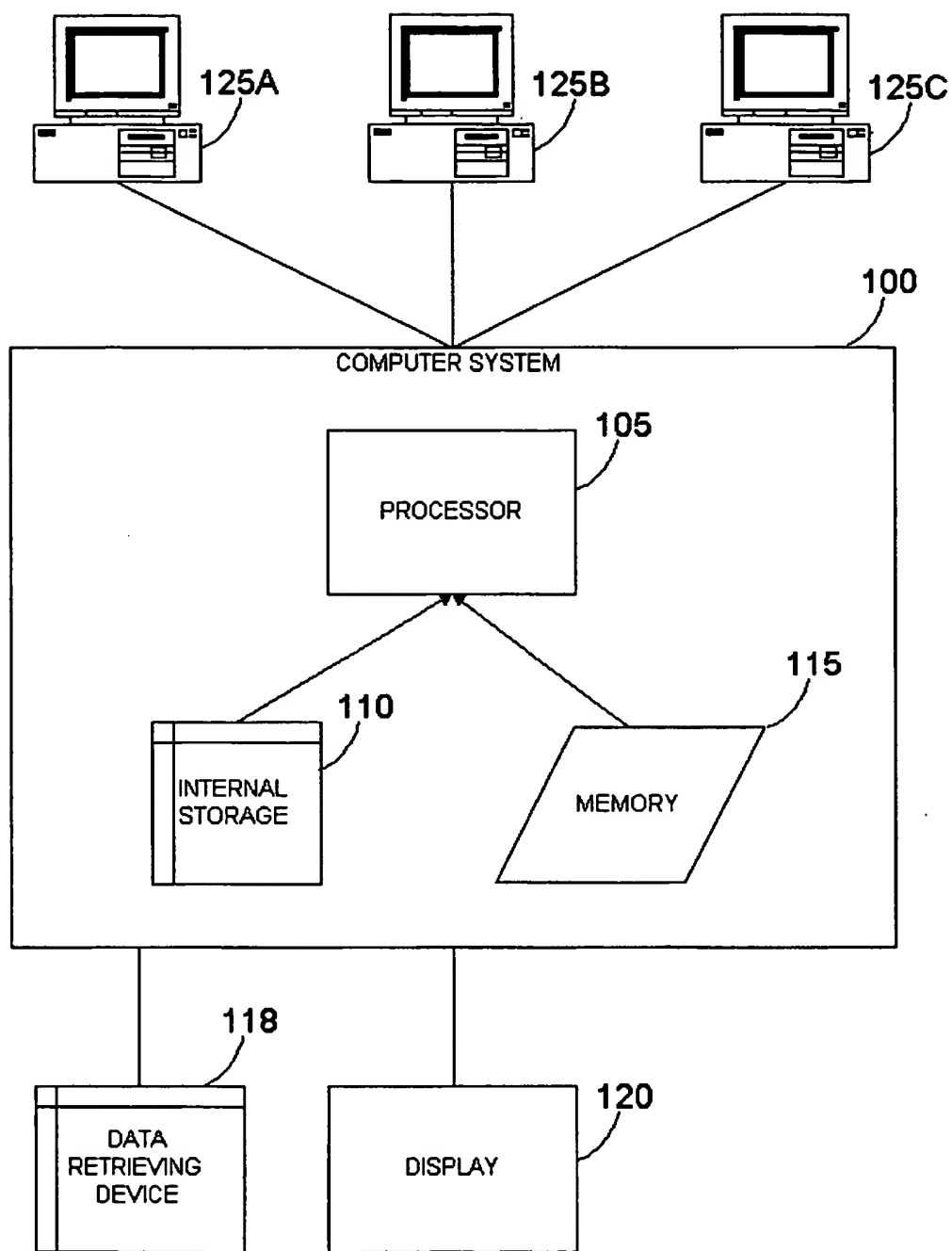


Figure 1

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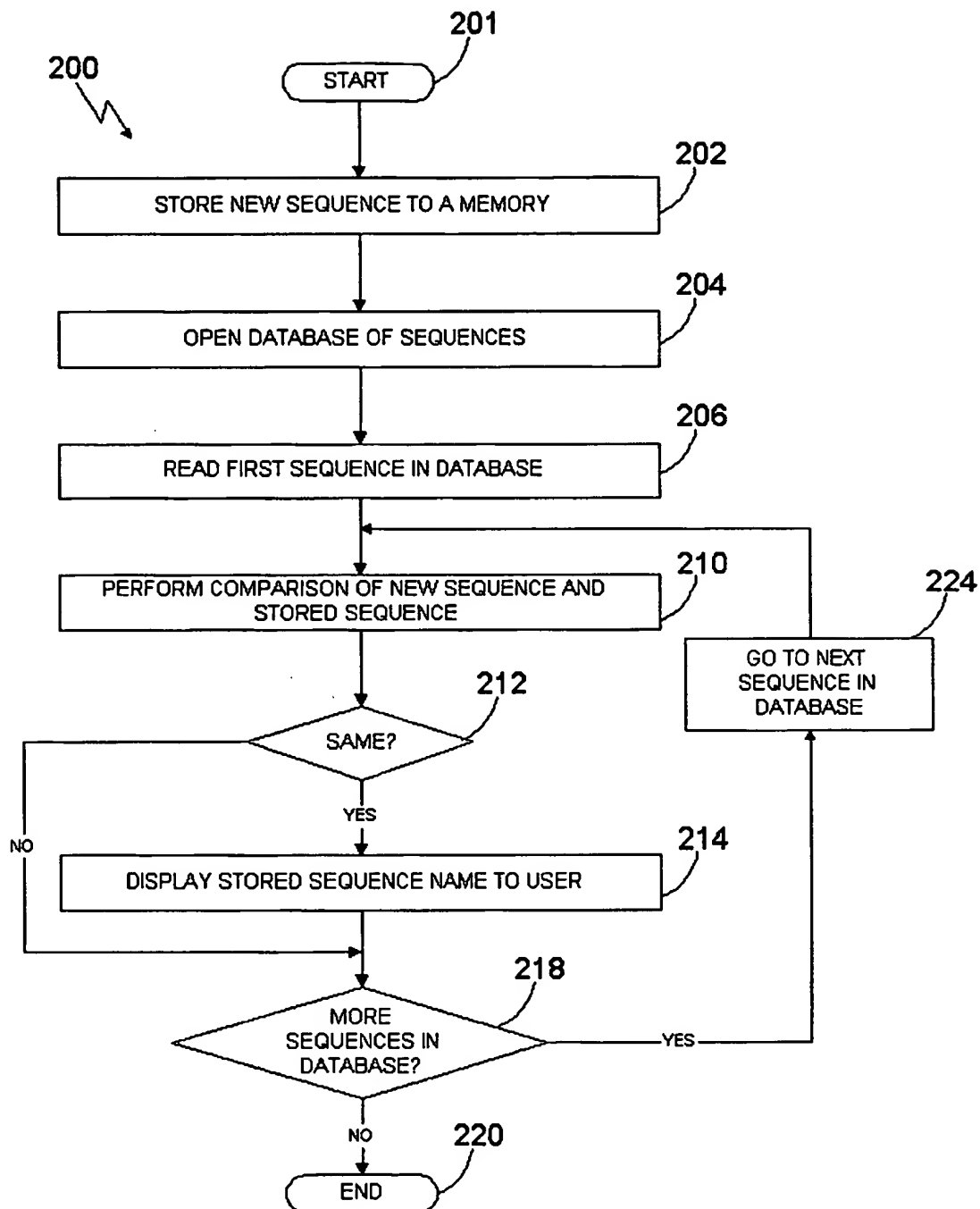


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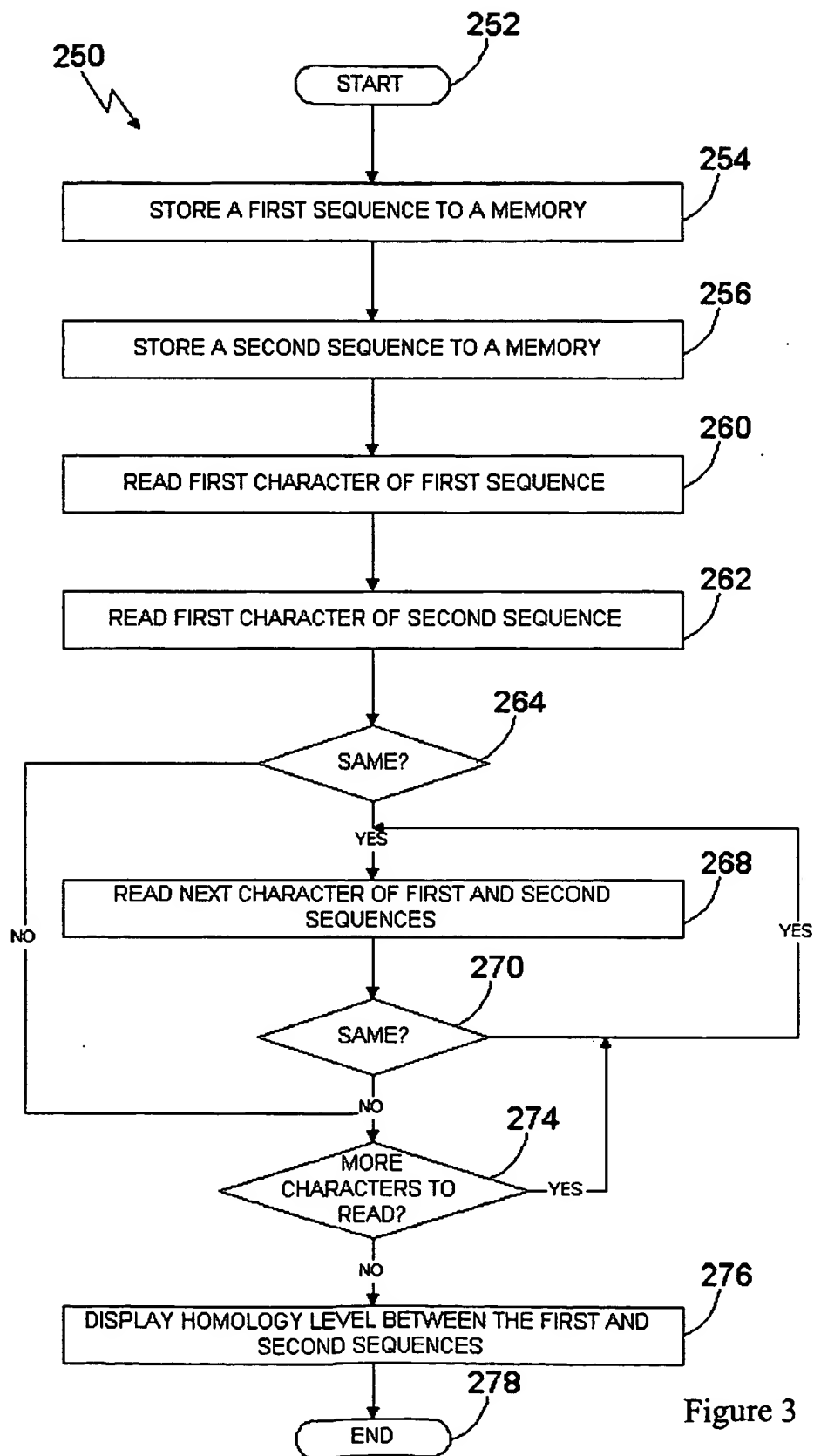
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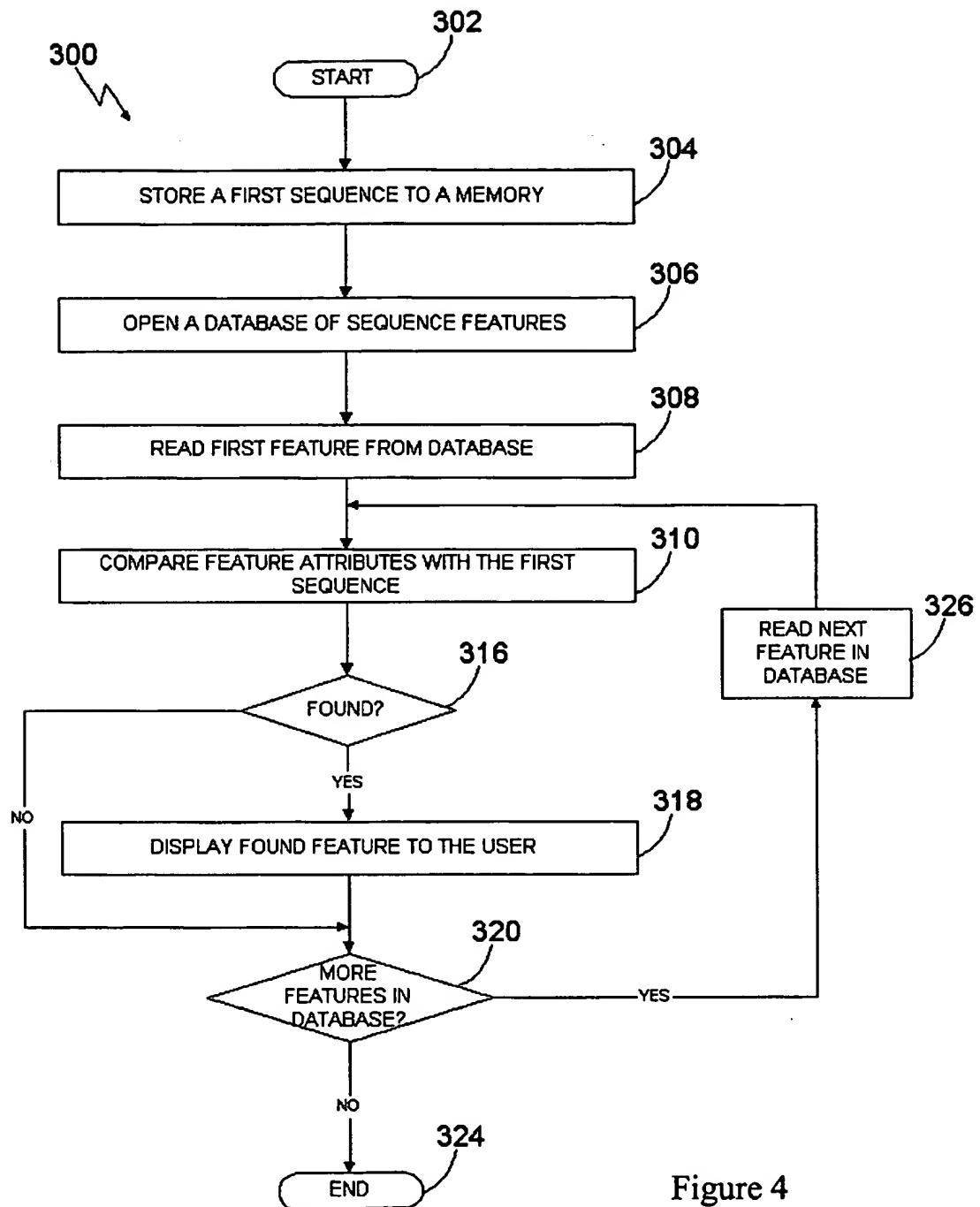


Figure 4

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Ile	Tyr	Thr	Pro	Thr	Ile	Glu	Ile	Asn	Ser	Ser	His	His	Ser	Ala	Met	
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Ser	Ser	Thr	Lys	Gly	His	Leu	Leu	Ile	His	Ser	Arg	Pro	Arg	Ser	Ser	
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105

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Cys	Trp	Val	Leu	Ser	Tyr	Asp	Trp	Val	Leu	Trp	Ser	Leu	Glu	Leu	Gly	
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Gly Thr Leu Phe Ala Asp Gln Pro Xaa Met Phe Val Ser Pro Ala Ser
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Ser Pro Pro Val Ala Lys Leu Cys Glu Leu Val His Leu Cys Gly Gly
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Ile	Tyr	Thr	Pro	Thr	Ile	Glu	Ile	Asn	Ser	Ser	His	His	Ser	Ala	Met
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Ser	Gly	Lys	Lys	Lys	Ala	Thr	Val	Lys	Tyr	Leu	Ser	Glu	Lys	Trp	Val
805					810					815					
Leu	Asp	Ser	Ile	Thr	Gln	His	Lys	Val	Cys	Ala	Xaa	Glu	Asn	Tyr	Leu
820					825					830					
Leu	Ser	Gln													
835															

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<212> DNA
<213> Artificial Sequence
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<220>
<223> sequencing oligonucleotide PrimerRP
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INTERNATIONAL SEARCH REPORT

International Application No

PCT/IB 00/01098

A. CLASSIFICATION OF SUBJECT MATTER

IPC 7 C12N15/12 C07K14/47 C12Q1/68 C07K16/18 A61K67/027

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 7 C12N C07K C12Q A61K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

EPO-Internal

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	DATABASE EMBL - EM_GSS 'Online! Entry B88009, 10 March 1998 (1998-03-10) ADAMS, M.D. ET AL.: "RPCI11-16G12.TVB RPCI-11 Homo sapiens genomic clone RPCI-11-16G12, genomic survey sequence." XP002152821 the whole document ---	1,10, 17-22, 56,58-62
X	DATABASE EMBL - EMBEST_HUM7 'Online! Entry/Acc.no. AI829673, 13 July 1999 (1999-07-13) STRAUSBERG, R.: "wf09b04.x1 Soares_NFL_T_GBC_S1 Homo sapiens cDNA clone IMAGE:2350063 3', mRNA sequence." XP002152822 the whole document --- -/--	2-5,10, 17-22, 56,58-62



Further documents are listed in the continuation of box C.



Patent family members are listed in annex.

* Special categories of cited documents:

A document defining the general state of the art which is not considered to be of particular relevance

E earlier document but published on or after the international filing date

L document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

O document referring to an oral disclosure, use, exhibition or other means

P document published prior to the international filing date but later than the priority date claimed

T later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

X document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

Y document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.

& document member of the same patent family

Date of the actual completion of the international search

15 November 2000

Date of mailing of the international search report

28/11/2000

Name and mailing address of the ISA

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INTERNATIONAL SEARCH REPORT

International Application No

PCT/IB 00/01098

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	DATABASE EMBL - EMBL_HUM15 'Online! Entry HS611123, Acc.no. R56611, 28 May 1995 (1995-05-28) HILLIER, L. ET AL.: "yg94b12.r1 Soares infant brain 1NIB Homo sapiens cDNA clone IMAGE:41184 5', mRNA sequence." XP002152823 the whole document ----	2-4,6, 17-22, 56,58-62
A	WO 99 32644 A (BOUGUELERET LYDIE ;CHUMAKOV ILYA (FR); COHEN DANIEL (FR); GENSET () 1 July 1999 (1999-07-01) the whole document ----	
A	WO 98 45420 A (DIAGNOCURE INC ;BUSSEMAKERS MARION J G (NL)) 15 October 1998 (1998-10-15) the whole document ----	
A	WO 99 37986 A (UNIV SOUTHERN CALIFORNIA) 29 July 1999 (1999-07-29) the whole document ----	
P,X	WO 00 09552 A (GENETICS INST) 24 February 2000 (2000-02-24) whole document, particularly passages relating to seq.ID.157. -----	2-5,16, 56-62

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

Continuation of Box I.1

Although claims 56-61 could be at least partially considered as a mere presentation of information (Rule 39.1.(v) PCT), and claims 59-62 at least partially as a computer program (Rule 39.1.(vi) PCT), the search has been carried out as far as possible in our systematic documentation.

INTERNATIONAL SEARCH REPORT

information on patent family members

International Application No

PCT/IB 00/01098

Patent document cited in search report		Publication date	Patent family member(s)	Publication date
WO 9932644	A	01-07-1999	US 5945522 A	31-08-1999
			AU 1574099 A	12-07-1999
			EP 1052292 A	15-11-2000
			EP 0991770 A	12-04-2000
WO 9845420	A	15-10-1998	AU 7019498 A	30-10-1998
			EP 1007650 A	14-06-2000
WO 9937986	A	29-07-1999	AU 2238499 A	09-08-1999
WO 0009552	A	24-02-2000	AU 5557099 A	06-03-2000

Before describing the invention in greater detail, the following definitions are set forth to illustrate and define the meaning and scope of the terms used to describe the invention herein.

The terms "PG-3 gene", when used herein, encompasses genomic, mRNA and cDNA sequences encoding the PG-3 protein, including the untranscribed regulatory regions of the genomic DNA.

The term "heterologous protein", when used herein, is intended to designate any protein or polypeptide other than the PG-3 protein. More particularly, the heterologous protein may be a compound which can be used as a marker in further experiments with a PG-3 regulatory region.

The term "isolated" requires that the material be removed from its original environment (e.g., the natural environment if it is naturally occurring). For example, a naturally-occurring polynucleotide or polypeptide present in a living animal is not isolated, but the same polynucleotide or DNA or polypeptide, separated from some or all of the coexisting materials in the natural system, is isolated. Such a polynucleotide could be part of a vector and/or such a polynucleotide or polypeptide could be part of a composition, and still be isolated in that the vector or composition is not part of its natural environment.

The term "purified" does not require absolute purity; rather, it is intended as a relative definition. Purification of starting material or natural material to at least one order of magnitude, preferably two or three orders, and more preferably four or five orders of magnitude is expressly contemplated. As an example, purification from 0.1 % concentration to 10 % concentration is two orders of magnitude. To illustrate, individual cDNA clones isolated from a cDNA library have been conventionally purified to electrophoretic homogeneity. The sequences obtained from these clones could not be obtained directly either from the library or from total human DNA. The cDNA clones are not naturally occurring as such, but rather are obtained via manipulation of a partially purified naturally occurring substance (messenger RNA). The conversion of mRNA into a cDNA library involves the creation of a synthetic substance (cDNA) and pure individual cDNA clones can be isolated from the synthetic library by clonal selection. Thus, creating a cDNA library from messenger RNA and subsequently isolating individual clones from that library results in an approximately 10^4 - 10^6 fold purification of the native message.

The term "purified" is further used herein to describe a polynucleotide or polynucleotide of the invention which has been separated from other compounds including, but not limited to other polynucleotides or polypeptides (such as the enzymes used in the synthesis of the polynucleotide), carbohydrates, lipids, etc.,. The term "purified" may be used to specify the separation of monomeric polypeptides of the invention from oligomeric forms such as homo- or hetero- dimers, trimers, etc. The term "purified" may also be used to specify the separation of covalently closed polynucleotides from linear polynucleotides. A polynucleotide is substantially pure when at least about 50%, preferably 60 to 75% of a sample exhibits a single polynucleotide sequence and conformation (linear versus covalently close). A substantially pure polypeptide or polynucleotide